



Centro de Pesquisa em Alimentos

Bioeconomia e Alimentos: Uso de subprodutos ou resíduos na produção de novos materiais

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Introdução

Definição do Conselho Alemão de Bioeconomia, 2015:
“Bioeconomia é a produção e utilização de recursos biológicos - incluindo o conhecimento - para fornecer produtos, processos e serviços em todos os setores dentro da estrutura de uma economia sustentável”.
(von Braun, 2018).



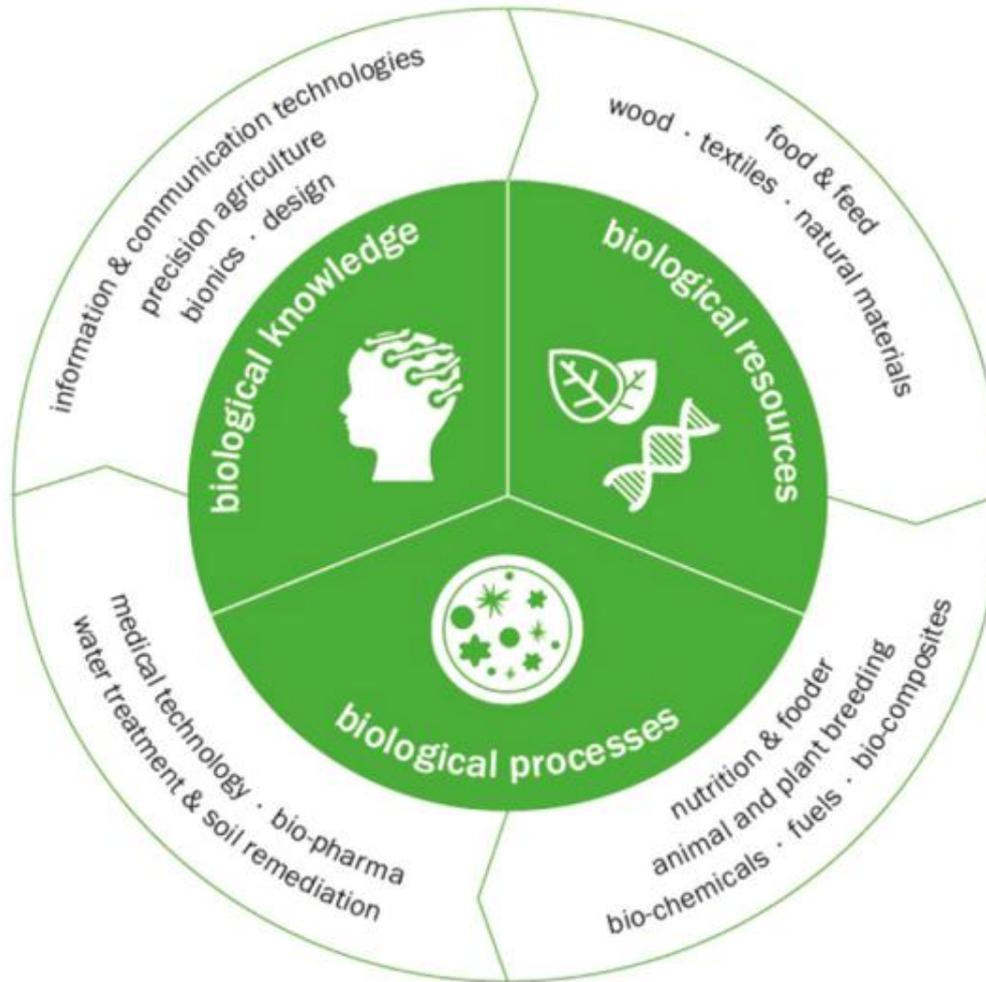


Fig. Bioeconomy Concept and Elements. Source: [German Bioeconomy Council \(2015\)](#). Fonte: von Braun et al. (2018)

A bioeconomia compreende as partes da economia que usam recursos biológicos renováveis da terra e do mar de forma responsável, visando beneficiar as empresas, a sociedade e a natureza (McCormic & Kautto, 2013), a produção de alimentos sendo um desses usos (Kristinsson & Jörundsdóttir, 2019).



Mais de 2 bilhões de euros em volume de negócios para a bioeconomia global na UE, que oferece mais de 18 milhões de empregos (Piotrowski et al., 2016).

Até 2030, o mundo precisa produzir 50% a mais de alimentos, 45% a mais de energia e 30% a mais de água (WWAP, 2012).

Em um mundo que se aproxima de 9 bilhões de pessoas em meados do século 21, a necessidade de uma transformação do sistema econômico e do estilo de vida torna-se cada vez mais importante em direção à sustentabilidade (von Braun, 2018).

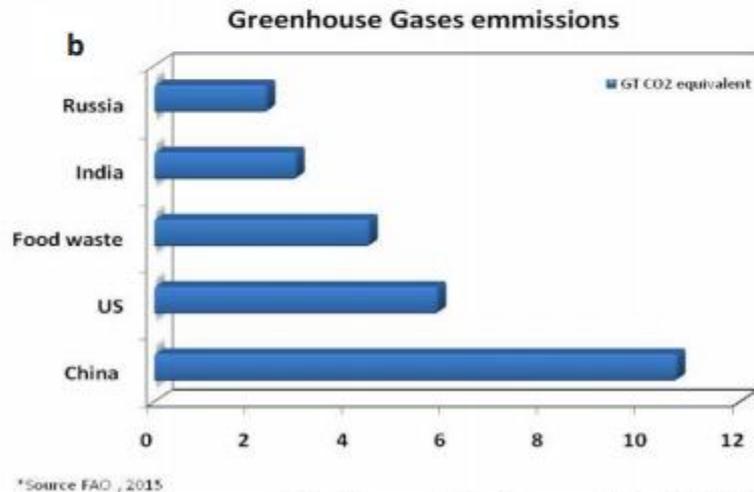
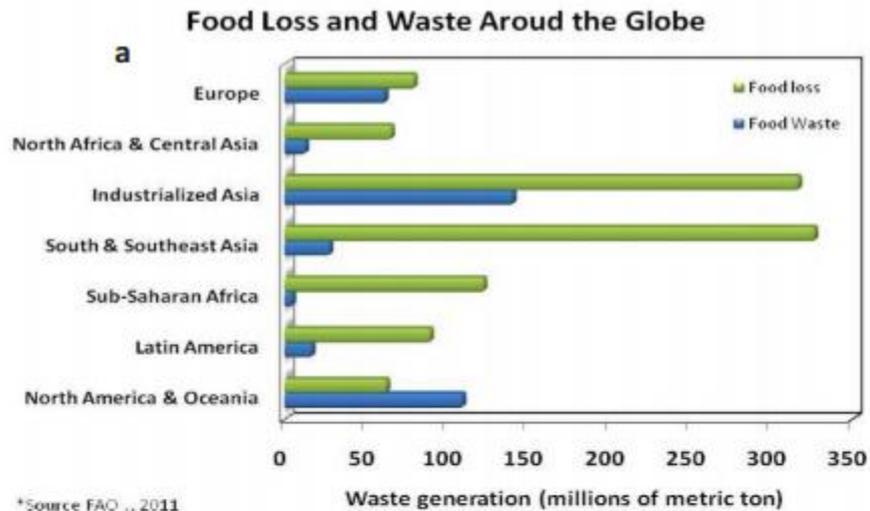


Nexus Água-Energia-Alimentos

Água, energia e alimentos são recursos essenciais e sistemas interdependentes. A necessidade de integração intersetorial da produção e provisão de recursos essenciais tem recebido maior atenção. Esta abordagem intersetorial é definida como o *Nexus Água-Energia-Alimentos* (Artioli et al., 2017).

A valorização de resíduos de alimentos é uma meta do desenvolvimento sustentável e ganha grande interesse, já que muitos produtos de base biológica podem ser derivados deles, além de energia e combustíveis (Rama Mohan, 2016).



