

Aplicação de técnica de metabolômica ao estudo de alimentos

Edy Sousa de Brito



FORMAÇÃO ACADÊMICA

- 1998: Bacharel em Química Industrial pela UFPB**
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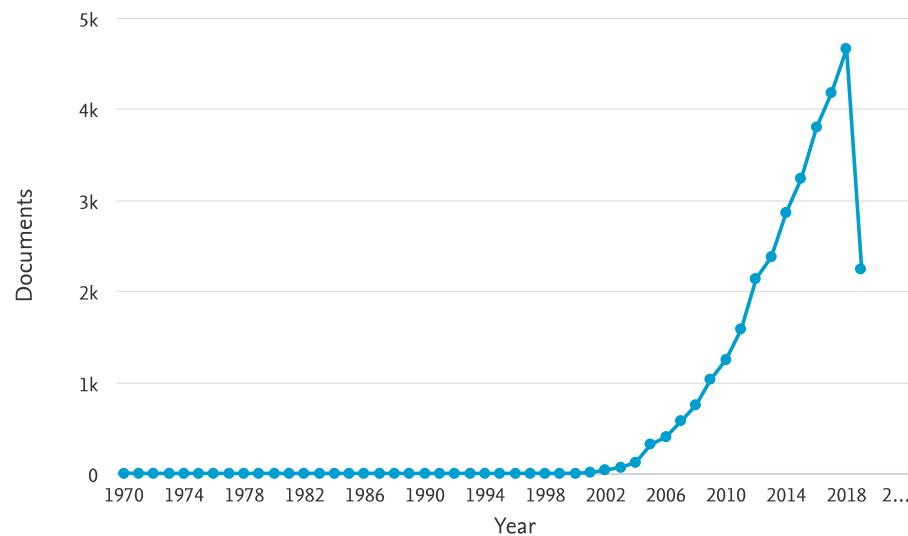
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metabolomic

Scopus junho 2019

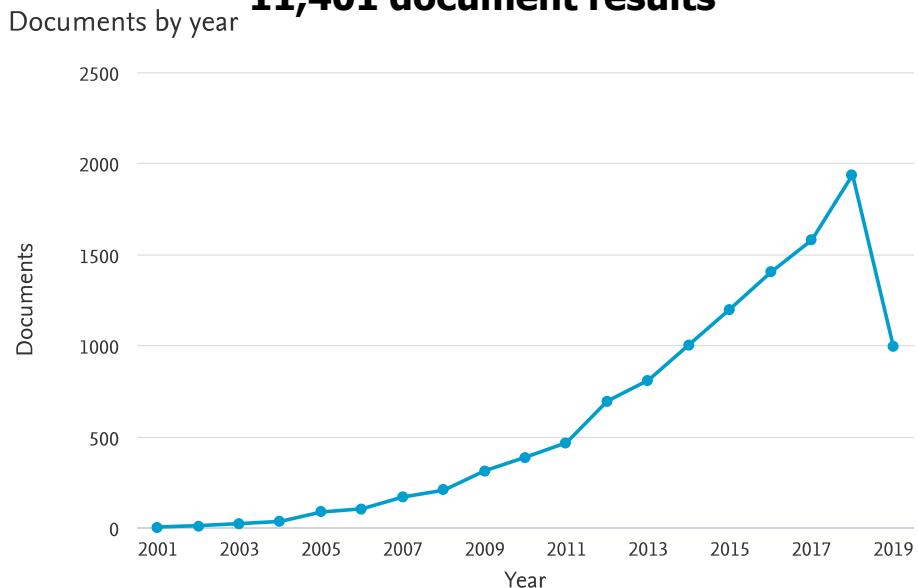
31,643 document results

Documents by year



metabolomic AND food

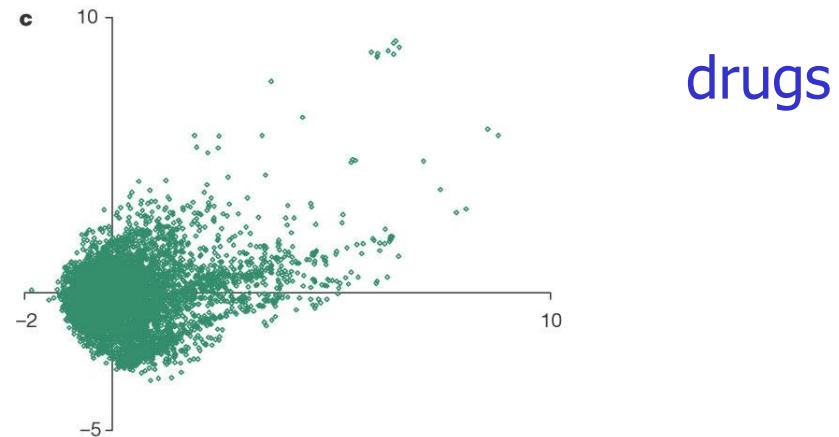
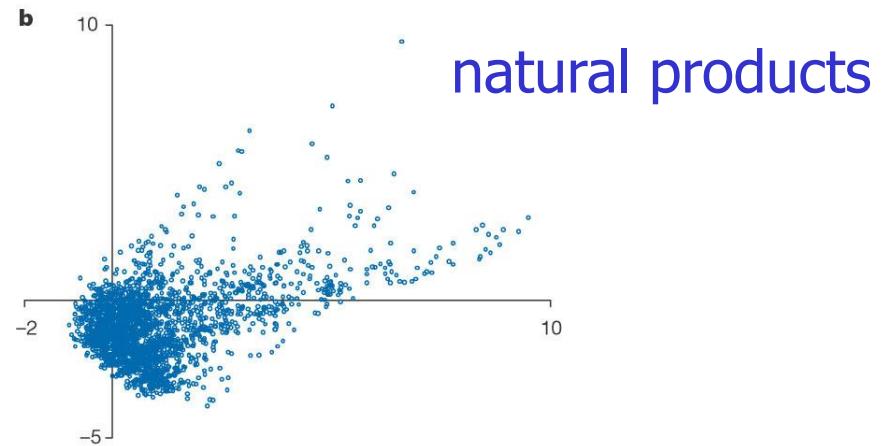
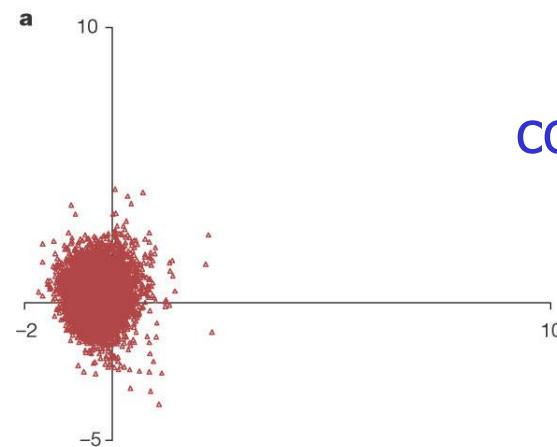
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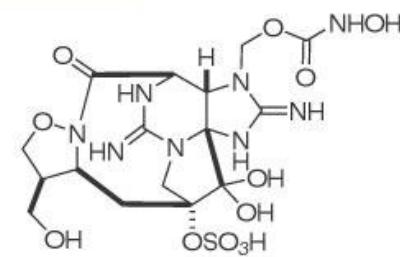
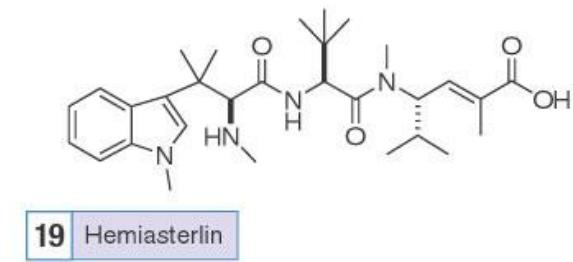
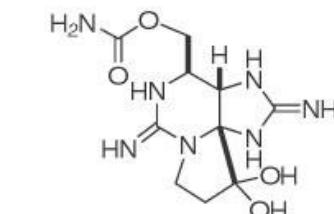
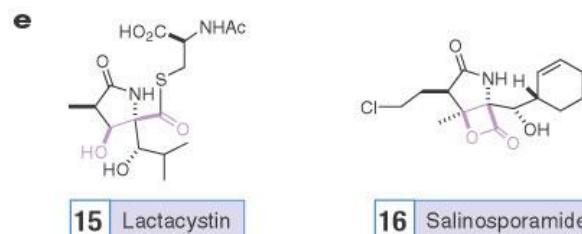
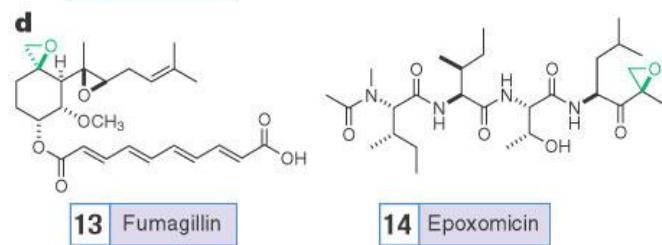
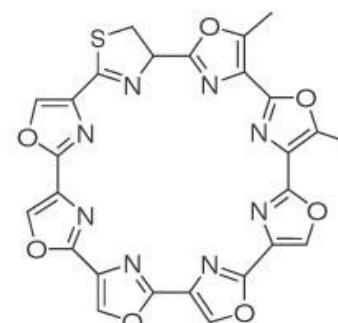
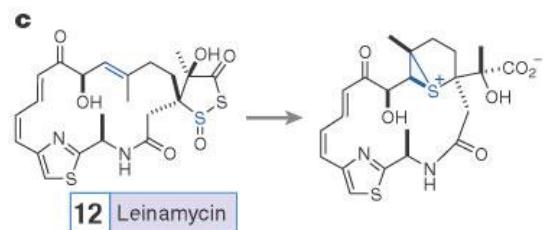
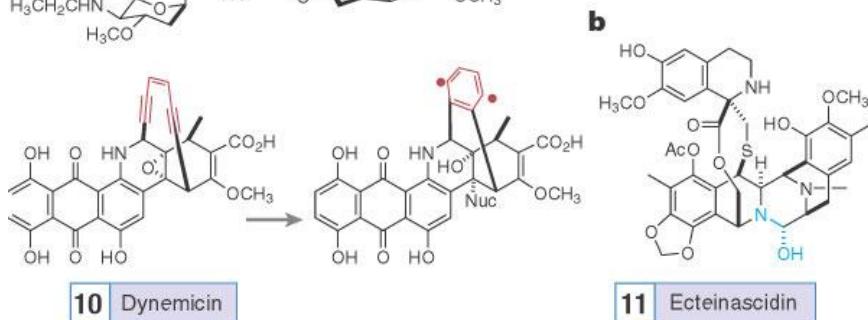
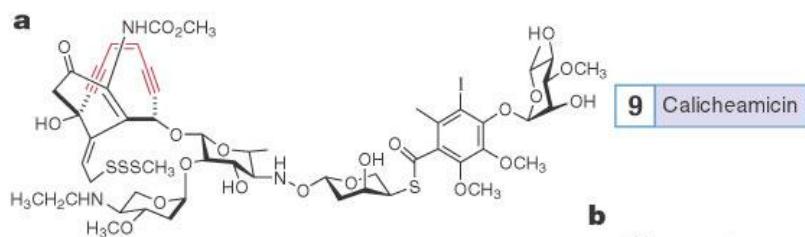