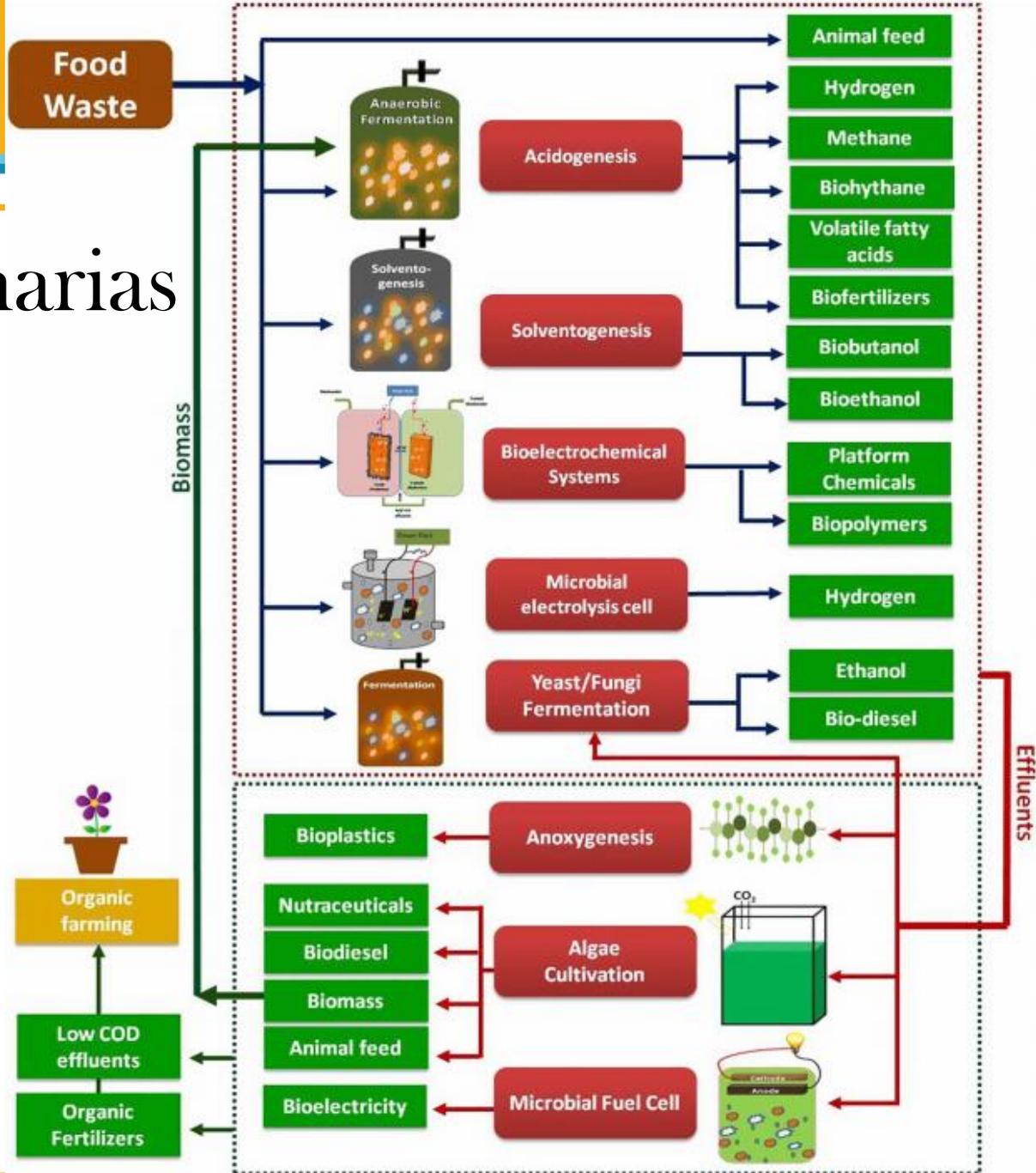


Fonte: Dahiya et al. (2018)

Fig. (a) Global food waste and food loss statistics (Source: FAO, 2011) (b) Global green house gases emissions in terms of GT CO₂ equivalents (Source: FAO, 2015)



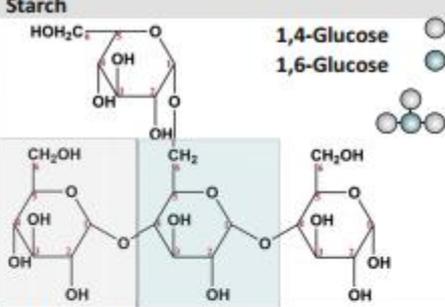
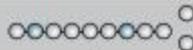
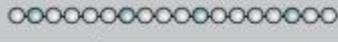
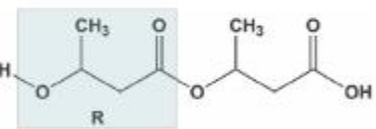
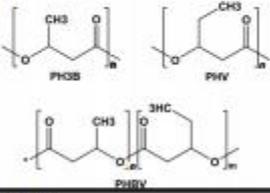
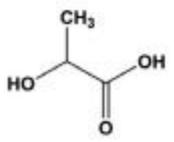
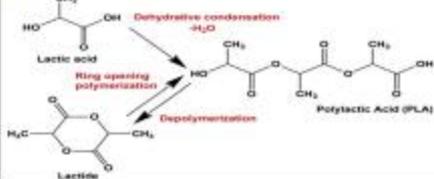
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Fonte:
Dahiya et al. (2018)

Fig. 5. Overview of multi-dimensional approaches for valorization of food waste to value added bio-based products in circular bioeconomy.

Figure Major Bioplastic Classes. Starch-based polymers, polyhydroxyalkanoates (PHAs) [including polyhydroxybutyrate (PHB), polyhydroxyvalerate (PHV), poly (3-hydroxybutyrate-co-3-hydroxyvalerate) (PHBV)], polylactic acid (PLA), cellulose-based polymers, polyethylene (PE), polyvinyl chloride (PVC), and protein-based polymers. Bioplastic input monomers and polymers (left-hand column), polymer production steps (middle column), properties, uses, and degradability characteristics (right-hand columns).

Natural monomer & polymer	Polymer processing	Property summary	
Starch 	Starch-based polymers Hydrolyzed Starch  Bioplastic polymer  Bioplastic plasticizer crosslinkers 	Properties ✓ Thermoplastic [20] ✓ Gas barrier [28] ✗ UV resistant [29] ✓ Biocompatible [30] ✗ Thermostable [31] ✓ Elastic [32] ✓ Rigid [32] ✗ Hydrophobic [35]	Uses Packaging [27] Food trays [27] Trash bags [27] Flower pots [27] Degradable ✓ In water [33] ✓ In soil [34] ✓ Ind. compost [36]
Polyhydroxyalkanoates 	PHA, PHB, PHV 	Properties ✓ Thermoplastic [37] ✓ Gas barrier [39] ✓ UV resistant [20] ✓ Biocompatible [37] ✗ Thermostable [37] ✓ Elastic [38] ✓ Rigid [40] ✓ Hydrophobic [37]	Uses Packaging [38] Adhesives [38] Fibers [38] Med. Implants [38] Degradable ✓ In water [33] ✓ In soil [36] ✓ Ind. compost [36]
Lactic acid 	Polylactic acid (PLA) 	Properties ✓ Thermoplastic [20] ✓ Gas barrier [42] ✓ UV resistant [43] ✓ Biocompatible [37] ✗ Thermostable [44] ✓ Elastic [43] ✓ Rigid [46] ✓ Hydrophobic [48]	Uses Packaging [41] Textiles [41] Med. Implants [41] Films [41] Degradable ✗ In water [45] ✓ In soil [47] ✓ Ind. compost [36]

Fonte: Karan et al. (2019)

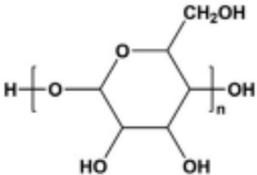
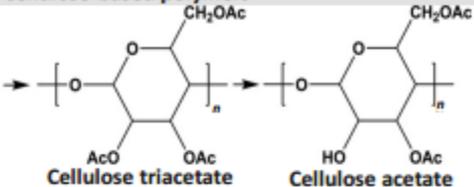
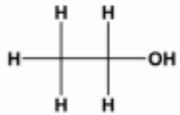
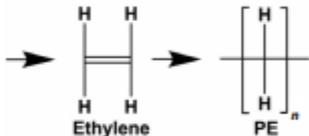
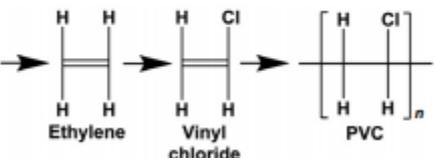
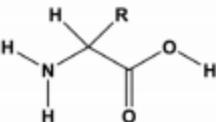
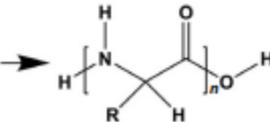
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FACULDADE DE ZOOTECNIA E ENGENHARIA DE ALIMENTOS

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Figure Major Bioplastic Classes. Starch-based polymers, polyhydroxyalkanoates (PHAs) [including polyhydroxybutyrate (PHB), polyhydroxyvalerate (PHV), poly (3-hydroxybutyrate-co-3-hydroxyvalerate) (PHBV)], polylactic acid (PLA), cellulose-based polymers, polyethylene (PE), polyvinyl chloride (PVC), and protein-based polymers. Bioplastic input monomers and polymers (left-hand column), polymer production steps (middle column), properties, uses, and degradability characteristics (right-hand columns).