Espaço Químico

basis of a variety of molecular properties

combinatorial chemistry







Molecular Plot

File



DATA SOURCES

Abbott kinase dataset

PLOTS

FLT1 vs FLT3

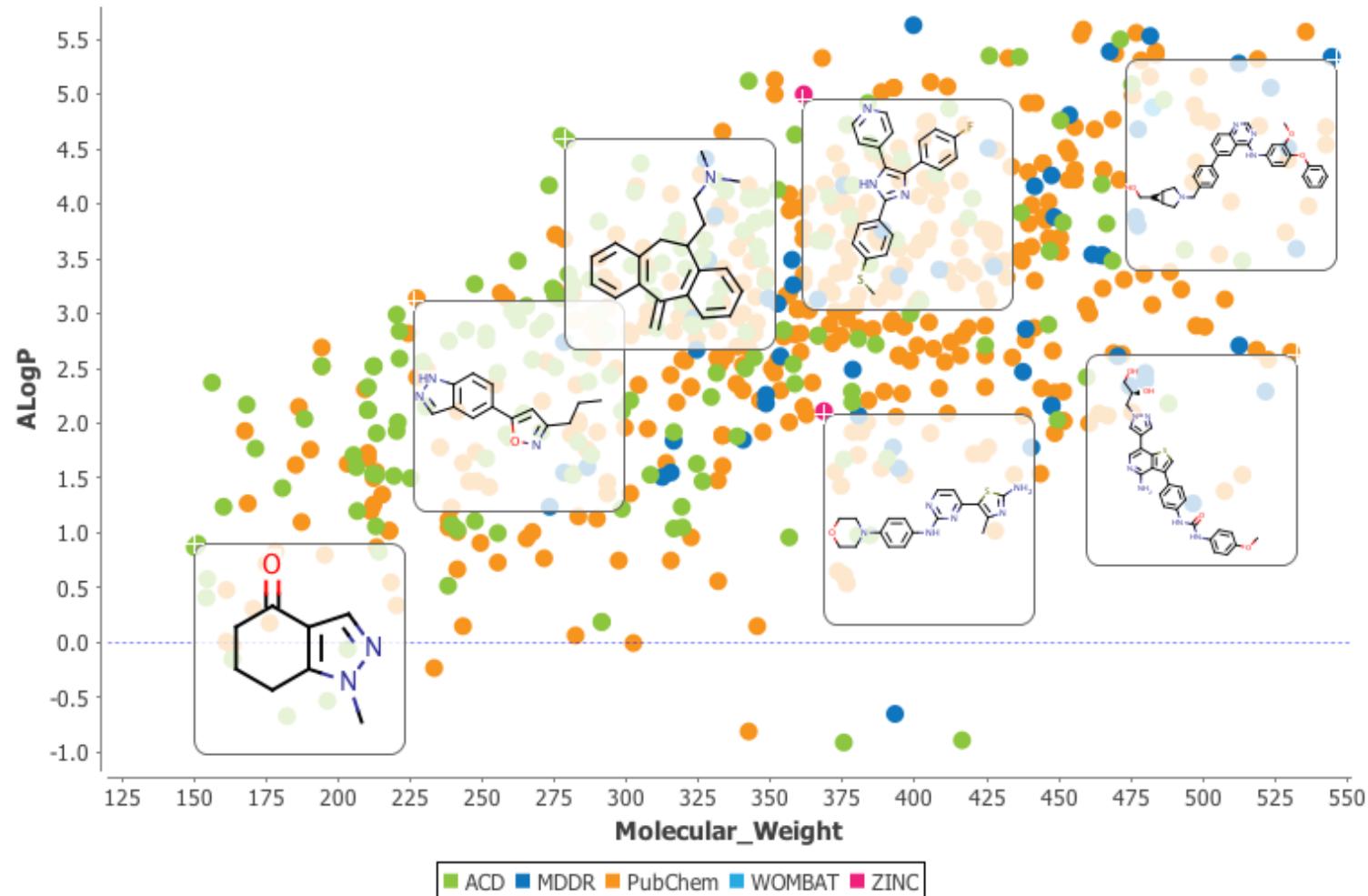
ALogP vs MWT

AKT1 vs AKT2

JAK2 vs JAK3

DYRK1A vs DYRK1B

ALogP vs MWT



Metabolômica: Desafios



Metabólitos

Primários

**Lipídeos, açucares,
ácidos graxos**

Secundários

**Policetídeos, alcaloides,
NRP, terpenos**





Metabolomica

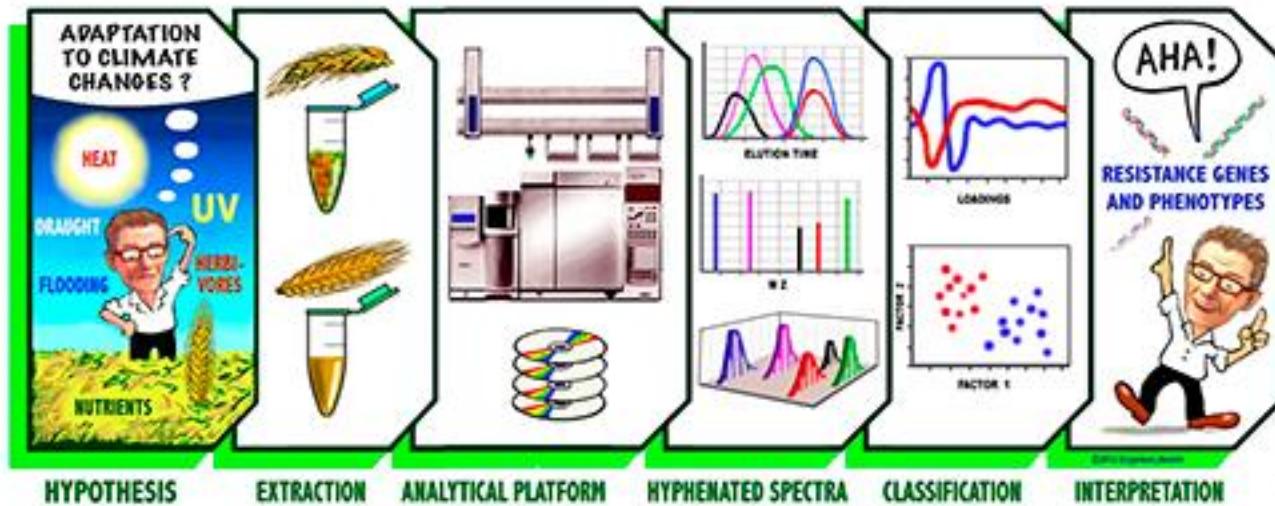
Foodomics é uma disciplina que examina todo o conjunto de substâncias presentes em nossos alimentos (foodome). A disciplina utiliza plataformas analíticas avançadas para investigar a composição do alimento e, assim, suas propriedades nutricionais e impacto na saúde. As novas técnicas também fornecem uma imagem detalhada da qualidade dos alimentos e podem ser usadas para detectar fraudes em alimentos e encontrar soluções para outros desafios na produção de alimentos. Os resultados de pesquisas de foodomics têm um impacto direto sobre os consumidores, a indústria de alimentos e a sociedade.

Engelsen

(https://food.ku.dk/english/research_at_food/research_fie_ids/foodomics/)

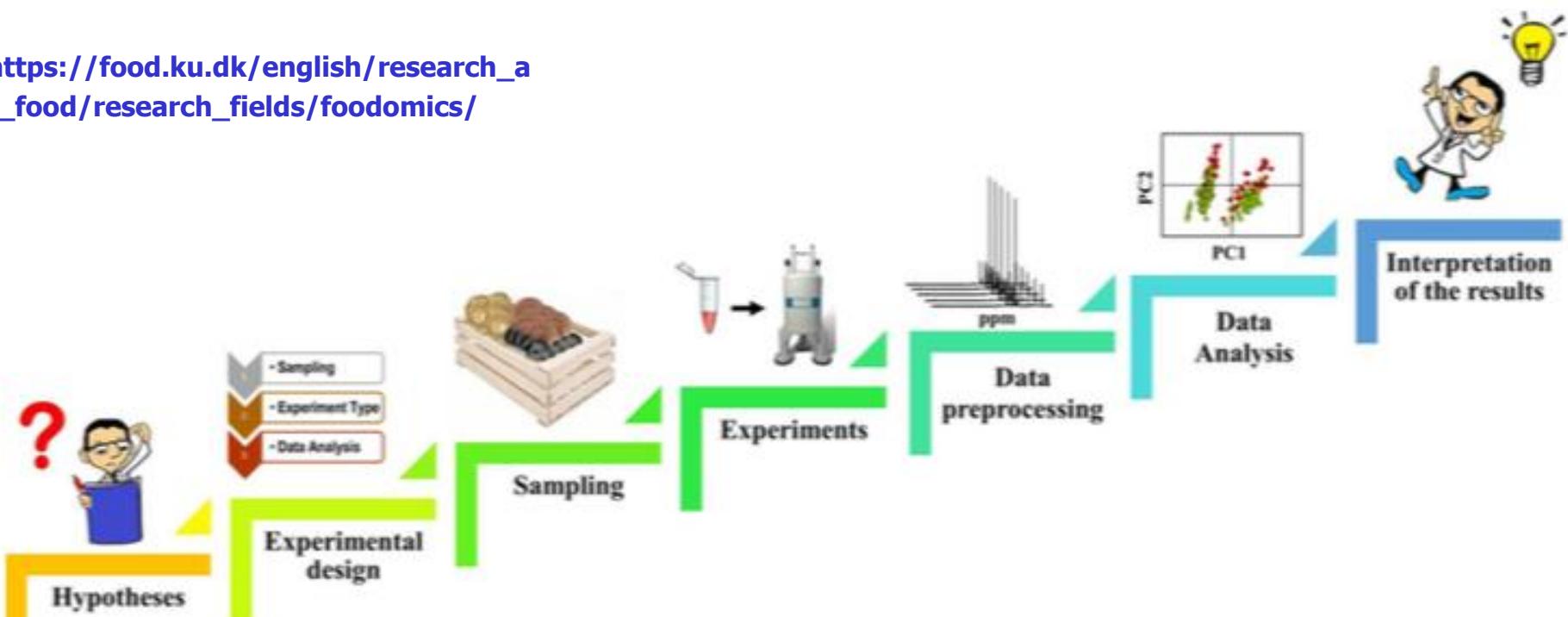
Foodomics—a discipline that studies the food and nutrition domains through the application of advanced omics technologies to improve the consumer's well-being, health, and knowledge.

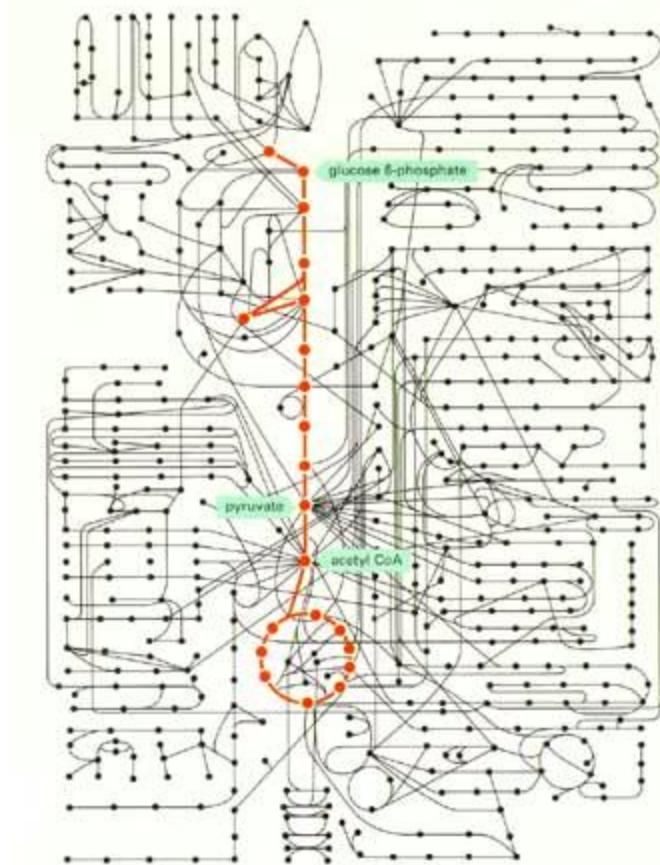
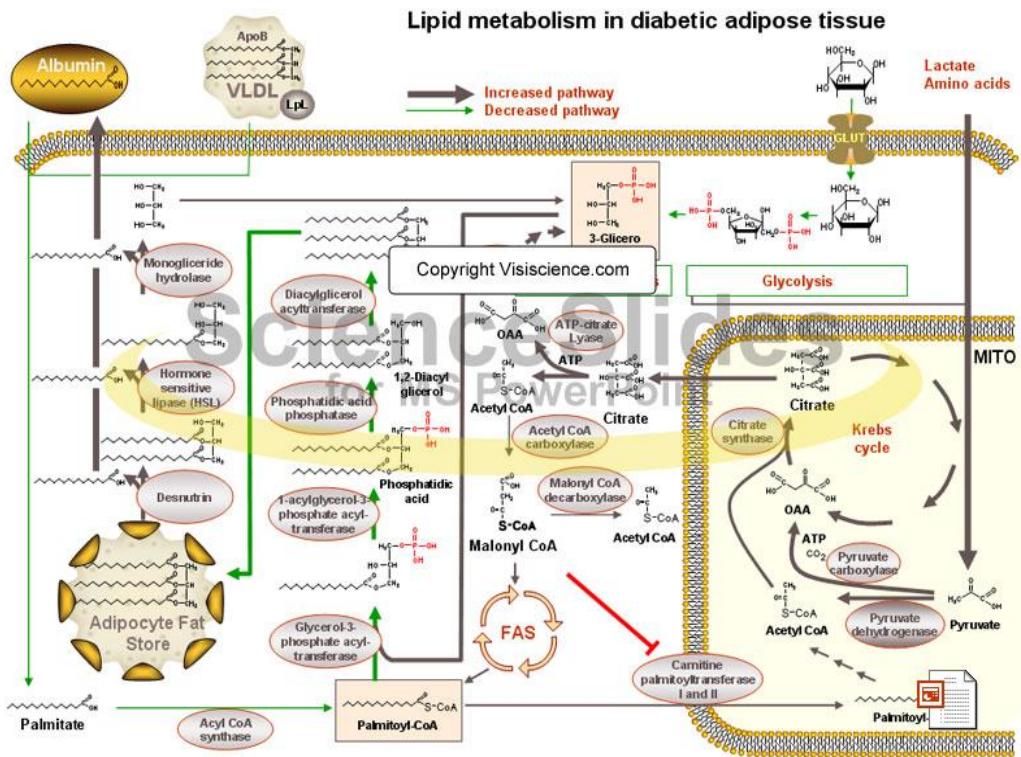
Khakimov & Engelsen, 2017



Foodomics. Illustration by Søren B. Engelsen and Tim Newlin ©

https://food.ku.dk/english/research_at_food/research_fields/foodomics/







Metabolite Profile



fingerprint

Table 1. Main Features of Most The Popular Techniques for Metabolomics

technique	CE-MS	GC-MS	LC-MS	NMR 1D	NMR 2D
universality	++	+	+/-	+++	+++
accuracy	-/+ ^a	-/+ ^a	-/+ ^a	+++	+++
reproducibility	++/- ^a	++/- ^a	++/- ^a	+++	+++
sensitivity	+	++/+++ ^a	+++	-	-
resolution	++	+++	+++	+	++
efficiency	++	-	+	++	++

^aAccuracy, repeatability, and sensitivity mainly depend on the analyzer. GC-MS and LC-MS are usually performed either with a quadrupole or time-of-flight analyzer. "+" or "-" in front of and behind "/" represent quadrupole and time-of-flight analyzer, respectively.

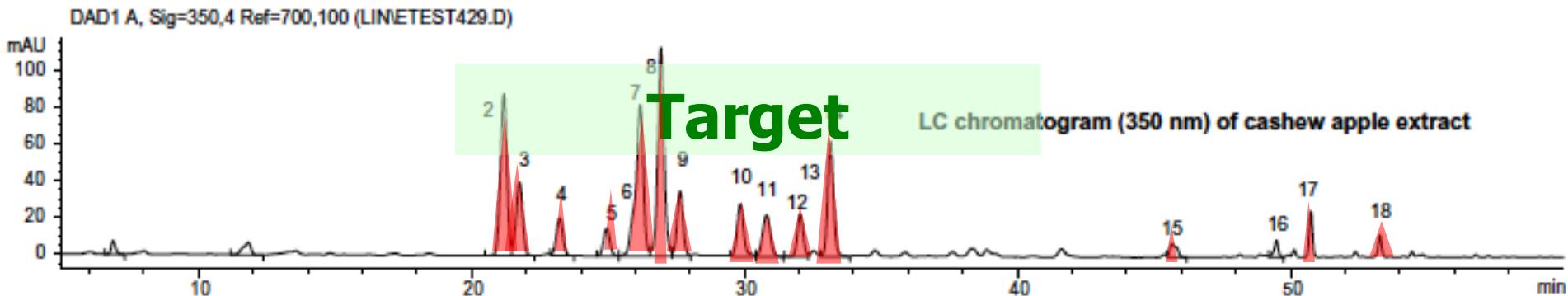
ABORDAGEM?

COMPOSTOS

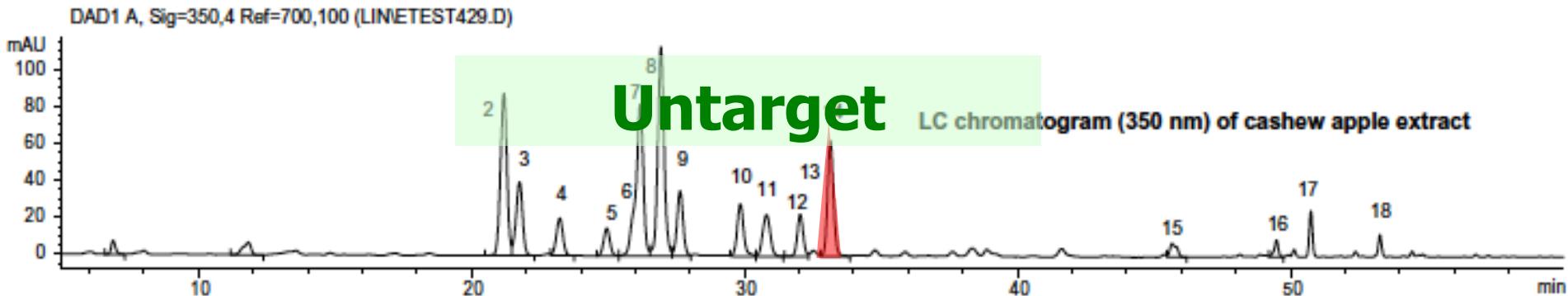
ALVO

NÃO ALVO

Caracterização completa e quantificação



Definição de marcadores e sua posterior quantificação



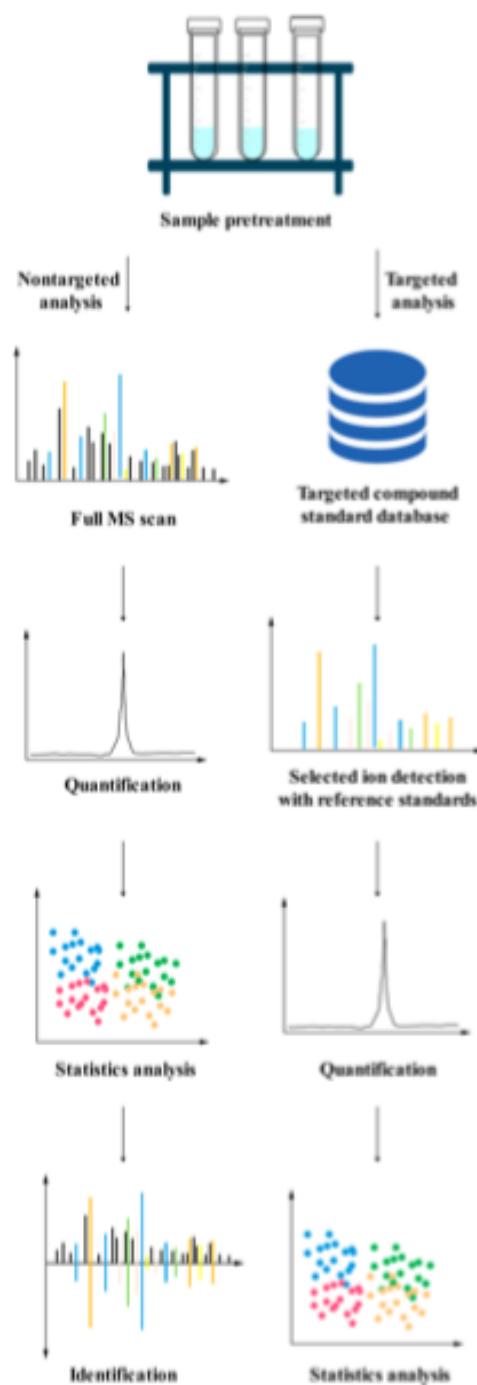
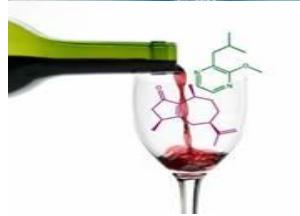
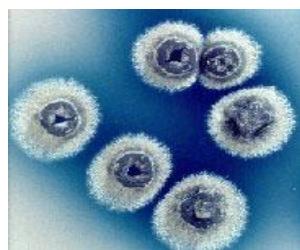
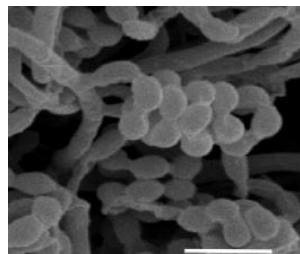


Table 1

Reported applications of metabolomics in life sciences research and practice

Basic research	References
Functional genomics	[2,5,6]
Interaction between metabolome, transcriptome and proteome	[7,8]
Discovery of new biochemical pathways	[9]
Interaction between species	[10]
Analysis of metabolic regulation	[11]
Applied research	References
Medical applications	
Understanding of disease pathophysiology	[12,13]
Disease biomarker identification	[14,15]
Early diagnosis	[16]
Personalized medicine	[17–19]
Clinical trial monitoring	[32,33]
Drug discovery	[34,35]
Toxicology—Drug safety	[36]
Agricultural/Nutrition Applications	
Identification of metabolic engineering targets	[20,21]
Understanding of stress response	[22,23]
Classification of special varieties of produce (e.g. tea, ginseng, fish)	[24–26]
Genetically modified (GM) food certification	[27–29]
Human nutrition	[30,31]
Industrial Applications	
Identification of metabolic engineering targets in <i>Escherichia coli</i> , yeast, algae	[37]
Fermentation process improvement	[38]
Biologics production and fermentation process optimization	[39]

Instrumentação: UPLC – QTOF – MS/MS

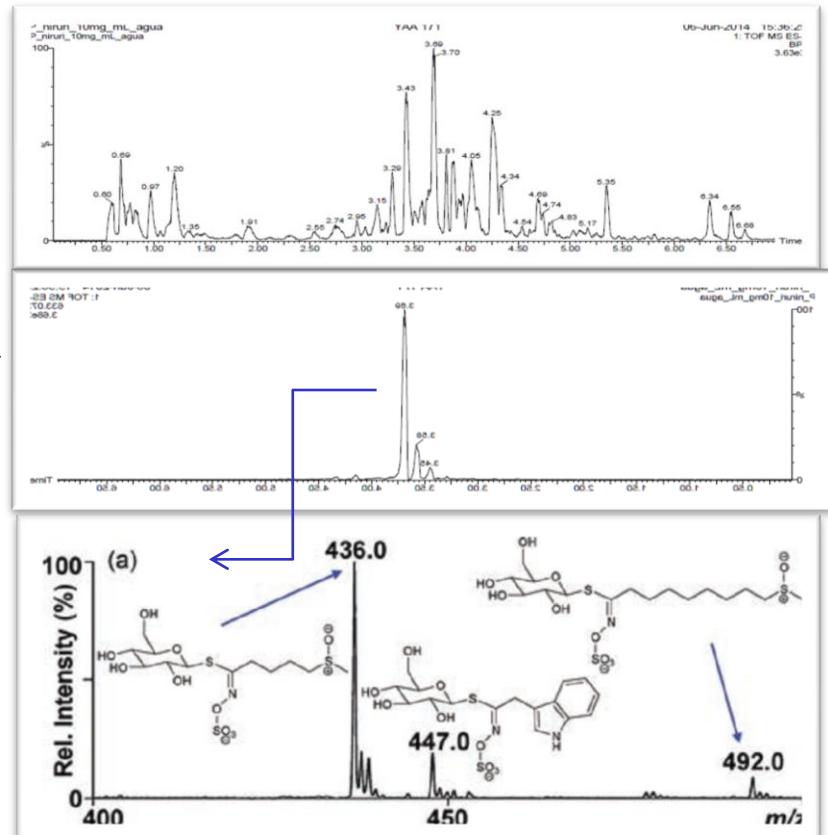


Algumas aplicações:

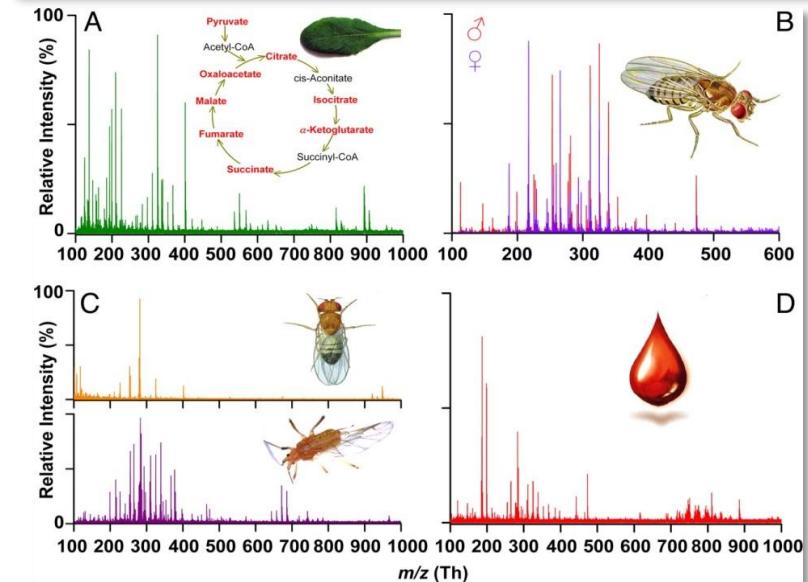
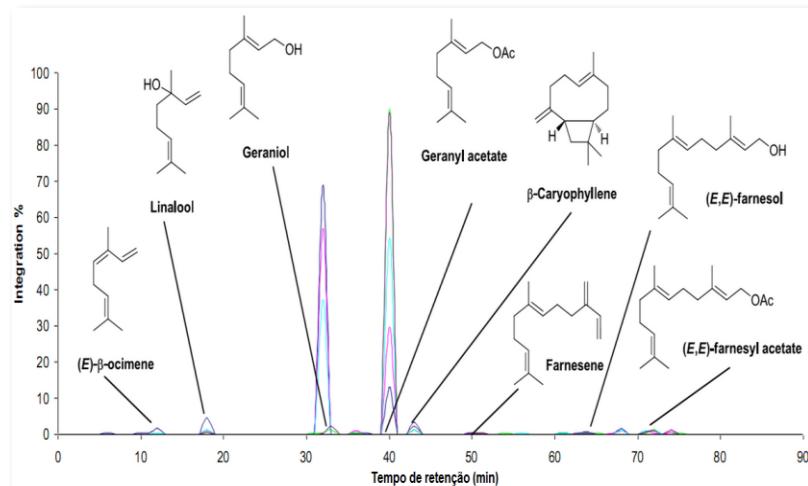
Produtos naturais -

Insumos agroindustriais

Determinação estrutural de substâncias de origem vegetal, animal e de microorganismos



Instrumentação: GC-MS/MS



Agumas aplicações:

Análises de aromas de vinhos

Óleos essenciais

Feromônios

Estudos de ecologia química

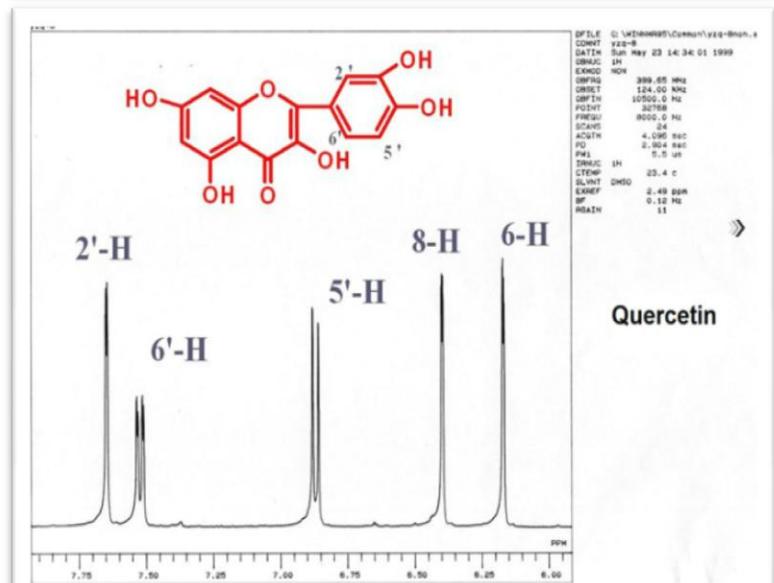
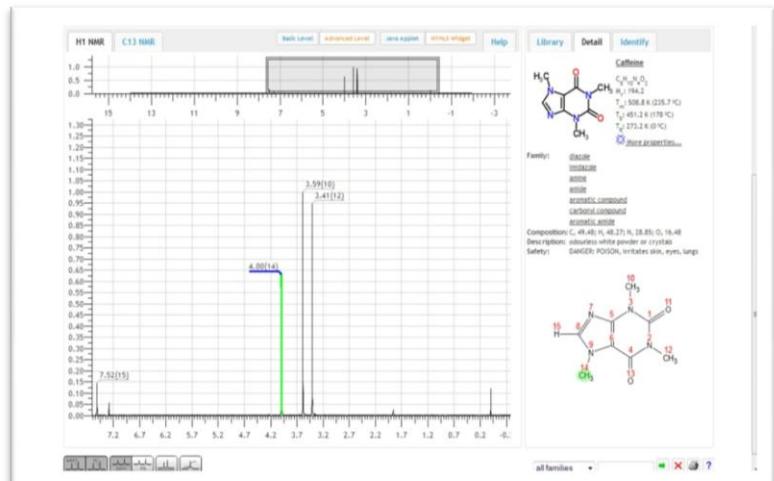
Instrumentação: Ressonância Magnética Nuclear (RMN)