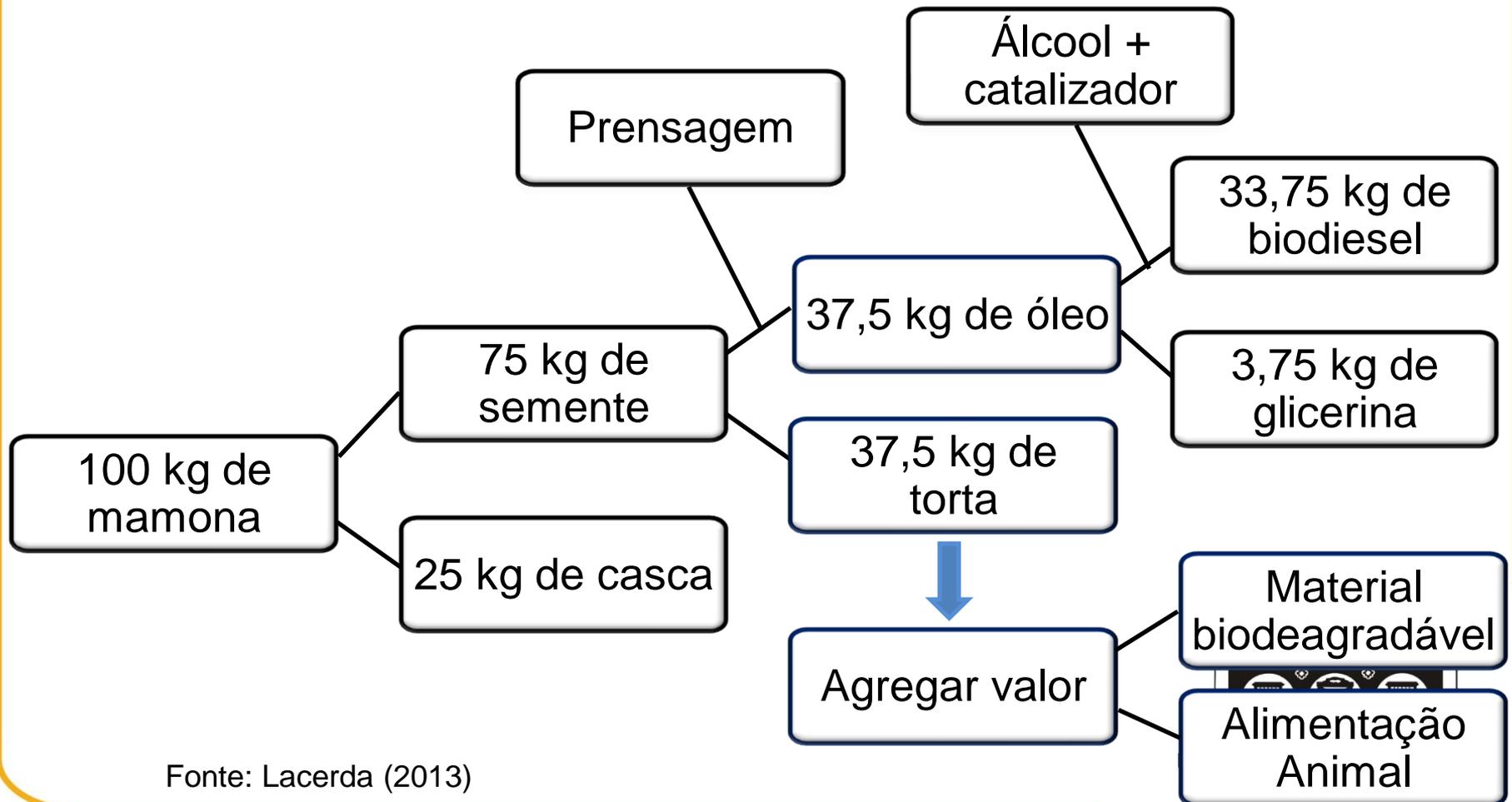
<p>Cellulose</p> 	<p>Cellulose-based polymers</p>  <p>Cellulose triacetate Cellulose acetate</p>	<p>Properties</p> <ul style="list-style-type: none"> ✗ Thermoplastic [31] ✗ Gas barrier [49] ✗ UV resistant^{viii} ✓ Biocompatible [50] ✓ Thermostable [51] ✗ Elastic [52] ✓ Rigid [54] ✗ Hydrophobic [54] 	<p>Uses</p> <ul style="list-style-type: none"> Wound dress.^{vii} Textiles^{vii} Air filters^{vii} Coatings^{vii} <p>Degradable</p> <ul style="list-style-type: none"> ✓ In water [53] ✓ In soil [55] ✓ Ind. compost [36]
<p>Ethanol</p> 	<p>Polyethylene</p>  <p>Ethylene PE</p>	<p>Properties</p> <ul style="list-style-type: none"> ✓ Thermoplastic [17] ✓ Gas barrier [57] ✗ UV resistant [58] ✓ Biocompatible^{ix} ✗ Thermostable^x ✗ Elastic^{xi} ✓ Rigid^{xii} ✓ Hydrophobic^{xiii} 	<p>Uses</p> <ul style="list-style-type: none"> Bottles [56] Ship container [56] Container lids [56] Adhesives [56] <p>Degradable</p> <ul style="list-style-type: none"> ✗ In water [11] ✗ In soil [34] ✗ Ind. compost [34]
<p>Ethanol</p> 	<p>Polyvinyl chloride</p>  <p>Ethylene Vinyl chloride PVC</p>	<p>Properties</p> <ul style="list-style-type: none"> ✓ Thermoplastic [17] ✗ Gas barrier [59] ✗ UV resistant^{xiv} ✓ Biocompatible^x ✗ Thermostable [60] ✓ Elastic^{xv} ✓ Rigid^{xiv} ✓ Hydrophobic [62] 	<p>Uses</p> <ul style="list-style-type: none"> Packaging [56] Window frames [56] Railings [56] Pipes [56] <p>Degradable</p> <ul style="list-style-type: none"> ✗ In water^{xvi} ✗ In soil [61] ✗ Ind. compost [17]
<p>Amino acid</p> 	<p>Protein-based polymers</p> 	<p>Properties</p> <ul style="list-style-type: none"> ✓ Thermoplastic [63] ✗ Gas barrier [65] ✓ UV resistant [66] ✓ Biocompatible [67] ✓ Thermostable [68] ✓ Elastic [65] ✓ Rigid [70] ✓ Hydrophobic [71] 	<p>Uses</p> <ul style="list-style-type: none"> Cast film [64] Injection mold. [64] Compr. mold. [64] Extrud. sheets [64] <p>Degradable</p> <ul style="list-style-type: none"> ✓ In water [69] ✓ In soil [69] ✓ Ind. compost [69]

Fonte: Karan et al. (2019)



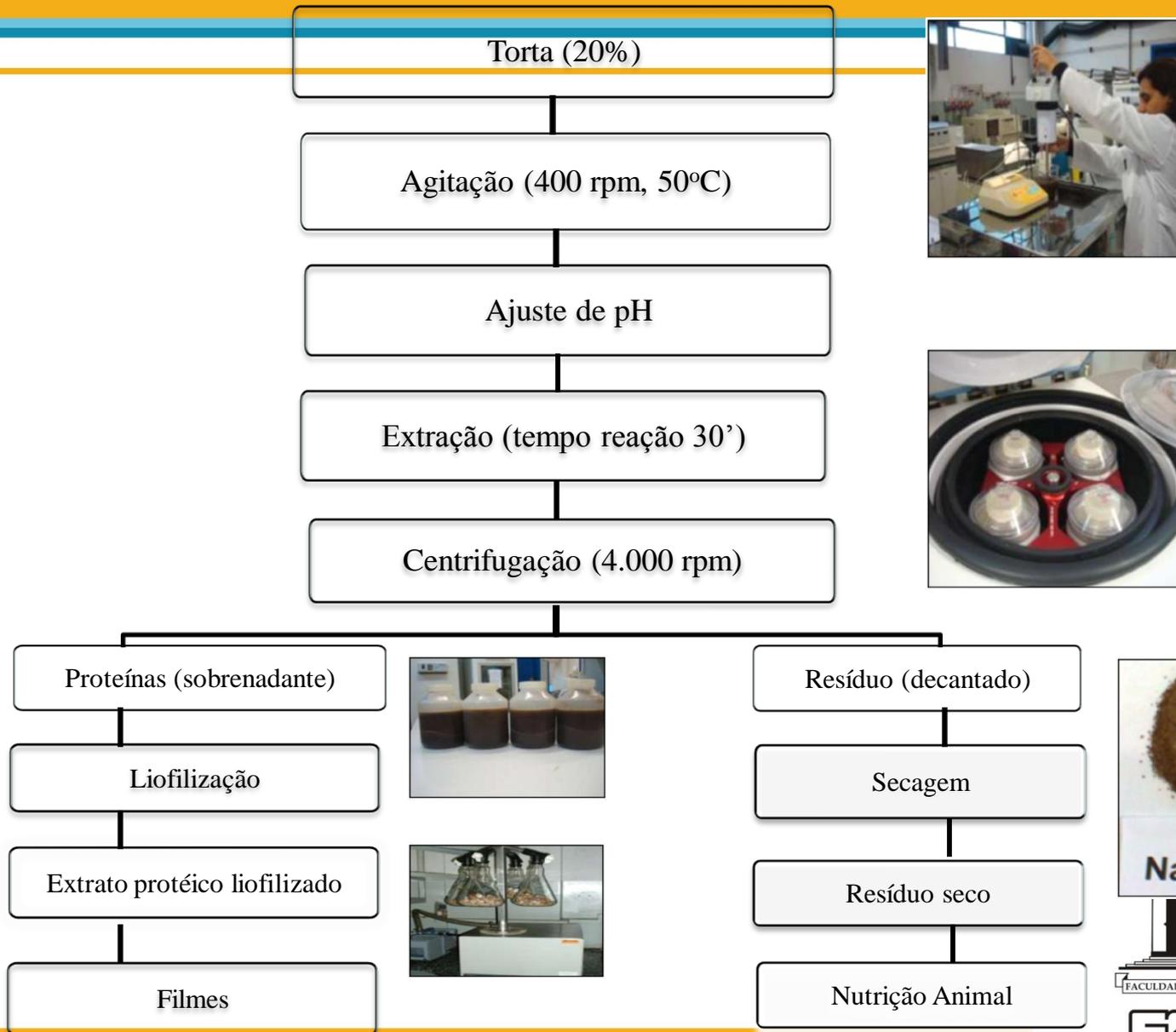
Alguns Resultados – biopolímeros

Cadeia do biodiesel – torta de mamona

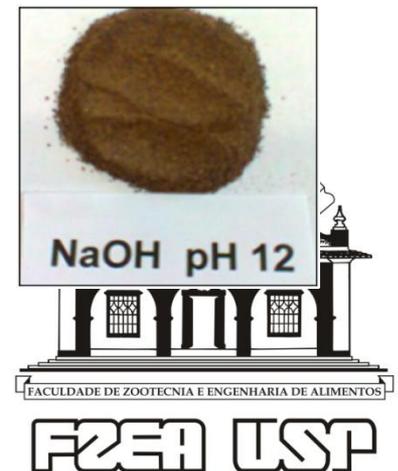


Fonte: Lacerda (2013)

Fluxograma de extração de proteína



Fonte: Lacerda (2013)



- Filmes a base de proteínas extraídas da torta da mamona



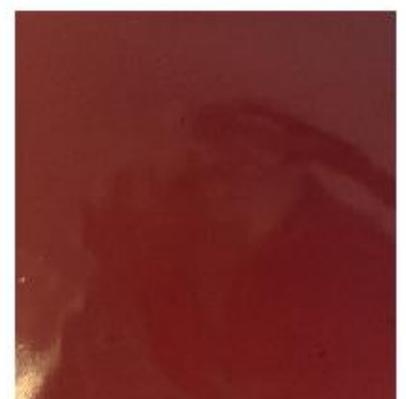
F1



F2



F3



F4

F1=filme com fibra e com glicol; F2=filme com fibra e sem glicol;
F3=filme sem fibra e com glicol; F4=filme sem fibra e sem glicol.

Fonte:
Bittante (2015)

pH=10



pH=11



pH=12



Composição centesimal dos dados de matéria seca, proteína e cinzas do resíduo

Resíduo Seco

	pH8	pH9	pH10	pH11
Matéria seca (%)	91,44 ± 1,01 ^a	93,89± 1,30 ^a	91,58±1,08 ^a	91,44±2,65^a
Proteína (b.s.%)	36,26 ± 1,08 ^a	33,40± 0,44 ^b	30,79±1,09 ^c	25,84±0,42^d
Cinzas (b.s.%)	12,60 ± 0,54 ^{ab}	11,91 ± 0,25 ^b	13,40±0,35 ^a	13,55±0,18^a

Fonte: Lacerda (2013)

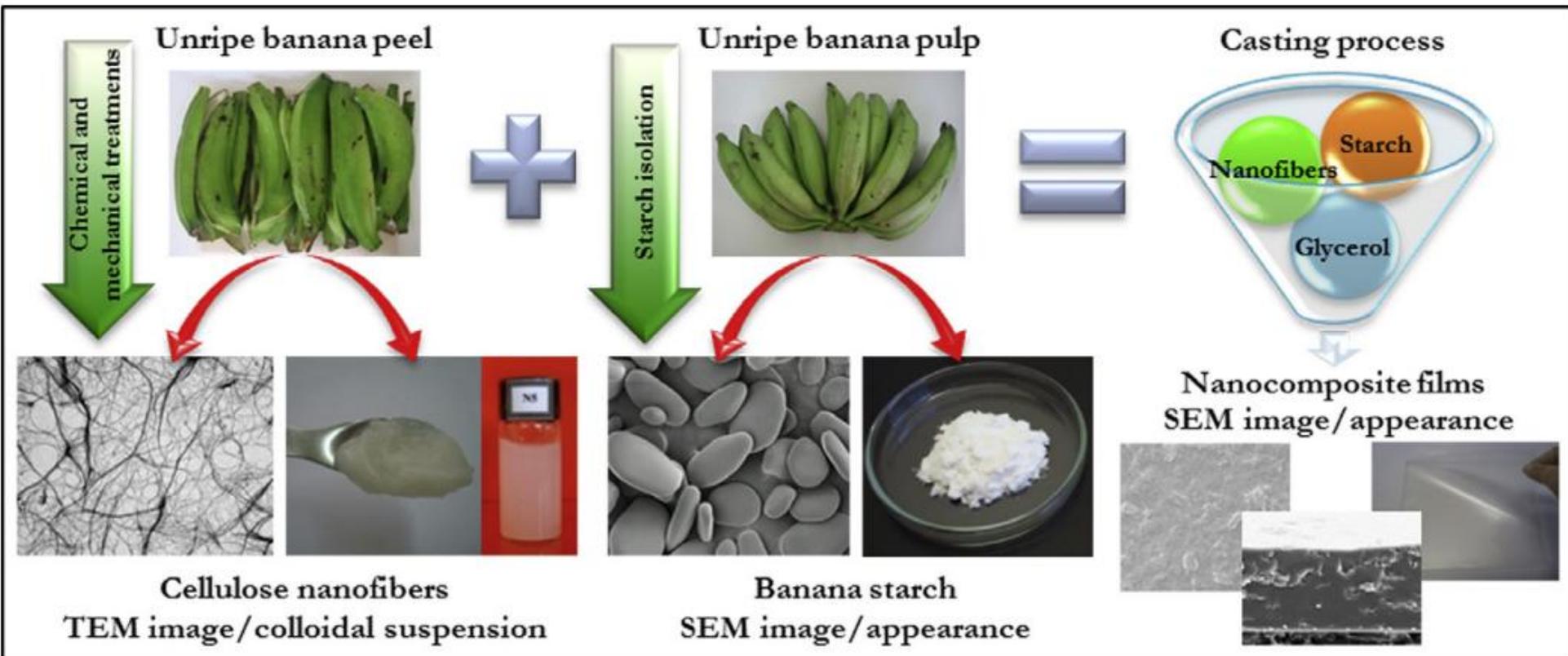
Perfil de aminoácidos dos resíduos de extração das proteínas em NaOH em diferentes pH

Aminoácidos (g/100g proteína)	pH de extração		
	10	11	12
Alanina	3,61±0,02 ^a	3,86±0,02 ^a	3,14±0,07 ^b
Arginina	9,57 ±0,05 ^a	9,89±0,04 ^a	7,67±0,01 ^b
Acido aspártico	7,16±0,25 ^a	7,45±0,21 ^a	5,98±0,30 ^a
Glicina	3,58±0,03 ^a	3,76±0,02 ^a	3,23±0,06 ^b
Isoleucina	2,93±0,17 ^a	3,26±0,06 ^a	2,56±0,04 ^a
Leucina	5,19±0,11 ^a	5,57±0,10 ^a	4,54±0,05 ^b
Acido glutâmico	13,44±0,06 ^a	13,39±0,01 ^a	11,63±0,26 ^b
Lisina	2,61±0,11 ^a	2,47±0,09 ^a	1,54±0,01 ^b
Cistina	1,82±0,22 ^a	0,89±0,09 ^a	0,19±0,00 ^a
Metionina	1,73±0,13 ^a	1,98±0,03 ^a	1,69±0,02 ^a
Fenilalanina	3,11±0,07 ^a	3,33±0,05 ^a	2,65±0,04 ^b
Tirosina	2,05±0,71 ^a	2,21±0,02 ^a	1,79±0,05 ^b
Treonina	2,70±0,76 ^a	3,03±0,01 ^a	1,83±0,03 ^b
Triptofano	7,89±0,04 ^a	7,75±0,06 ^a	6,23±0,13 ^b
Prolina	3,14±0,03 ^a	3,53±0,04 ^a	2,61±0,03 ^b
Valina	3,37±0,20 ^a	3,79±0,13 ^a	2,04±0,04 ^b
Histidina	2,14±0,08 ^a	2,27±0,07 ^{ba}	1,52±0,11 ^b
Serina	4,49±0,03 ^a	4,71±0,01 ^a	

Fonte: Lacerda (2013)



Cadeia da banana – nanofibras de celulose



Fonte: Pelissari et al., (2017)

Composição da casca da banana:

Umidade: 5.2 %

Cinzas: 9.8 %

Celulose: 12.1 %

Hemicelulose: 10.2 %

Lignina: 2.9 %.

Fonte: Pelissari et al., (2014)

Tratamento mecânico: homogeneizador de duplo-estágio (GEA Niro Soavi modelo NS 1001L—Panda 2 K, Parma Italy): 500 e 50 Bar.

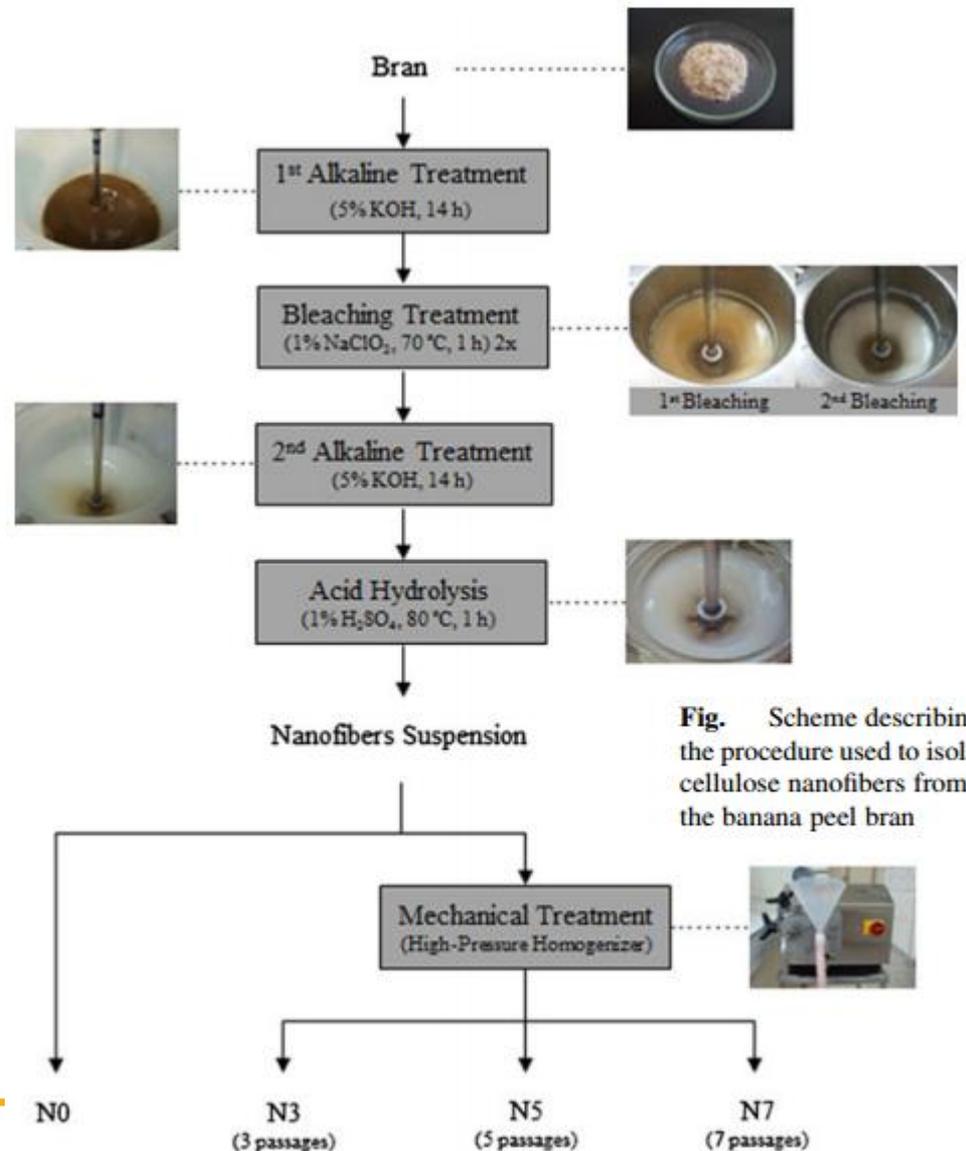


Fig. Scheme describing the procedure used to isolate cellulose nanofibers from the banana peel bran

Fonte: Pelissari et al., (2014)