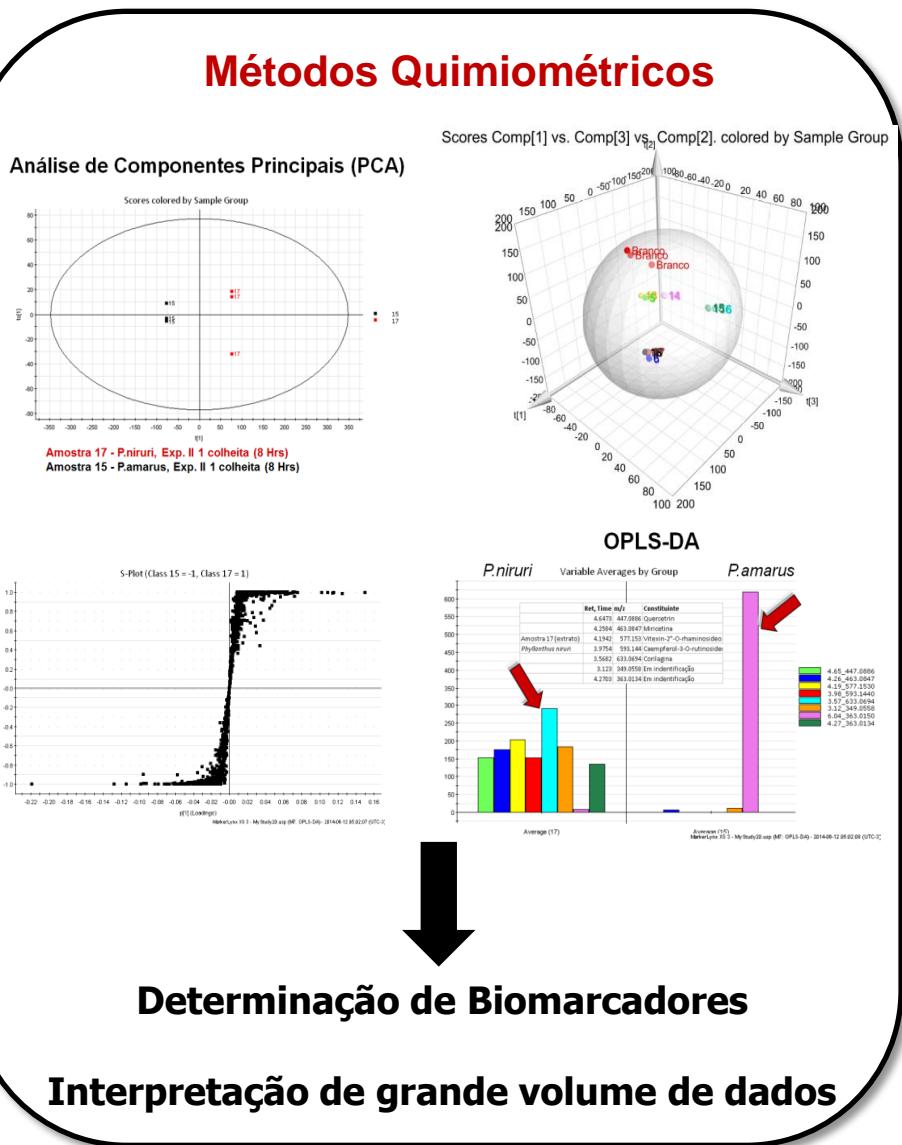
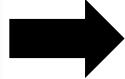
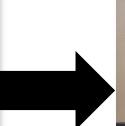
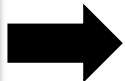


Sistema de detecção universal

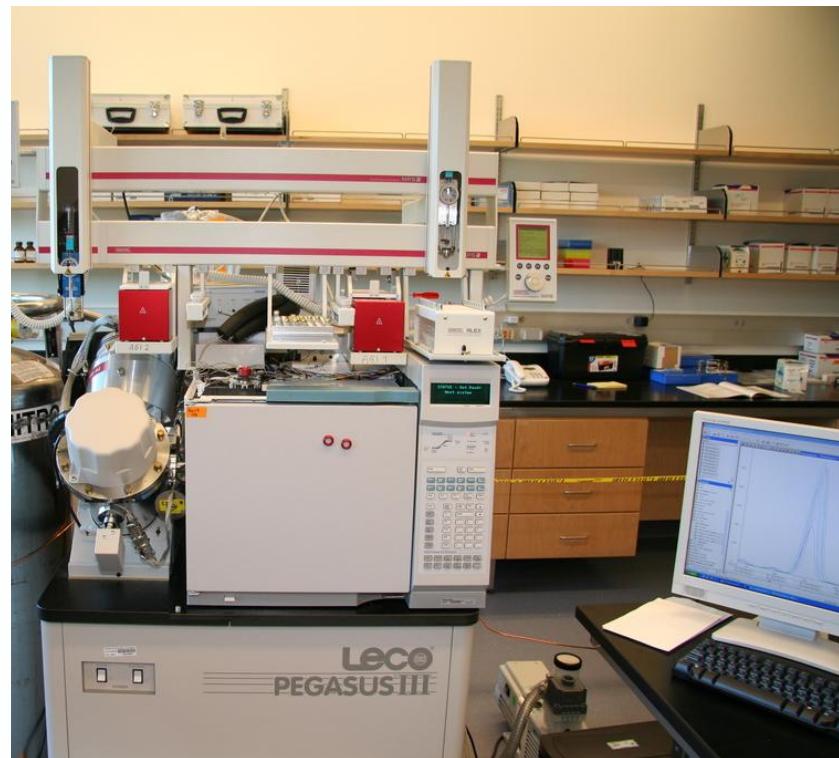
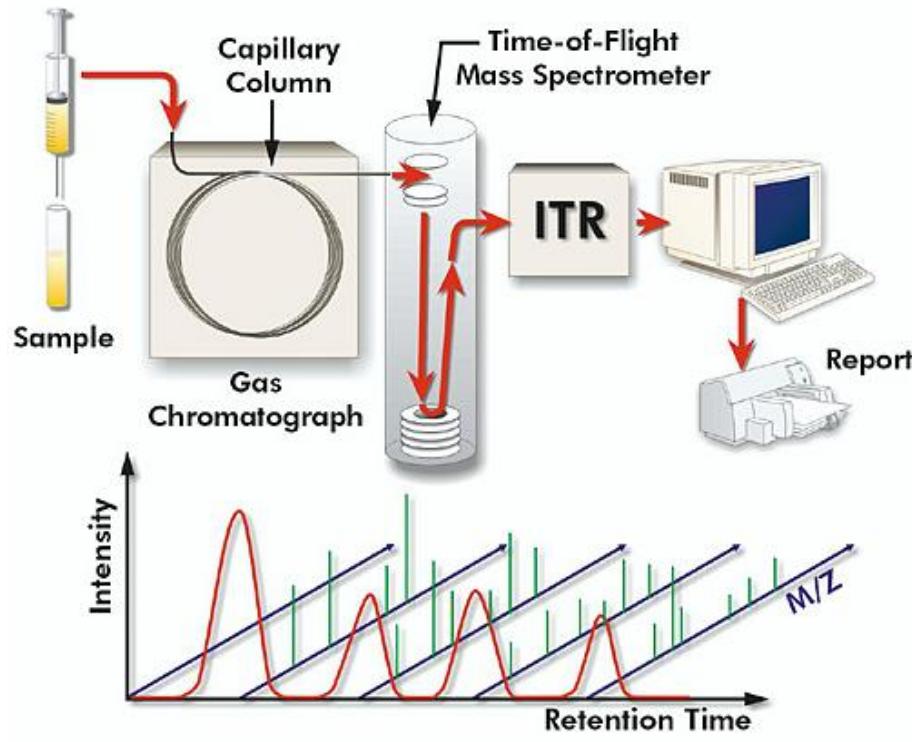
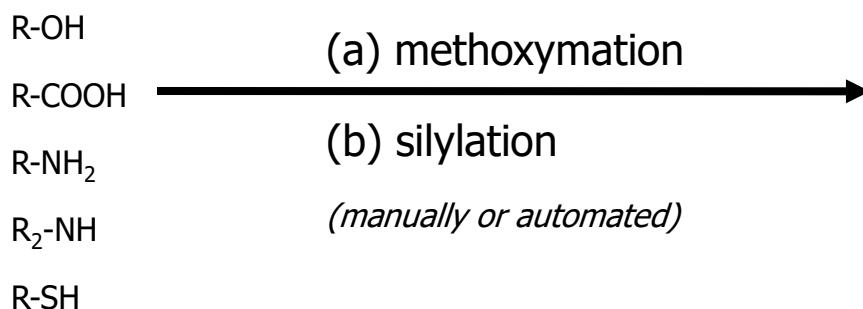
Estudos de Metabolômica e Elucidação estrutural

Quantificação sem necessidade de padrões analíticos



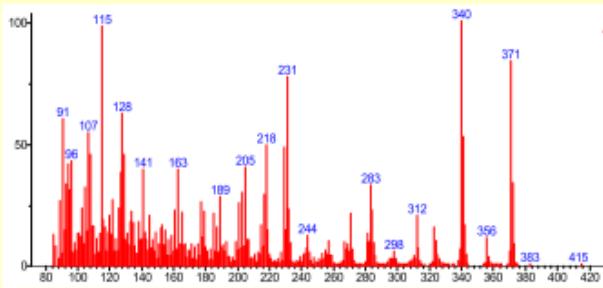


Metabolite profiling by GC-EI-TOF covers ~ 50-550 Da

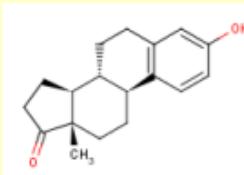
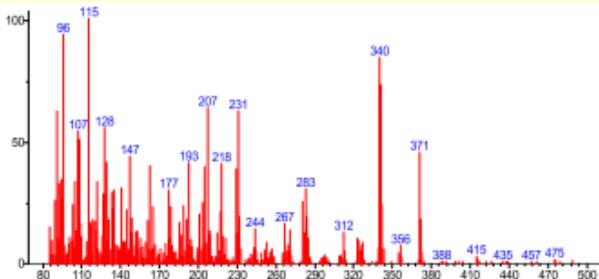


Library – Naming convention must be consistent but is not trivial

Estrone major
FiehnRI 948753



Estrone minor
FiehnRI 950990



Estrone

Solution:

mass spectra + retention time point to the biological name estrone with PubChem [CID: 5870](#) and KEGG ID and InChIKey=DNXHEGUUPJUMQT-UHFFFAOYAI

[Information](#)[Viewer](#)[Compounds](#)[Edit](#)[Log](#)**C.1304.2 (1H NMR)**

2.737E2

1.368E2

-0.126

11.098

8.098

5.098

2.098

-0.902

ppm

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11.0985

Imax: 273.6926

Ok

<

0.01

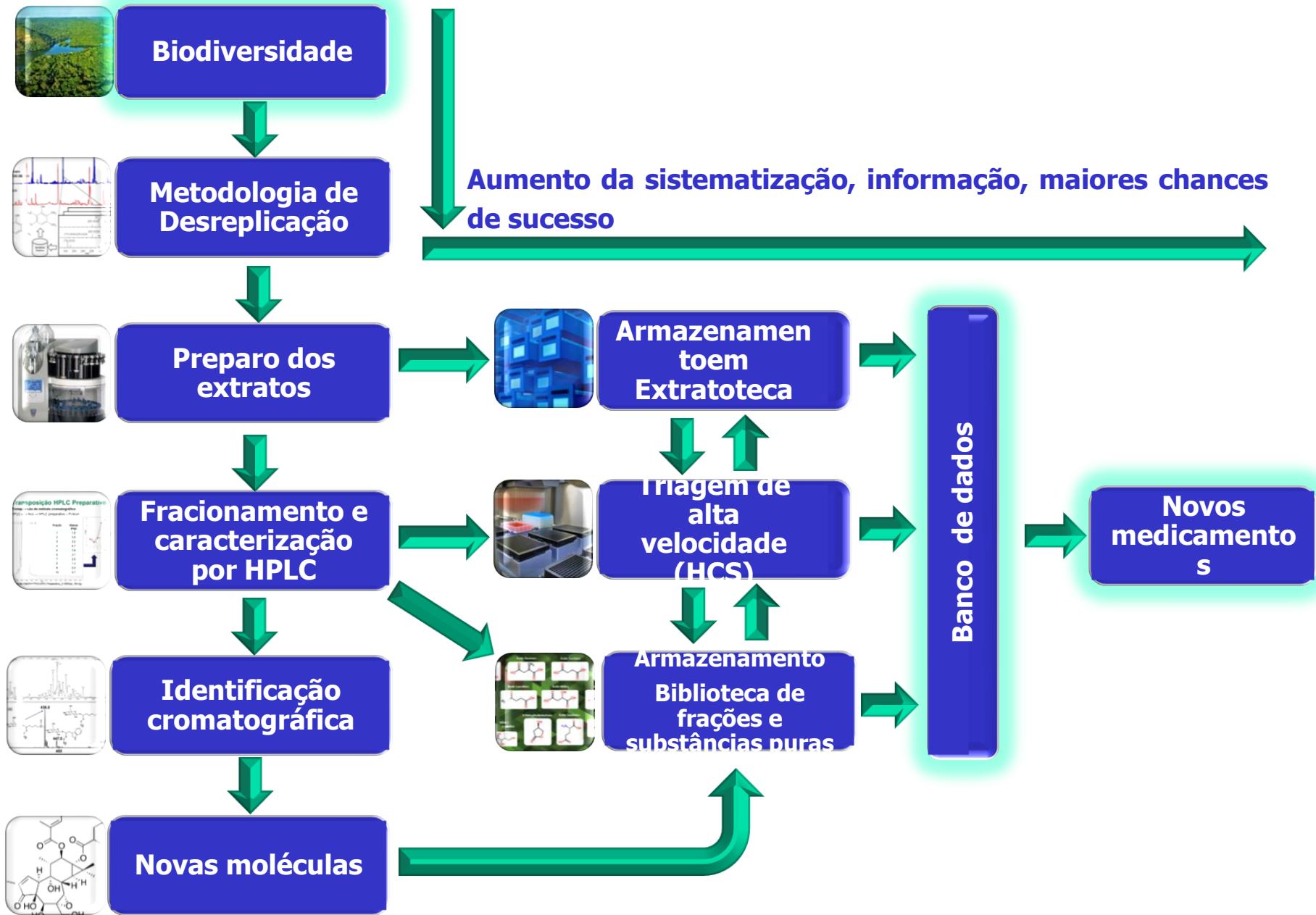
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Zoom: -