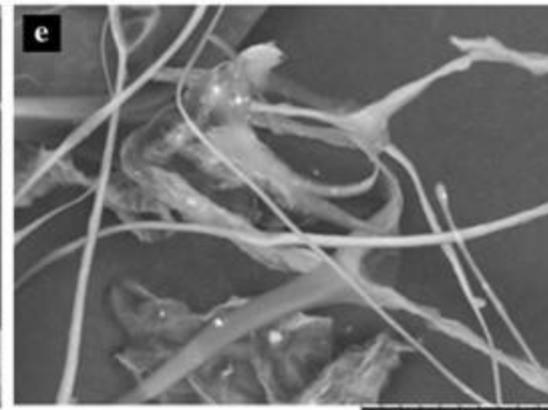
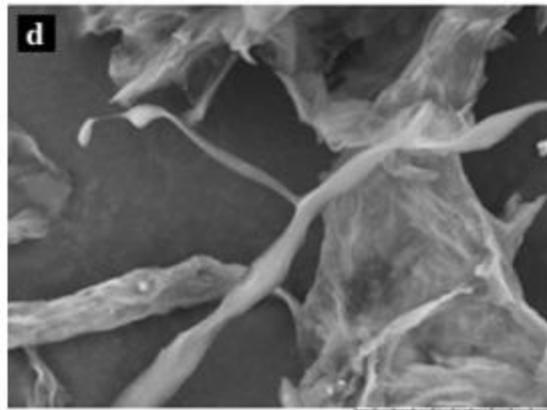
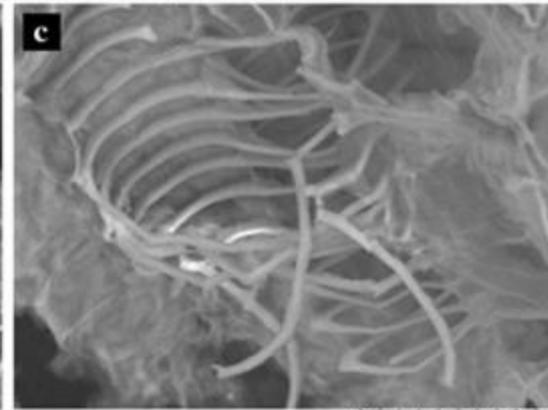
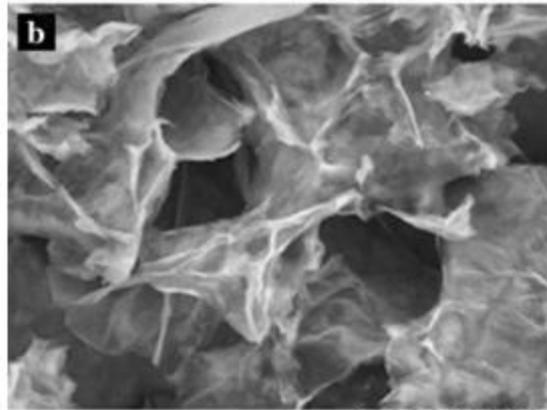
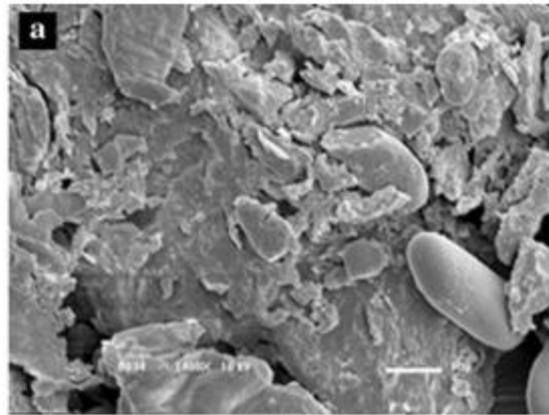


Fig. SEM images of the **a** banana peel bran ($\times 1,000$, scale bar 10 μm) and of the steps involved in the chemical treatment for the isolation of cellulose nanofibers: **b** first alkaline

treatment, **c** first bleaching, **d** second bleaching, and **e** second alkaline treatment ($\times 2,000$, scale bar 30 μm)

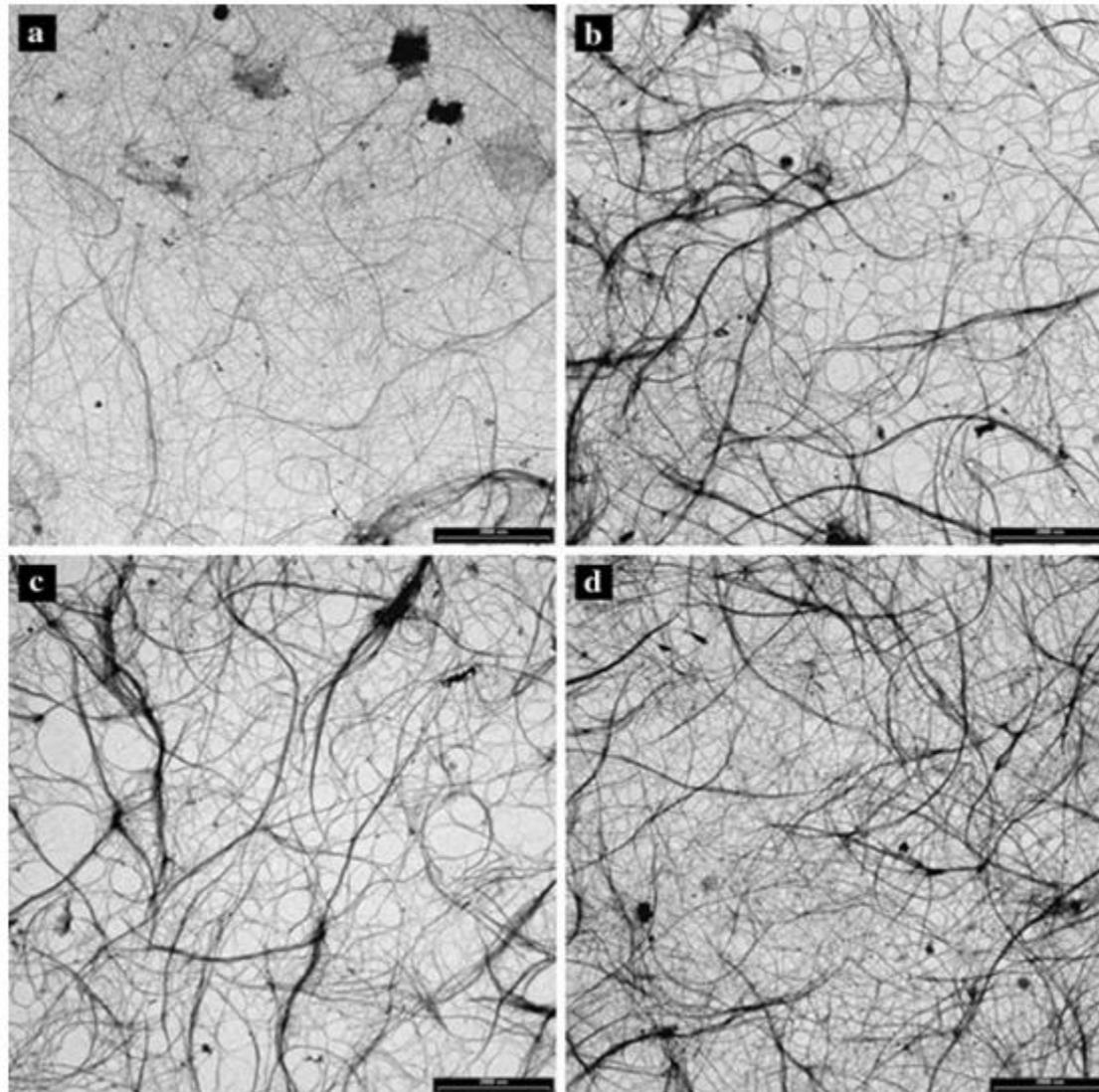
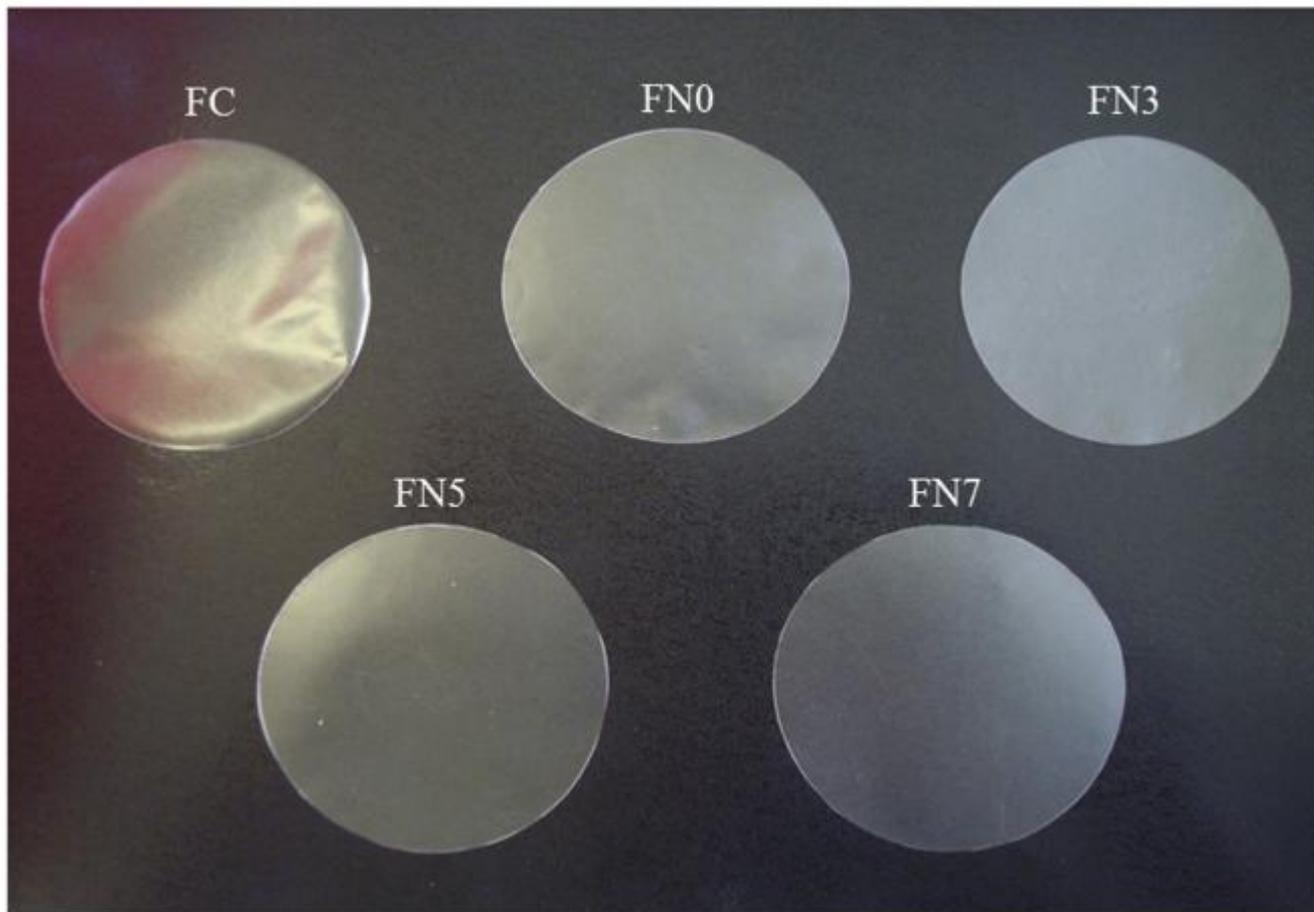


Fig. TEM images of cellulose nanofibers obtained by different number of passages in the high-pressure homogenizer: **a** N0, **b** N3, **c** N5, and **d** N7 ($\times 1,400$, scale bar 2,000 nm)

Fonte: Pelissari et al., (2014)



Amido extraído da polpa da banana da terra de acordo com Pelissari et al. (2013).