All

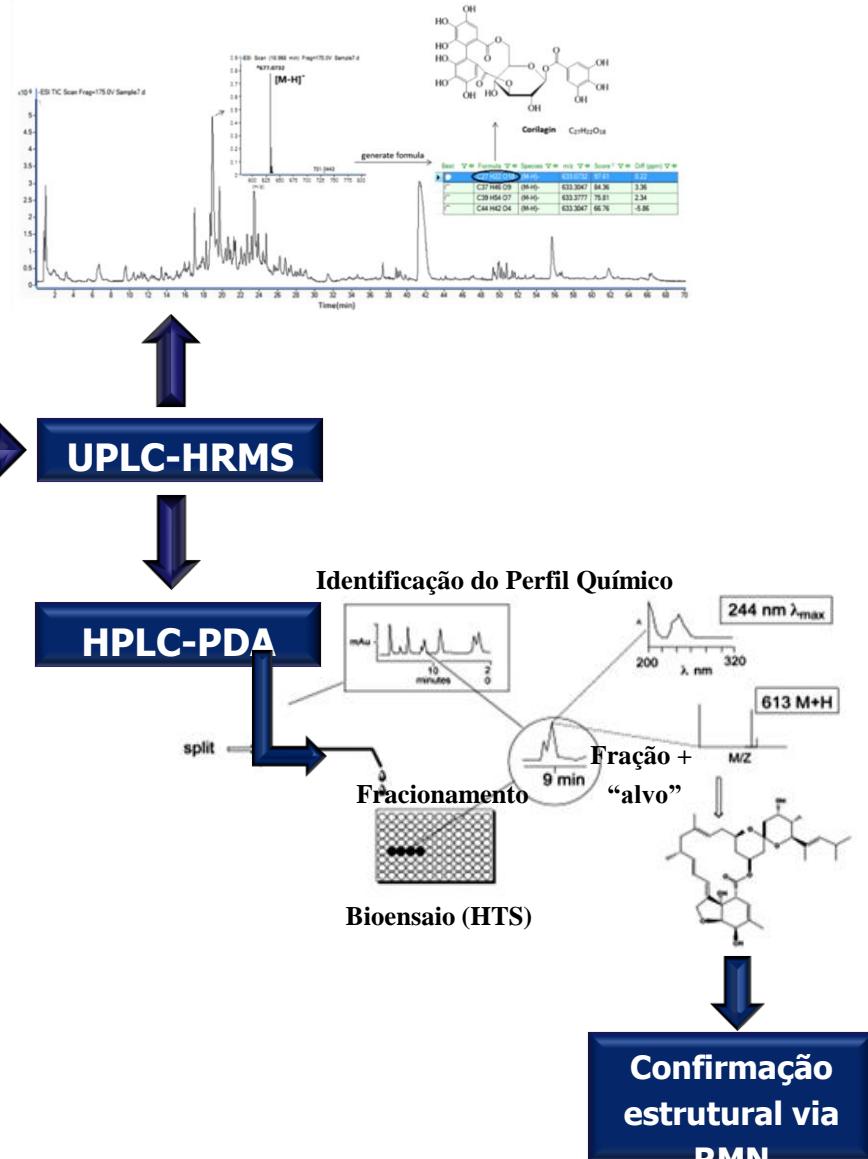
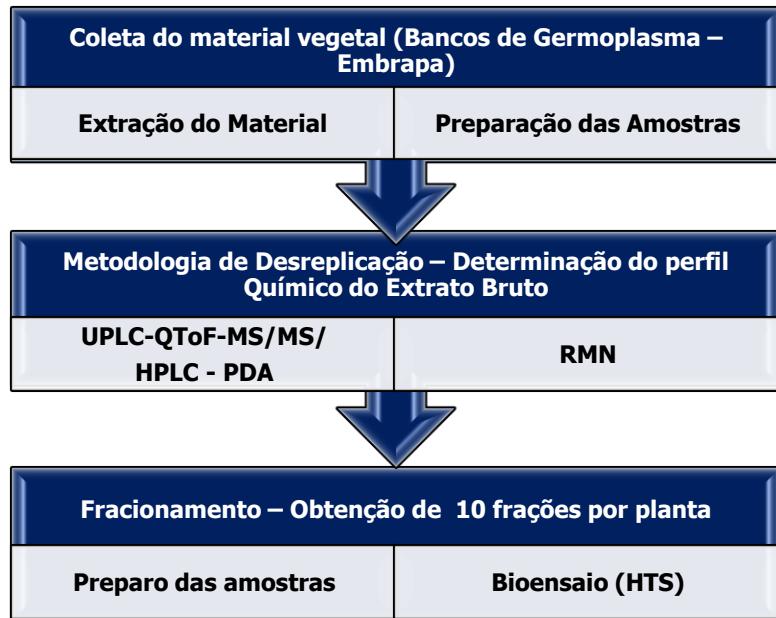
RMN

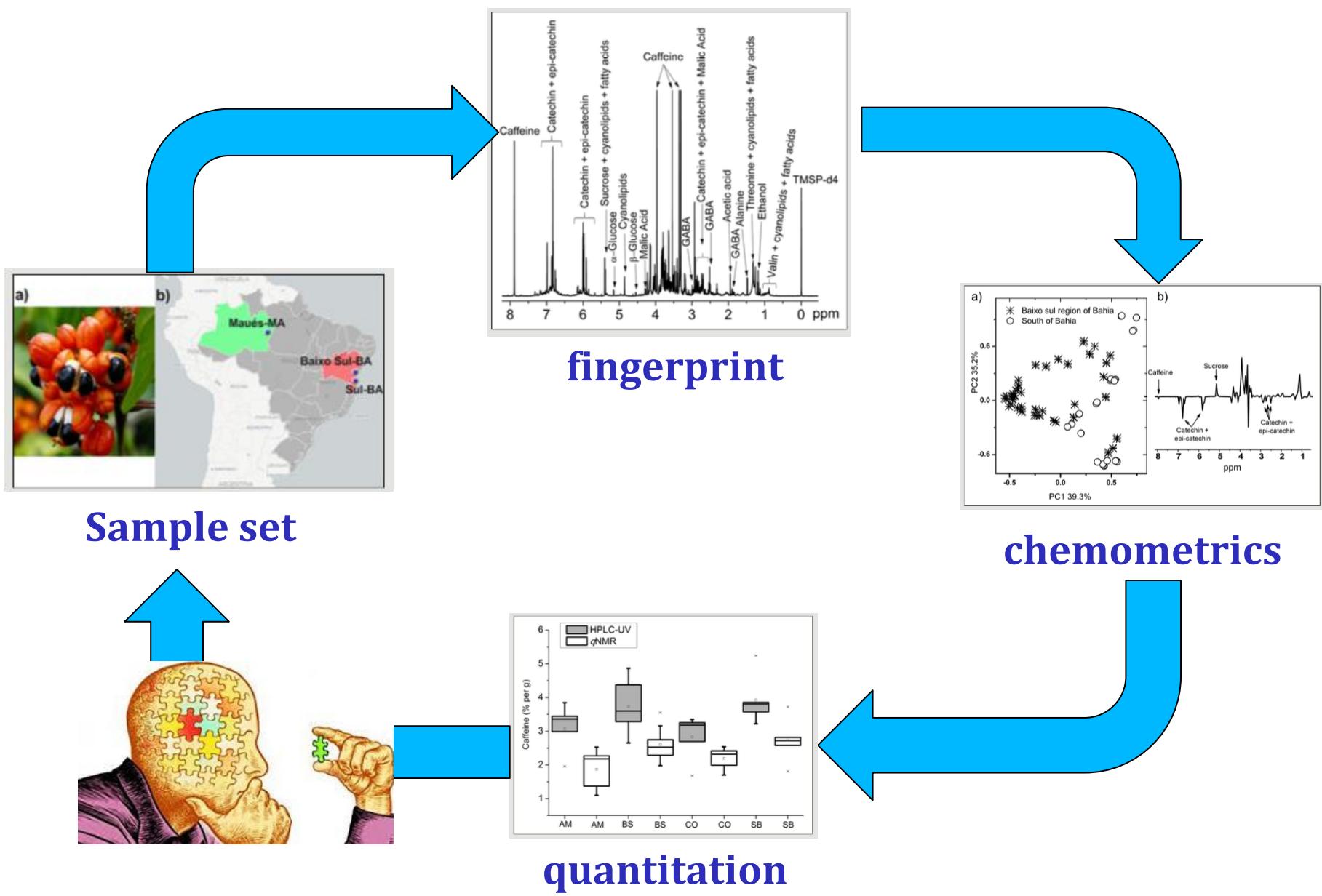
Bioprospecção de Moléculas com Potencial Terapêutico



Estratégia de Desreplicação

Dinâmica de Pesquisa





Qual a minha hipótese?

Metabolômica e Nutrição: condições experimentais



Review | Free Access

Nutrimetabolomics: An Integrative Action for Metabolomic Analyses in Human Nutritional Studies

Marynka M. Ułaszewska, Christoph H. Weinert, Alessia Trimigno, Reto Portmann ... See all authors ▾



Volume 63, Issue 1
Special Issue: Metabolomics:
A Powerful Tool to Enrich our
Understanding of the Impact
of Food on Health

January 2019
1800384

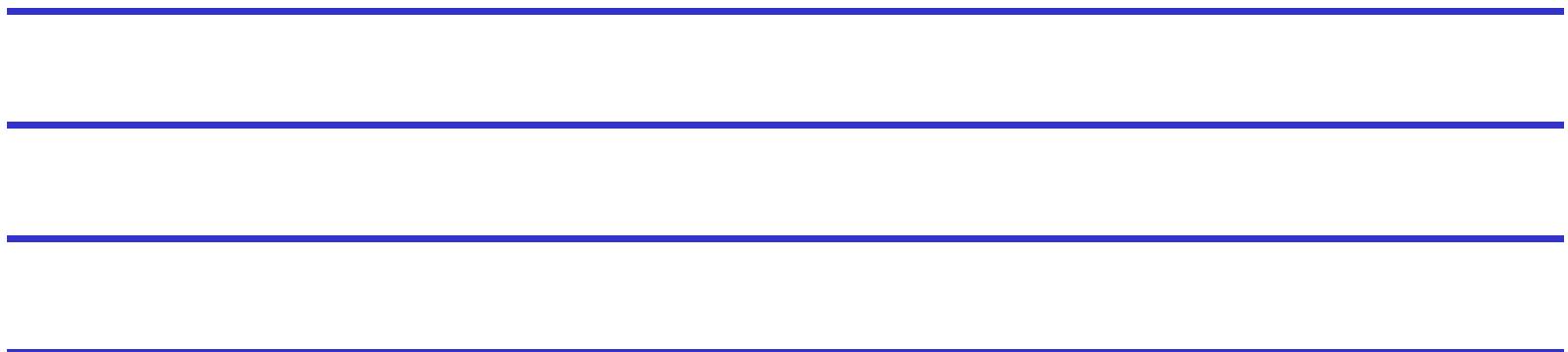
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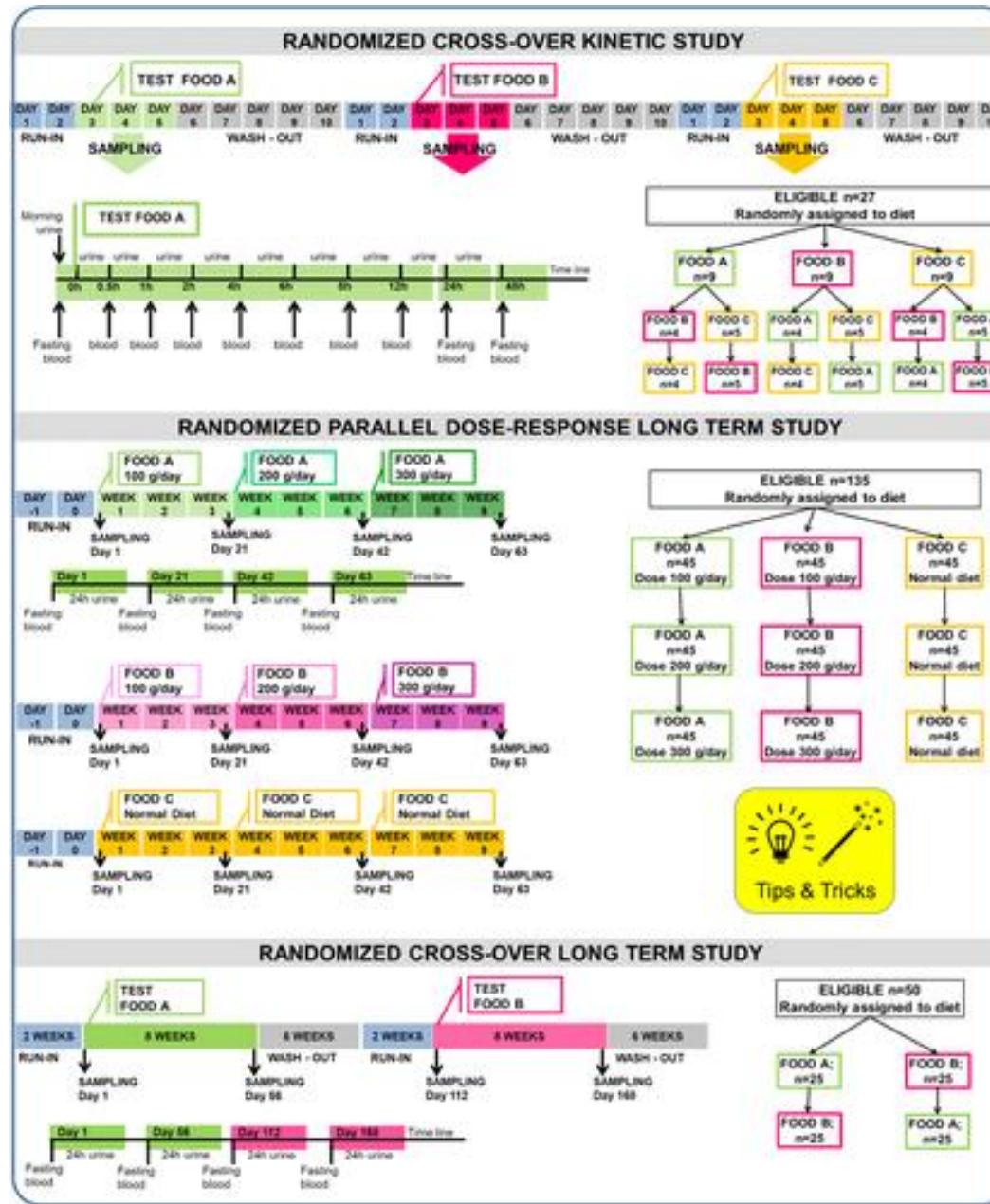
Nutrimetabolomics: An Integrative Action for Metabolomic Analyses in Human Nutritional Studies

*Marynka M. Ułaszewska, Christoph H. Weinert, Alessia Trimigno, Reto Portmann, Cristina Andres Lacueva, René Badertscher, Lorraine Brennan, Carl Brunius, Achim Bub, Francesco Capozzi, Marta Cialiè Rosso, Chiara E. Cordero, Hannelore Daniel, Stéphanie Durand, Bjoern Egert, Paola G. Ferrario, Edith J.M. Feskens, Pietro Franceschi, Mar Garcia-Aloy, Franck Giacomoni, Pieter Giesbertz, Raúl González-Domínguez, Kati Hanhineva, Lieselot Y. Hemeryck, Joachim Kopka, Sabine E. Kulling, Rafael Llorach, Claudine Manach, Fulvio Mattivi, Carole Migné, Linda H. Münger, Beate Ott, Gianfranco Picone, Grégory Pimentel, Estelle Pujos-Guillot, Samantha Riccadonna, Manuela J. Rist, Caroline Rombouts, Josep Rubert, Thomas Skurk, Pedapati S. C. Sri Harsha, Lieven Van Meulebroek, Lynn Vanhaecke, Rosa Vázquez-Fresno, David Wishart, and Guy Vergères**

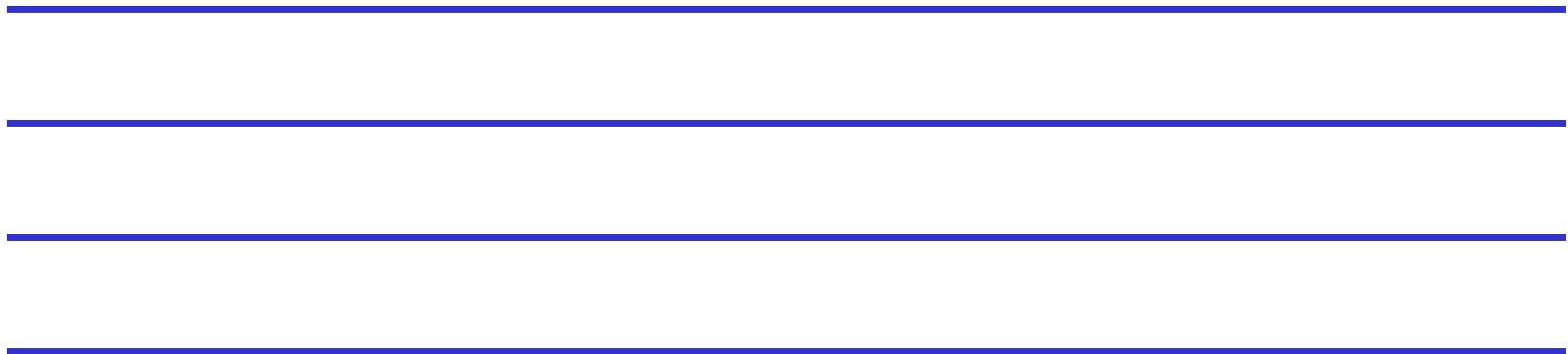
Qual o desenho experimental?



Nutrimetabolomics: An Integrative Action for Metabolomic Analyses in Human Nutritional Studies



Amostras?





Spot urine sample



24 h urine sample



Sarstedt®

MarketLab®

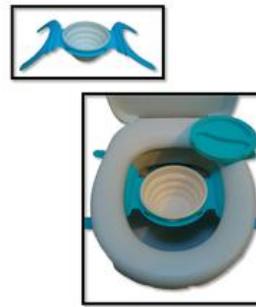
URINE COLLECTION

Wide-mouth plastic bag and a plastic container



FECES COLLECTION

Stool specimen collection units



Fecotainer®

Freezing toilet at -30°C



Toilet type T-1970, Gisebo;
Privetti® Pikkuvihrea



collection of 24h urine sample
requires **an instruction for
volunteer.**

Samples should be transferred/delivered to laboratory as soon as possible for further storage (< 2h).



avoid stool contamination with water, urine or other
materials (e.g. toilet paper).
An instruction for volunteer is required.

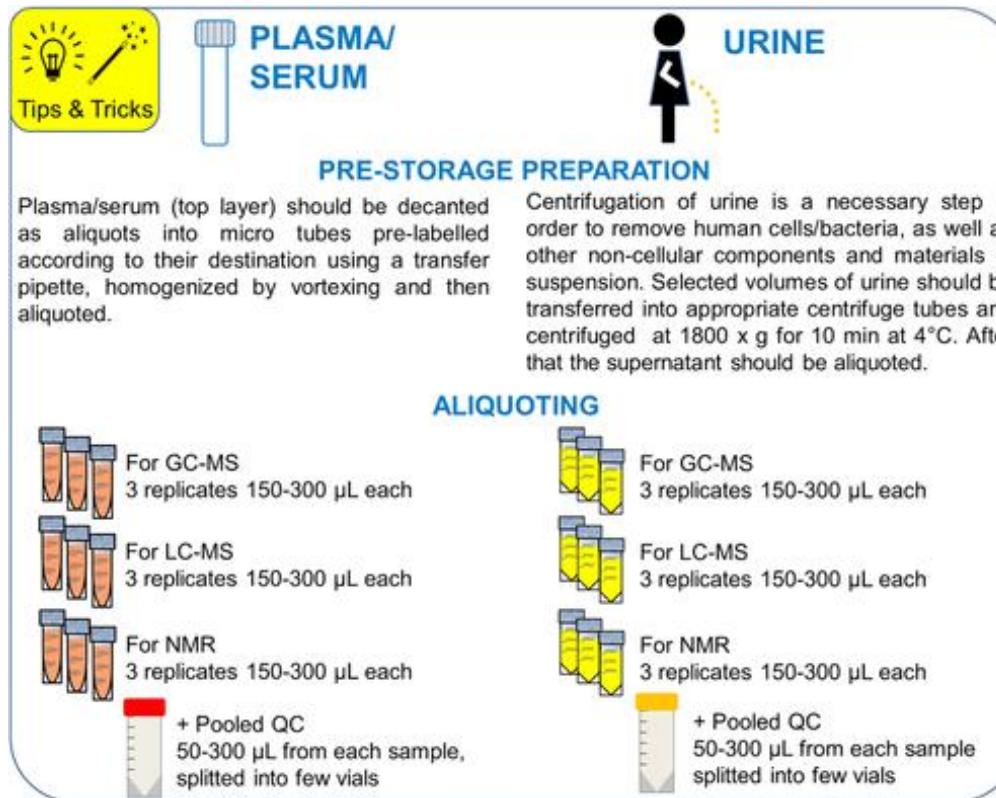
In contrast to serum/plasma, urine and feces require sample specific normalization.

Volume and weight of urine and feces and thus the overall concentration of metabolites may vary drastically.
Information such as volume and weight for both matrices should be collected at sample arrival to the laboratory,
before samples aliquotiation.

Preparação de amostras

Table 2. Overview of commonly used solvents for microbial metabolite extraction.

Method	Organism	Ref.
<i>For general extraction of different chemical classes</i>		
80:20 methanol:water	<i>Escherichia coli</i>	[103–105]
50:50 methanol: water	<i>Schizosaccharomyces pombe</i>	[106]
60:40 methanol:water	<i>Streptomyces coelicolor</i>	[33]
72:28 methanol	<i>Saccharomyces cerevisiae</i>	[107]
30:30:20 methanol:water:chloroform	<i>Corynebacterium glutamicum</i>	[108]
60:20:20 methanol:water:chloroform	<i>Leishmania donovani</i>	[80]
40:40:20 acetonitrile:methanol:water with 0.1% formic acid	<i>S. cerevisiae</i> <i>E. coli</i>	[31] [66]
40:40:20 acetonitrile:methanol:water	<i>Mycobacterium tuberculosis</i> <i>E. coli</i> <i>S. cerevisiae CEN.PK 113–7D</i>	[109] [110] [61]
Boiling HEPES-buffered ethanol solution (75:25 v/v ethanol:water; pH=5.2)	<i>Methylbacterium extorquens</i>	[111]
60:40 ethanol:water	<i>Staphylococcus aureus</i>	[58]
Pure cold methanol	<i>E. coli</i>	[66,71,72,112]
Boiling ethanol	<i>S. cerevisiae CEN.PK 113–7D</i> <i>E. coli, B. subtilis, S. cerevisiae</i> <i>S. cerevisiae</i> <i>Pichia pastoris</i>	[61] [49] [60,69,113–115] [116]
<i>For specific classes of compounds</i>		
Alkaline extraction (for basic molecules)	<i>M. extorquens</i>	[111]
Acid extraction (for organic acids)	<i>M. extorquens</i>	[111]
Bligh–Dyer method (for lipids)	<i>Trypanosoma brucei</i>	[117]



PRE-STORAGE PREPARATION



Tips & Tricks

FECES



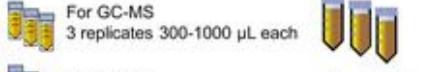
HOMOGENIZATION OF STOOL SAMPLE:

- automatic homogenization of a whole plastic bag in stomacher or blender
- stirring of fresh sample with a sterile spatula directly in delivery bag/container
- collecting multiple aliquots of i.e. 20 mg from the same area below the surface of the stool

PREPARATION OF SAMPLE AFTER HOMOGENIZATION:

- a. fresh feces freezing at -80° C
- b. centrifuging of fresh feces with or without portions of extracting agent (ice-cold PBS, 95% ethanol, etc.) and collection of supernatants (fecal water)
- c. feces freeze-drying (fecal powder)

ALIQUOTING

Fresh feces freezing	Fresh feces centrifuging: fecal water
	
For GC-MS 3 replicates 10-50 g each	For GC-MS 3 replicates 300-1000 µL each
For LC-MS 3 replicates 10-50 g each	For LC-MS 3 replicates 300-1000 µL each
For NMR 3 replicates 10-50 g each	For NMR 3 replicates 300-1000 µL each
+ Pooled QC 0.5-10 g from each sample splitted into several vials	
Feces freeze-drying: fecal powder	
	
For GC-MS 3 replicates 50-400 mg each	+ Pooled QC approx. 30-100 mg from each sample splitted into several vials
For LC-MS 3 replicates 50-400 mg each	
For NMR 3 replicates 50-400 mg each	

!

Fecal powder is hygroscopic, weigh with caution. Verify the weight of one spatula of fecal powder, and fill eppendorf tube/vial with only approximative amount (i.e. ca 50 mg or 100mg). Take note of exact weight on the sample label.

Molecular Nutrition & Food Research, Volume: 63, Issue: 1, First published: 03 September 2018, DOI: (10.1002/mnfr.201800384)

Nutrimetabolomics: An Integrative Action for Metabolomic Analyses in Human Nutritional Studies

Tips & Tricks

SERUM
Clot for 30 min at room temperature, centrifuge 10 min at 4°C and 2,500 × g

PLASMA
Mix with anticoagulant, centrifuge 10 min at 4°C and 2,500 × g

HEMOLYSIS PROBLEM:
The breakdown of blood cells strongly alters metabolic profiles of blood-derived samples, by increasing the concentrations of numerous metabolites coming from the intracellular space as well as by inducing the degradation of some compounds by the action of released enzymes

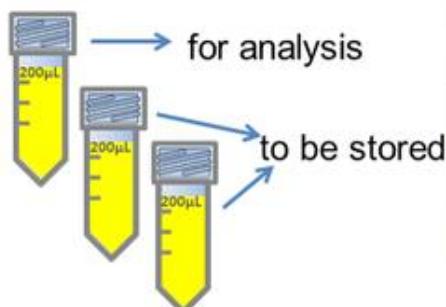
NORMAL SAMPLE **HEMOLYSIS SAMPLES**

HEMOLYZED SAMPLES SHOULD BE AVOIDED IN METABOLOMIC STUDIES

Qual abordagem analítica?

UNTARGETED ANALYSIS

- LC-MS: polar and medium-polar metabolites (i.e., polyphenols and its metabolites, amino acids)
- LC-MS: non-polar metabolites (i.e., lipidomics)
- GC-MS: volatile metabolites
- GC-MS: metabolites after derivatization
- NMR



MINIMUM REQUIREMENT

TARGETED ANALYSIS

- LC-MS (kits, etc)
- GC-MS quantitative analysis with/without derivatization (i.e., SCFA, sugars etc.)
- Clinical analysis (multiple choices)

In general multiple aliquots of smaller volume (i.e., 200 μ L) are recommended, rather than few aliquots of high volume.

Higher number of aliquots is recommended, which allows for additional analysis (i.e., additional analytical technique that arises in the future)



Tips & Tricks

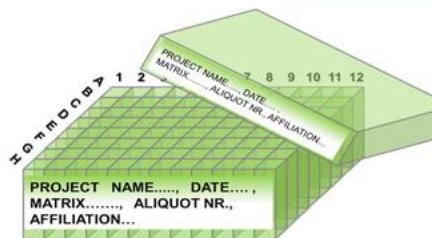
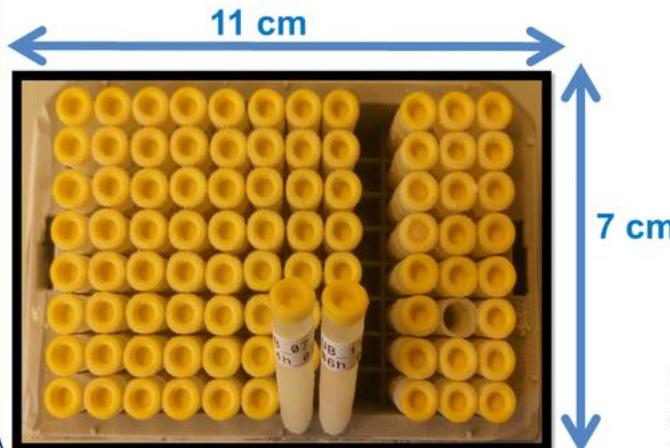
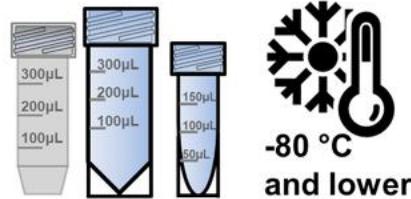
3 ALIQUOTS FOR ONE TYPE OF ANALYSIS (i.e., GC-MS)

Use containers adapted for the volume of matrix that will be stored. Matrix should fill possibly 80%-90% of container with minimum amount of air above its surface. Use containers with low adherence in the interior of the tube, and twisted caps that avoid splashing while opening