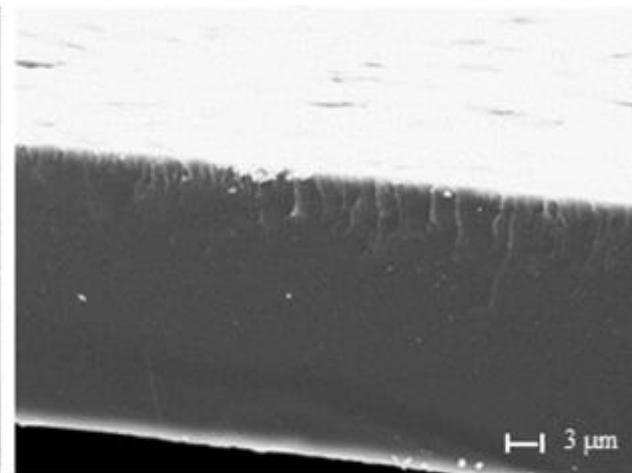
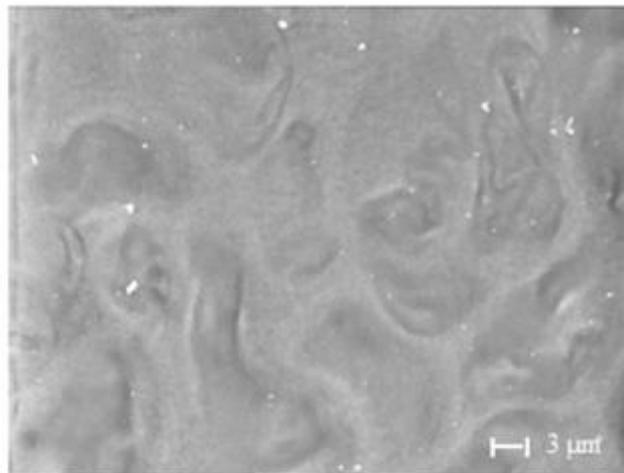
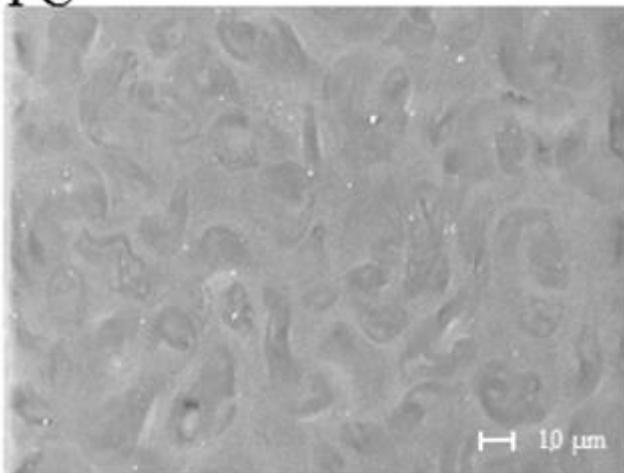
Filmes nanocompósitos:

- 4 g amido/100 g solução
- 5 g nanofibras/100 g amido
- 25 g glicerol/100 g amido

Fig. Appearance of the control film (FC) and nanocomposites reinforced with cellulose nanofibers that were passed through the high-pressure homogenizer zero (FN0), three (FN3), five (FN5), and seven (FN7) times.

Fonte: Pelissari et al., (2017)

FC



FN7

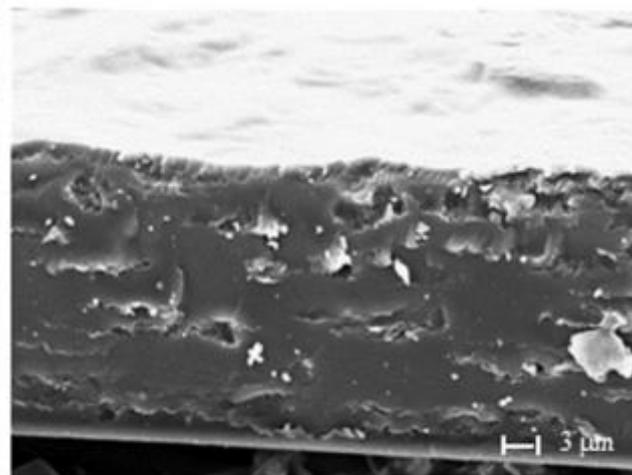
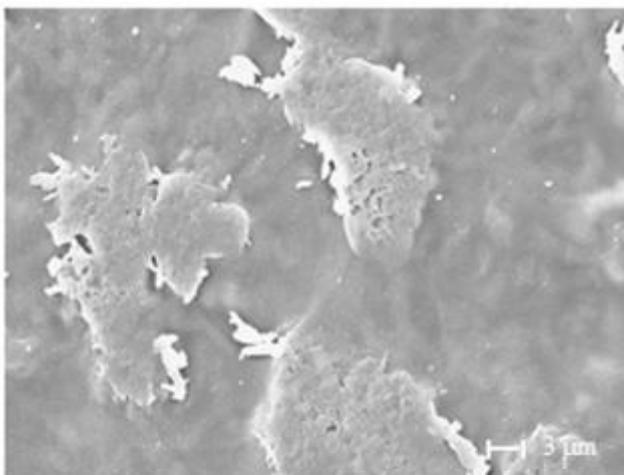
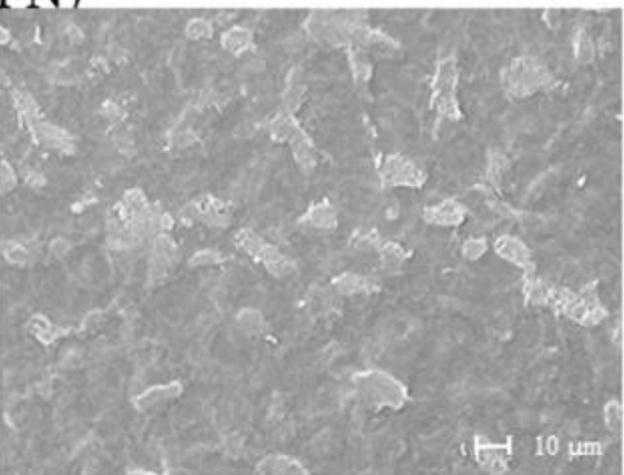


Fig. 1. SEM surfaces (500x, scale bar = 10 μm ; 2000x, scale bar = 3 μm) and cross-sections (2000x, scale bar = 3 μm) of the control film (FC) and nanocomposites reinforced with cellulose nanofibers that were passed through the high-pressure homogenizer zero (FN0), three (FN3), five (FN5), and seven (FN7) times.

Fonte: Pelissari et al., (2017)