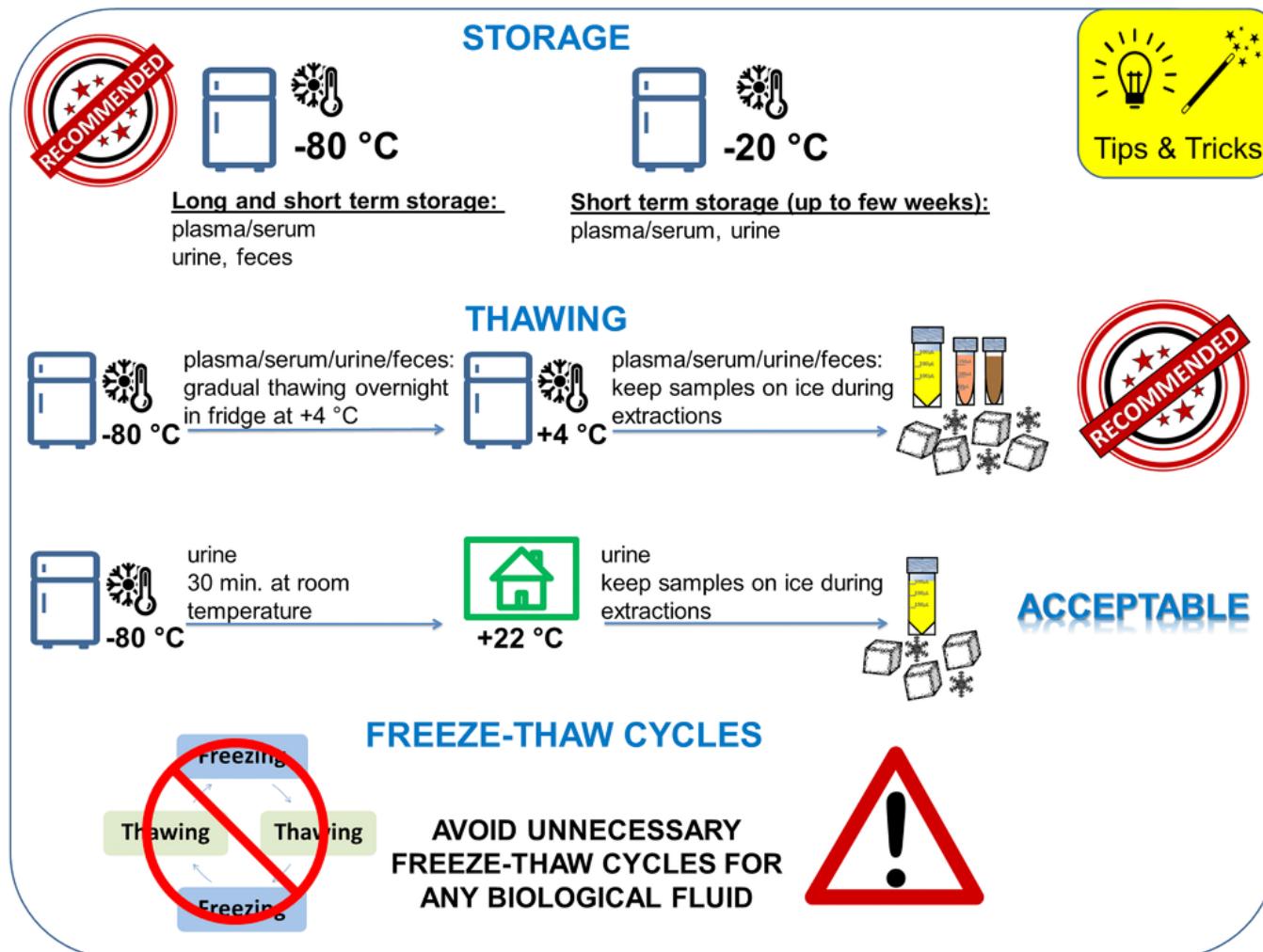


Conical/skirted vials for better recovery of matrix, twisted caps.

Recommended cryovial



Optimization of storage capacity in freezers of small aliquotes in tight boxes. Avoid long term storage in paper boxes.



Nutrimetabolomics: An Integrative Action for Metabolomic Analyses in Human Nutritional Studies



QUALITY CONTROL



Tips & Tricks

Extraction with 96 well plate, preparation of ca. 200 samples/day for LC-MS

DAY 1												DAY 2											
A	Blank	A	Blank																				
B												B											
C												C											
D												D											
E												E											
F												F											
G												G											
H												H											
DAY 1												DAY 2											
A	Blank	A	Blank																				
B												B											
C												C											
D												D											
E												E											
F												F											
G												G											
H												H											

Legend:

- Blank: Blanks of extraction process
- Real samples in random order
- QC pooled sample extracted for evaluation of well plate performance
- QC pooled sample, extracted multiple times

QCs extracts are combined in one **QC pooled vial**, and injected multiple times along injection queue. This quantity is sufficient to cover injections of full sample set at least twice – in both ionization modes. It is recommended to split this QC pooled vial into smaller aliquots.

Example of injection queue, with double QCs injections incorporated every 10 samples. Depending on chromatography time duration, QCs can be incorporated more or less frequently.

Examples:

- Run time is 12 min: double QCs every ca. 8/10 injections
- Run time is 5 min: double QCs every ca. 20 injections

RANDOMIZATION



IT'S
A MUST
!!!

EXAMPLE OF AN INJECTION QUEUE in LC-MS	
DAY 1	x001_solvent x002_solvent x003_QC_equilibration_run x004_QC_equilibration_run x005_QC_equilibration_run x006_QC_equilibration_run x007_QC_equilibration_run x008_Blank1 x009_Blank2 x010_Blank3 x011_solvent x012_QC_pooled x013_QC_pooled x014_QC_pooled x015_QC_pooled x016_urine x017_urine x018_urine x019_urine x020_QC_plate1_1 x021_urine x022_urine x023_urine x024_urine x025_urine x026_solvent x027_QC_pooled x028_QC_pooled x029_urine x030_urine x031_urine ...
DAY 2	x032_urine x033_urine x034_urine x035_urine x036_urine x037_urine x038_urine x039_urine x040_urine x041_QC_pooled x042_QC_pooled x043_urine x044_urine x045_QC_plate2_3 x046_urine x047_urine x048_urine x049_urine x050_urine x051_solvent x052_QC_pooled x053_QC_pooled ...

Molecular Nutrition & Food Research, Volume: 63, Issue: 1, First published: 03 September 2018, DOI: (10.1002/mnfr.201800384)



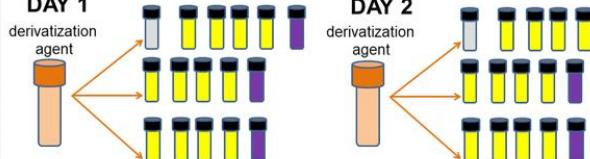
QUALITY CONTROL



Tips & Tricks

**Extraction with derivatization,
preparation of ca. 15/20 samples/day for GC-MS**

DAY 1



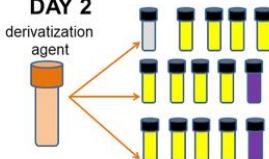
derivatization agent

Blanks of extraction process

QC pooled sample extracted every day few times (i.e., 3)

Real samples in random order

DAY 2



derivatization agent

Time-consuming extraction methods such as derivatization procedures in GC-MS require a different strategy for sample organization. Preparation of one large QC pooled extract is not recommended due to low stability of derivatized extracts.

Every day, a blank, and series of QC samples must be extracted together with randomized study samples. Fresh samples should be analysed within 24-36 h. In the reported queue example, given a run time of ca. 70-75 min, double QCs samples are injected every 4 samples.

RANDOMIZATION

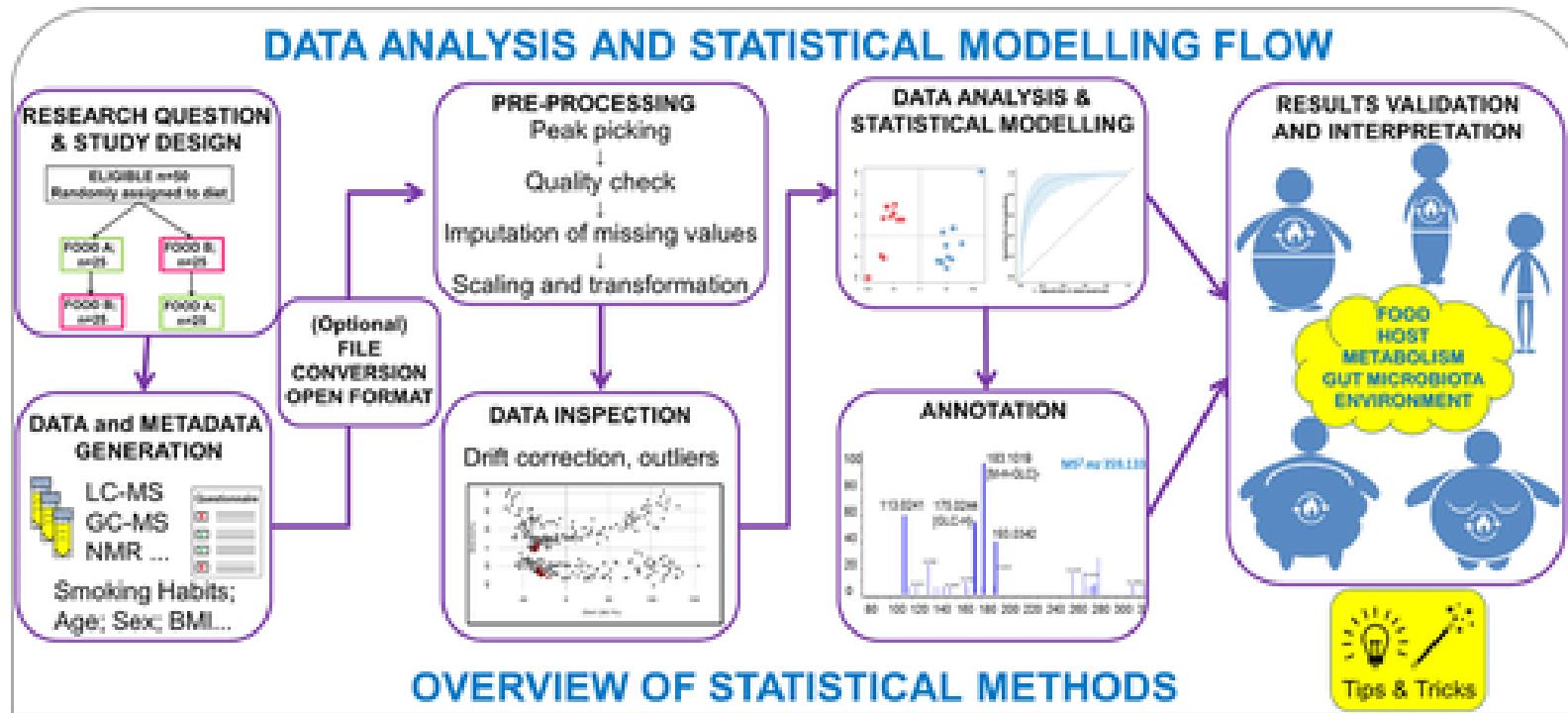


IT'S
A MUST
!!!

EXAMPLE OF AN INJECTION QUEUE in GC-MS

	x001_QC_equilibration_run
	x002_QC_equilibration_run
	x003_QC_equilibration_run
	x004_QC_equilibration_run
	x005_QC_equilibration_run
	x006_Blank01
DAY 1	x007_QC
	x008_QC
	x009_urine
	x010_urine
	x011_urine
	x012_urine
	x013_QC
	x014_QC
	x015_urine
	x016_urine
	x017_urine
	x018_urine
	x019_QC
	x020_QC
	x021_urine
	x022_urine
	x023_urine
	x024_urine
	x025_QC
	x026_QC
DAY 2	x027_Blank02
	x028_QC
	x029_QC
	x030_urine
	x031_urine
	x032_urine
	x033_urine
	x034_QC
	x035_QC
	x036_urine
	x037_urine
	x038_urine
	x039_urine
	x040_QC
	x041_QC
	x042_urine
	x043_urine
	x044_urine
	x045_urine
	x046_QC
	x047_QC

Qual análise estatística?

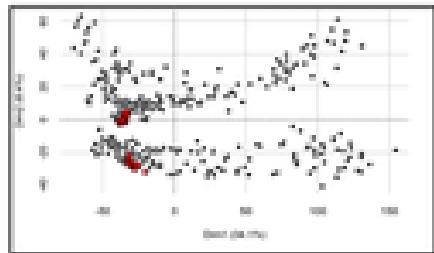


OVERVIEW OF STATISTICAL METHODS

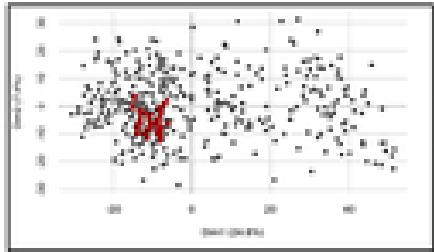
	Visualisation	Regression	Independent 2-group test	Dependent 2-group test	Multiple factors	Time series
Parametric	Plot	Linear models	t-test	Paired t-test	ANOVA Mixed models	Repeated measures
Non-parametric	Rank plot	Rank regr. Kernel LOESS	MW U-test	Wilcoxon signed-rank	Kruskal-Wallis Friedman rank ANOVA	Friedman rank-repeated
Multivariate	PCA	PLS; RF; SVM	PLS-DA; RF; SVM	ML-PLS; ML-RF; ML-SVM	ANOVA decomposition	ANOVA decomposition

VERIFICATION OF QUALITY CONTROL SAMPLES AND OUTLIERS WITH PCA

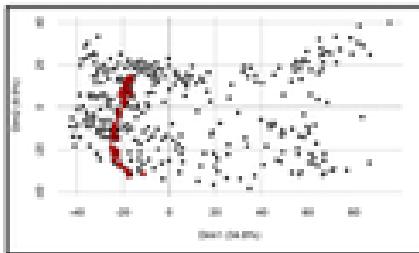
Example of Batch Effect – samples divided into two clouds due to a prompt problem during injections (RAW, not normalized data)



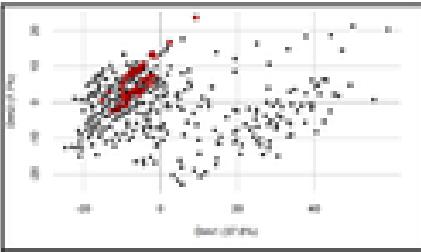
The same sample set normalized by:
1) median of each feature/plate = 1
2) Intensity of creatinine m/z feature



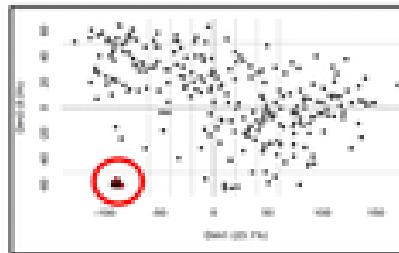
Example of Drift due to loss of signal intensity– Samples and QCs suffered from drop in the signal intensity during injections (RAW, not normalized data)



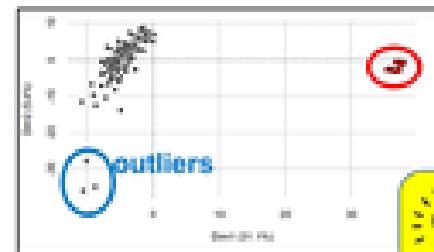
The same sample set normalized by:
1) median of each feature/plate = 1
2) Intensity of creatinine m/z feature



Example of QC prepared from study samples
– a tight QCs cloud is located nearby a wide cloud of real samples (RAW, not normalized data)



Example of QC prepared from commercial biological fluid (i.e. plasma)– a tight QCs cloud is located far from a wider cloud of real samples (RAW, not normalized data). Three samples (outliers) are separated from the rest of samples (blue circle)



Tips & Tricks

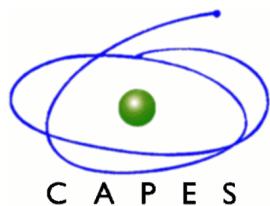
Exemplos



Universidade Federal do Ceará
Departamento de Engenharia Química



UNIVERSITY OF COPENHAGEN



inct
institutos nacionais
de ciência e tecnologia

¹H NMR spectroscopy and chemometrics evaluation of non-thermal processing of orange juice