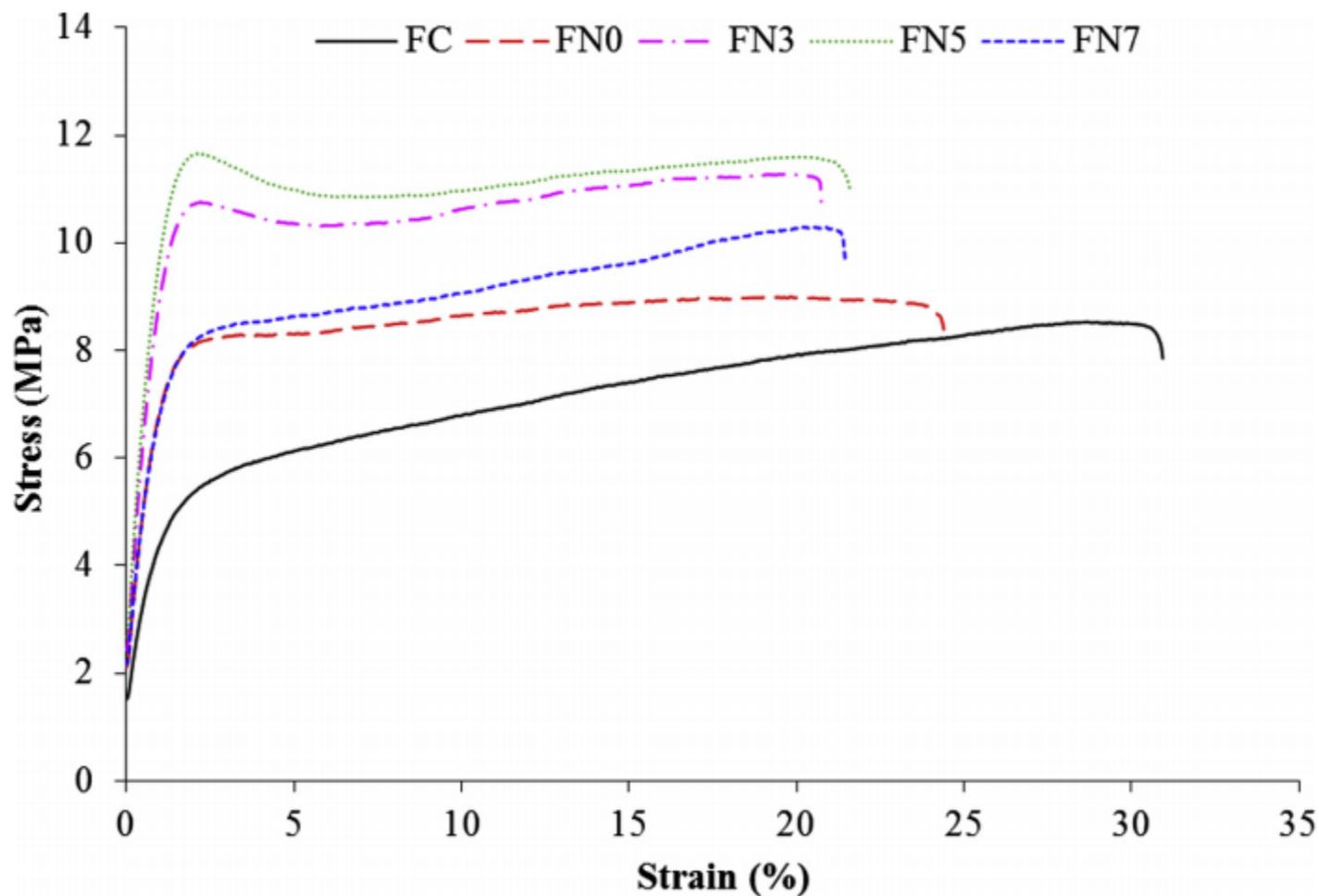


Fig. 3. Typical stress-strain curves obtained for the control film (FC) and nanocomposites reinforced with cellulose nanofibers that were passed through the high-pressure homogenizer zero (FN0), three (FN3), five (FN5), and seven (FN7) times.

Cadeia de ervas – extratos em filmes ativos



Componentes determinados por CG-MS (GC-2010, Shimadzu, Suzhou, China):

Cafeína.



1,8-cineol, p-cymeno, fitol, ácido eicosanoico, cis,cis,cis-7,10,13-hexadeca trienal, e α -ironeo.



Cinamaldeído e α -copaeno.



Eucalyptol, linalol, cânfora, α -pineno, terpineno, 4-ol, cânfeno, β -pineno, D-limoneno, p-cymeno, ácido n-hexadecanoico, endo-borneol, L- α -terpineol, caryofileno, bornyl acetato and tetradecanal.



Table: Results obtained for the extracts in the Trolox equivalent antioxidant capacity (TEAC) and DPPH assays at specific time points and for the Folin-Ciocalteu assay (Mean values with the standard deviations in brackets).

Sample		Rosemary	Boldo	Cinnamon	Guarana
TEAC (mM)	1 min	2.96 (0.04) ^{a,x}	5.50 (0.01) ^{b,x}	2.53 (0.11) ^{c,x}	0.25 (0.01) ^{d,x}
	3 min	3.32 (0.24) ^{a,y}	6.23 (0.21) ^{b,y}	2.75 (0.07) ^{c,y}	0.26 (0.01) ^{d,y}
	6 min	3.69 (0.42) ^{a,z}	6.66 (0.17) ^{b,z}	2.86 (0.06) ^{c,z}	0.26 (0.01) ^{d,z}
DPPH (mg/l)	15 min	0.37 (0.02) ^{a,x}	0.23 (0.02) ^{b,x}	0.06 (0.01) ^{a,x}	0.10 (0.01) ^{c,x}
	30 min	0.48 (0.02) ^{a,y}	0.37 (0.03) ^{b,y}	0.13 (0.04) ^{a,y}	0.14 (0.01) ^{c,y}
	45 min	0.80 (0.14) ^{a,z}	0.56 (0.05) ^{b,z}	0.23 (0.02) ^{c,z}	0.26 (0.04) ^{c,z}
Folin-Ciocalteu (mg GA/g extract)	---	115.00 (0.01) ^a	108.00 (0.04) ^a	172.00 (0.03) ^a	139.00 (0.03) ^a

The same superscript (abc) compares different extracts horizontally and (xyz) compare them vertically at different times in the LSD test. Values with different letters differ significantly in same row. Level of significance: 95.0%.



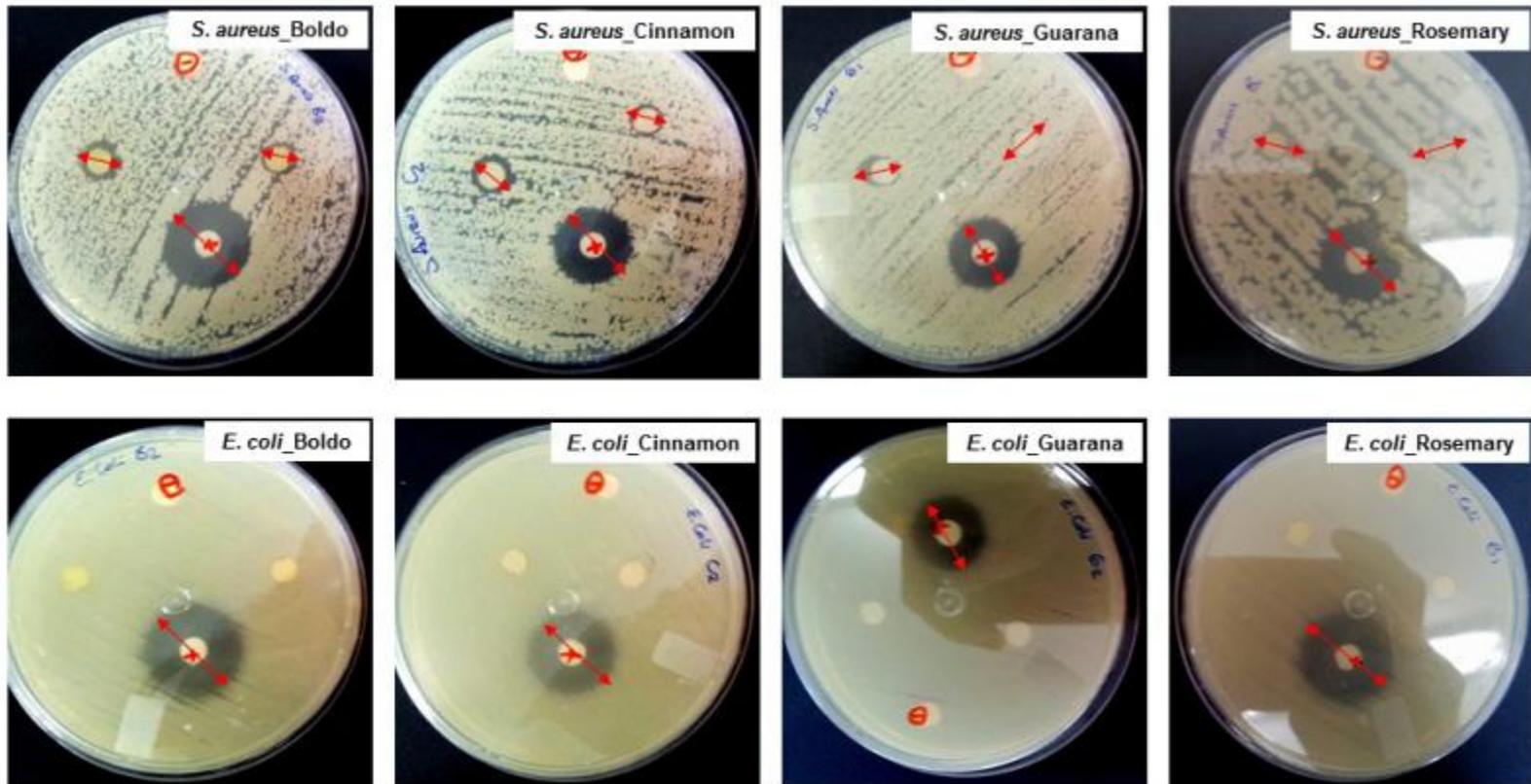
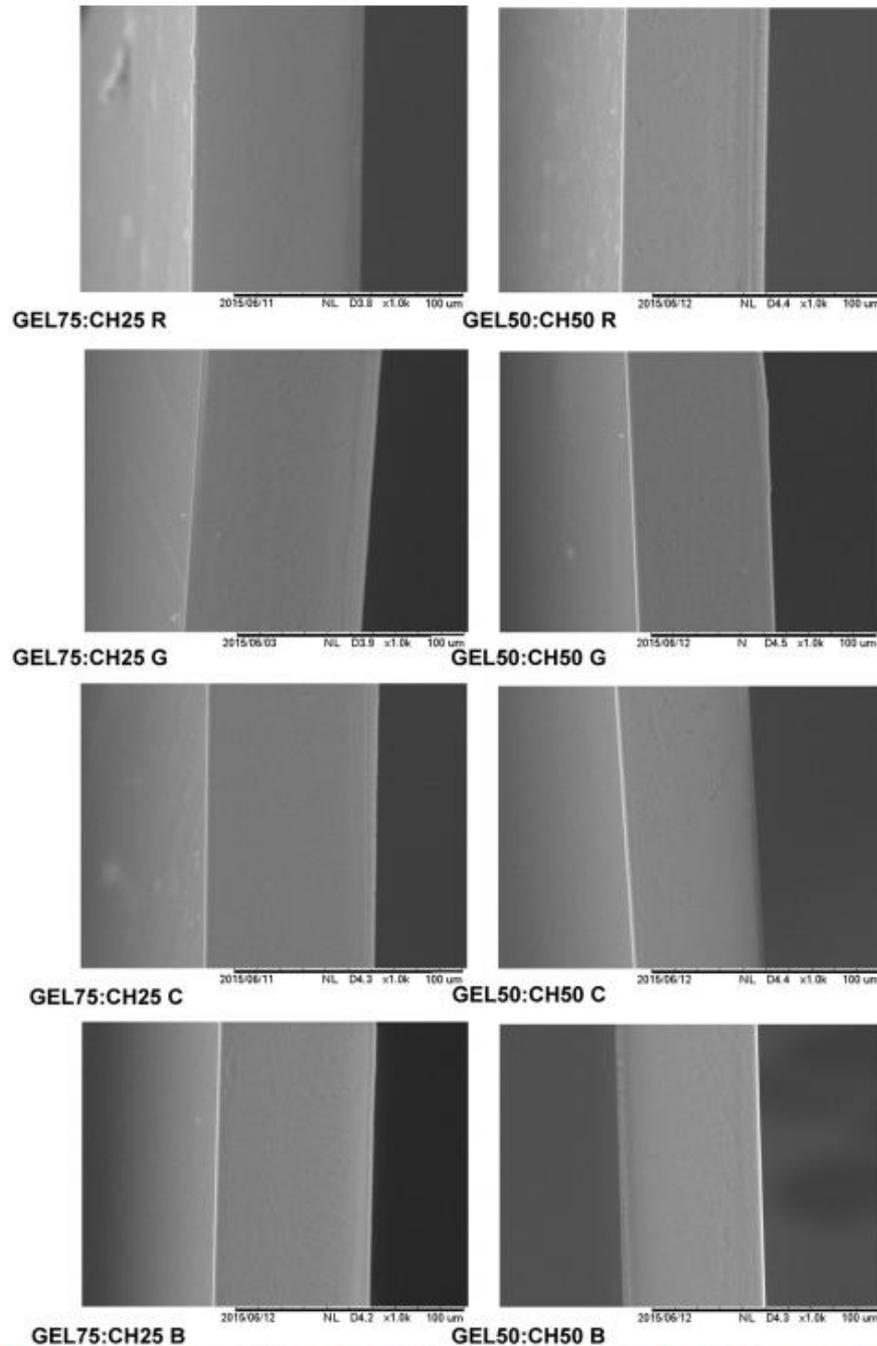


Figure . Inhibition of bacterial growth (*E. Coli* and *S. Aureus*) by the boldo of Chile (*Peumus boldus molina*), cinnamon (*Cinnamomum sp.*), guarana (*Paullinia cupana*) and rosemary (*Rosmarinus officinalis*) extracts.



Fonte: Bonilla & Sobral (2016)



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Fig. Scanning electron microscopy (SEM) images of cross-sections of blend films (Gel75:CH25 or GEL50:CH50) with guarana (G), boldo (B), cinnamon (C) and rosemary (R) extracts.

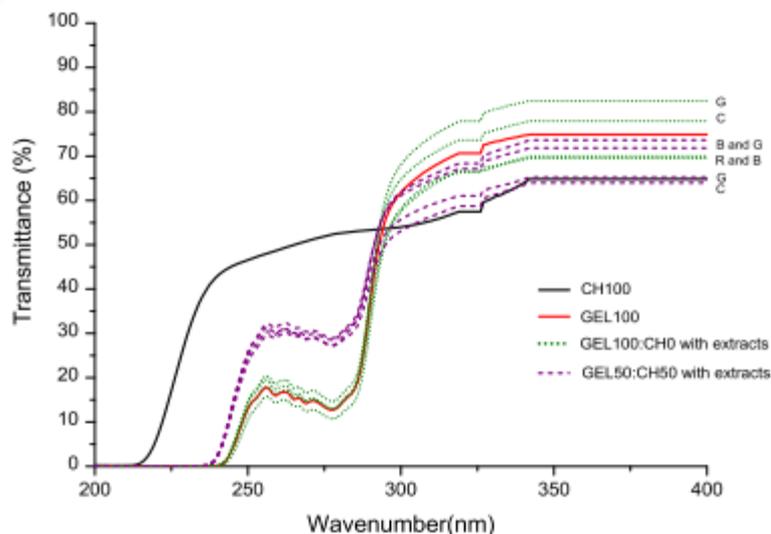


Fig. UV-Vis spectra for pure chitosan films (CH100), pure gelatin films (GEL100) and GEL100:CH0 and GEL50:CH50 blend films with guarana (G), boldo (B), cinnamon (C) and rosemary (R) extracts.

Fonte: Bonilla & Sobral (2016)

Table

Trolox equivalent antioxidant capacity (TEAC) of the films (mg/l) with or without ethanolic extract at specific time points.

Film	Extract	1 min	3 min	6 min	SEM
GEL100	Without	—	—	—	—
CH100	Without	0.15 ^{C,d}	0.16 ^{C,d}	0.16 ^{C,d}	0.13
GEL 75:CH25	Boldo	0.94 ^{B,a}	1.4 ^{B,a}	1.5 ^{B,a}	0.13
GEL 50:CH50	Boldo	1.1 ^{A,a}	1.5 ^{A,a}	1.8 ^{A,a}	0.13
GEL 75:CH25	Rosemary	0.17 ^{B,b}	0.21 ^{B,b}	0.27 ^{B,b}	0.13
GEL 50:CH50	Rosemary	1.2 ^{A,b}	1.6 ^{A,b}	1.9 ^{A,b}	0.13
GEL 75:CH25	Cinnamon	0.17 ^{B,c}	0.23 ^{B,c}	0.31 ^{B,c}	0.13
GEL 50:CH50	Cinnamon	0.27 ^{A,c}	0.31 ^{A,c}	0.32 ^{A,c}	0.13
GEL 75:CH25	Guarana	0.89 ^{B,ab}	1.2 ^{B,ab}	1.5 ^{B,ab}	0.13
GEL 50:CH50	Guarana	0.93 ^{A,ab}	1.3 ^{A,ab}	1.6 ^{A,ab}	0.13

— Not showed antioxidant capacity. GEL: Gelatin; CH: Chitosan. Means with the same superscript (^{ABCD}) were compared vertically for different GEL:CH ratios, and means (^{abcd}) were compared vertically for different extracts in LSD test. SEM: Standard error of the mean based on Mean Square Error of ANOVA analysis.



Cadeia de ervas – extratos como aditivos

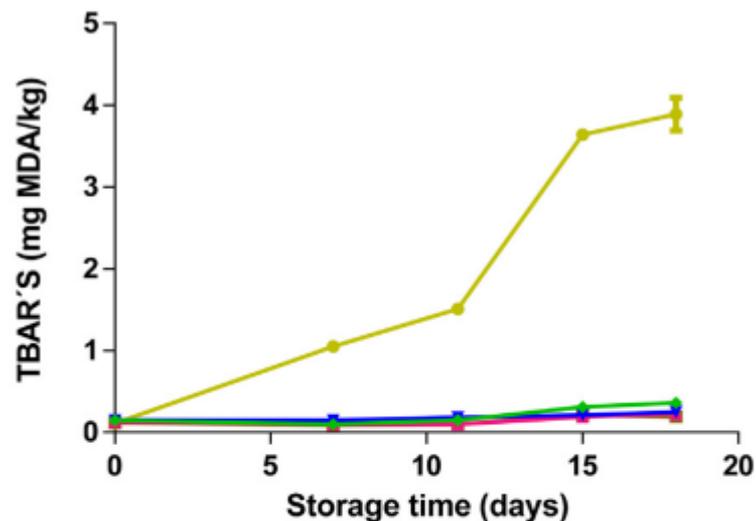
Extrato de folhas de pitanga (EFP) foi estudado como aditivo antioxidante e antimicrobiano em “burgers” de carne suína embalada em atmosfera modificada e estocada a 2 °C.

Cinco tratamentos:

- i) Sem antioxidante.
- ii) Com BHT (200 mg/kg).
- iii) Com EFP (250, 500 e 1000 mg/kg).

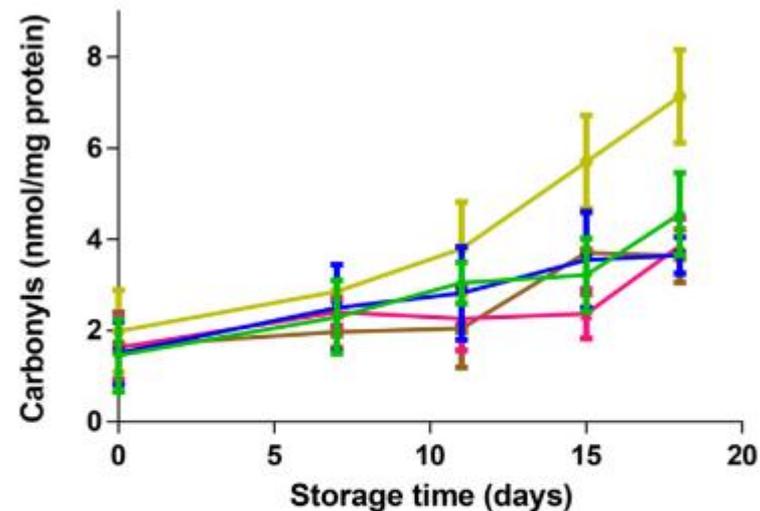
Foram identificados (UHPLC-ESI/QTOF): ácido hidroxicinâmico, tirosol e outros fenólicos (acilmetoxifenóis, hidroxicomarinos e hidroxifenilpropenos).





—●— Control —■— BHT —◆— PLL —▲— PLM —▼— PLH

Fig. Effect of addition of antioxidant on the evolution of TBARS values of pork burgers packed under modified atmosphere packaging (80% O₂ + 20% CO₂) during storage at 2 ± 1 °C. Treatments: CON: control, without antioxidant; BHT: Butylated hydroxytoluene at 200 mg/kg; PLL pitanga leaf extracts at 250 mg/kg; PLM: pitanga leaf extracts at 500 mg/kg; PLH: pitanga leaf extracts at 1000 mg/kg;



—●— Control —■— BHT —◆— PLL —▲— PLM —▼— PLH

Fig. Effect of addition of antioxidant on the evolution of carbonyl of pork burgers packed under modified atmosphere packaging (80% O₂ + 20% CO₂) during storage at 2 ± 1 °C. Treatments: CON: control, without antioxidant; BHT: Butylated hydroxytoluene at 200 mg/kg; PLL pitanga leaf extracts at 250 mg/kg; PLM: pitanga leaf extracts at 500 mg/kg; PLH: pitanga leaf extracts at 1000 mg/kg;

Fonte: Lorenzo et al. (2018)



Considerações finais

- Bioeconomia e biorefinarias envolvem produção e processamento de Alimentos.
- Alimentos seguros e a segurança alimentar devem estar nesse contexto.
- Estudos mais sistêmicos são necessários.
- Custos?
- Água, Energia e Alimentos não são inesgotáveis!



Muito obrigado pela atenção!!!

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