Amostra: Suco laranja

Processamento: Plasma e O₃

Análise: RMN ¹H

Quimiometria: PCA e quantificação (*q*NMR)

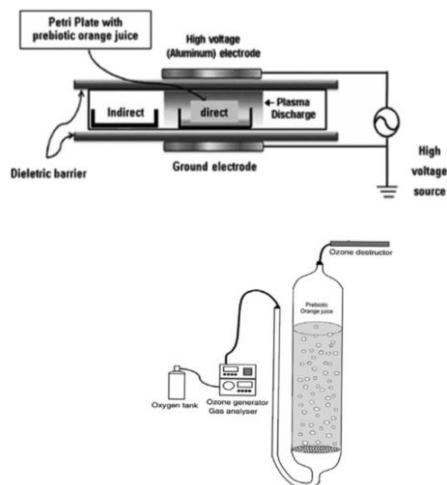
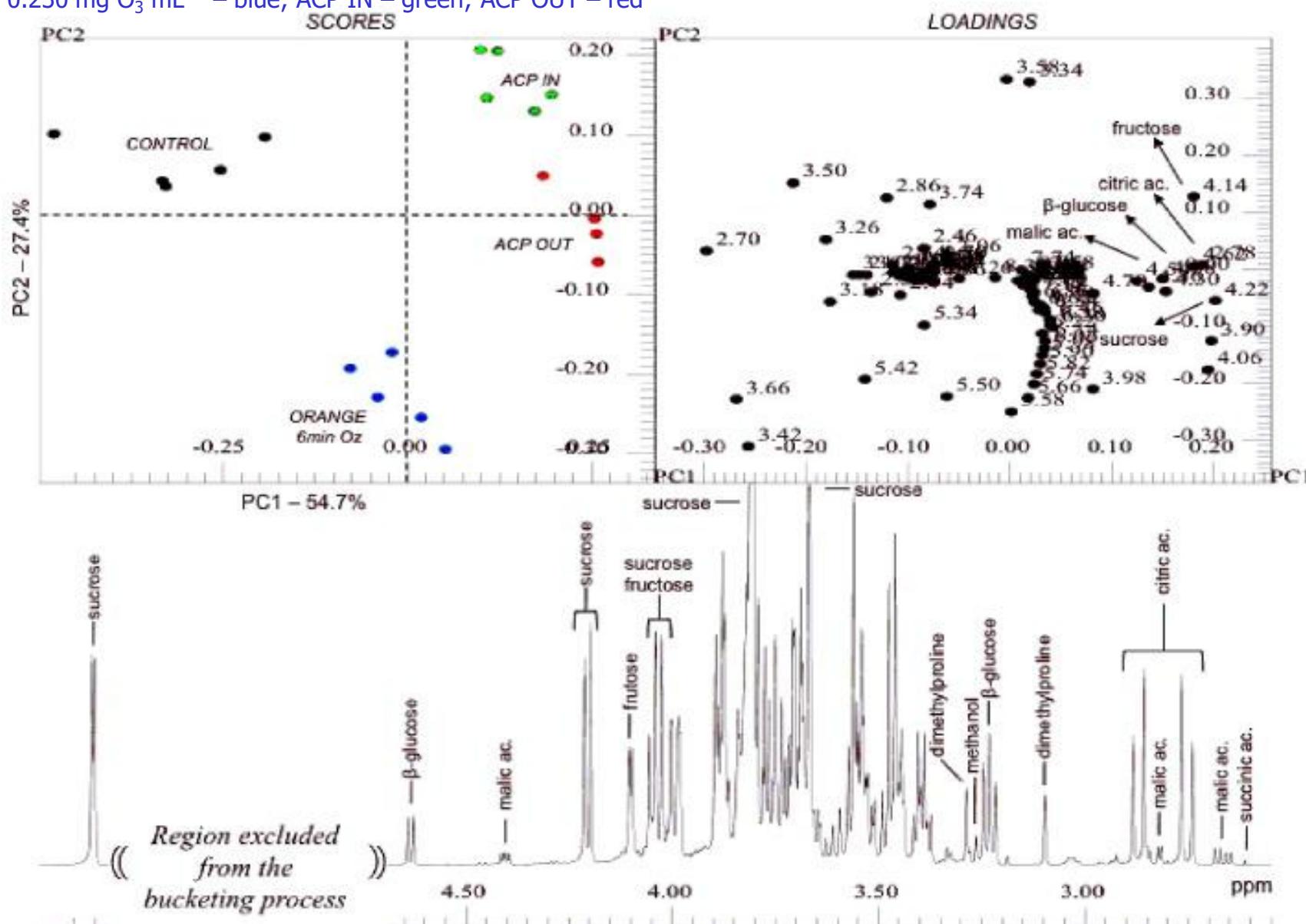
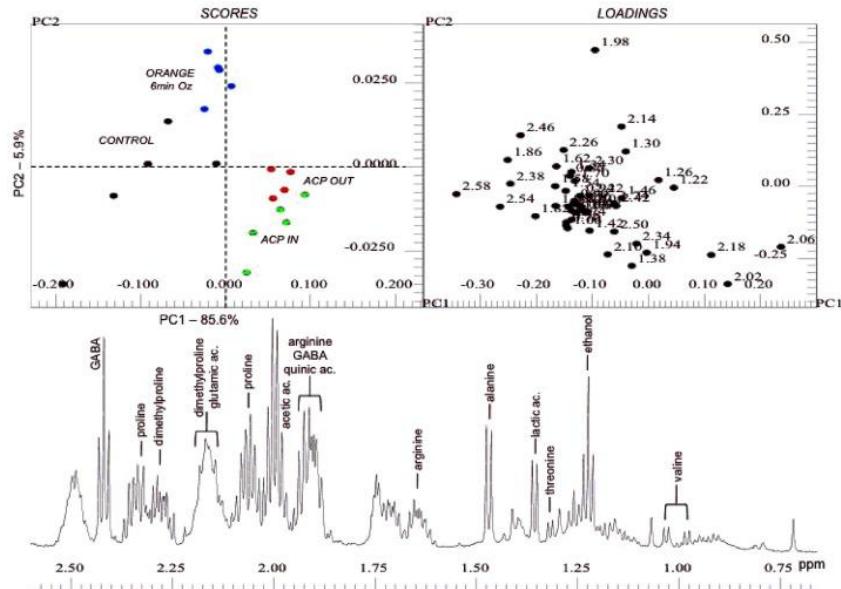


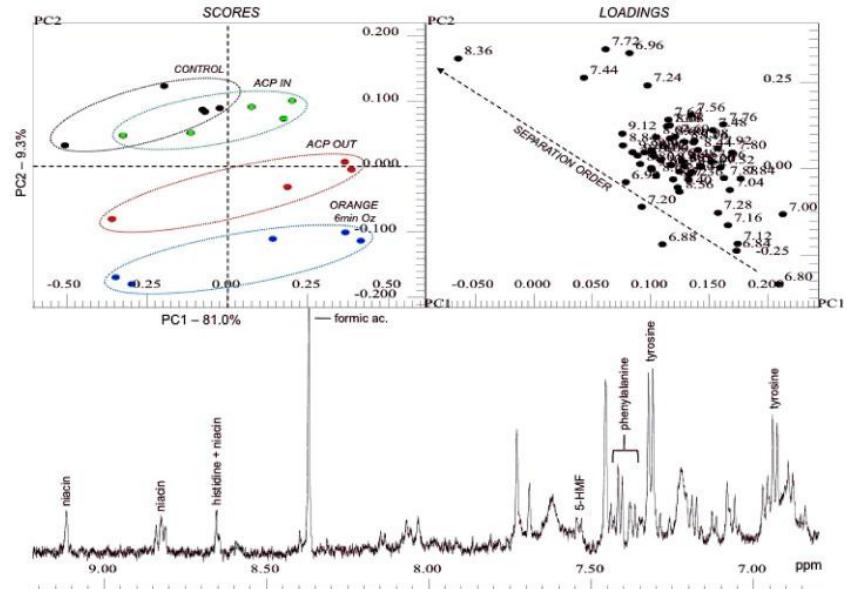
Fig. 1. ^1H NMR spectrum (C), PC1 vs. PC2 scores (left side – A) and loadings (right side – B) coordinate system for the orange juice submitted to different processing: control – black; $0.230 \text{ mg O}_3 \text{ mL}^{-1}$ – blue; ACP IN – green; ACP OUT – red



Região alifáticos



Região aromáticos



- The processes promoted slight variation in concentration of primary metabolites.
- The variations did not result in significant changes in orange juice composition.
- Plasma and ozone are suitable non-thermal alternatives for orange juice processing.

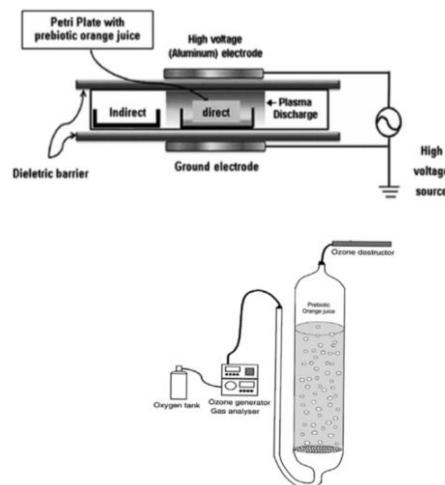
An untargeted chemometric evaluation of plasma and ozone processing effect on volatile compounds in orange juice

Amostra: Suco laranja

Processamento: Plasma e O₃

Análise: GC-MS

Quimiometria: análise hierárquica + PCA



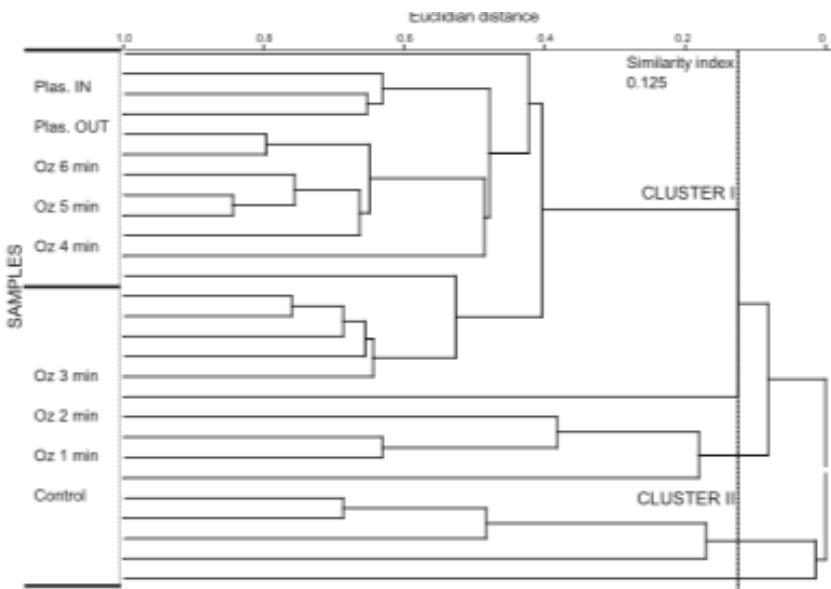


Fig. 2. Dendrogram representing chemical composition similarity relationships among orange juices to whose cluster it belongs: I) plasma IN, plasma OUT, and 4–6 min ozone; II) control, and 1–3 min ozone.

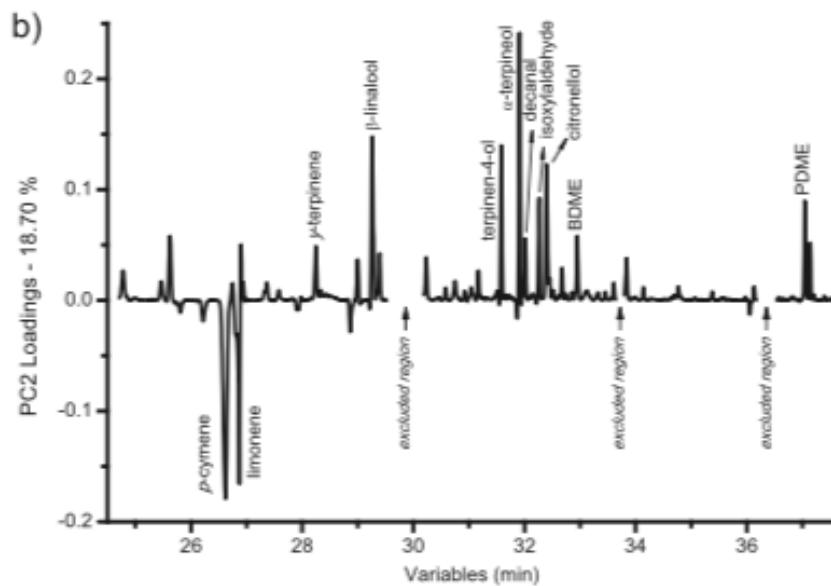
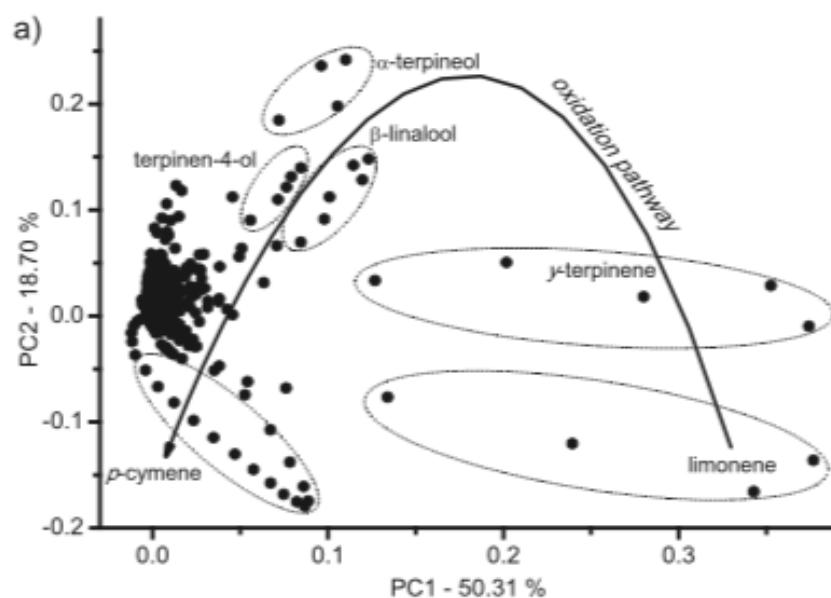
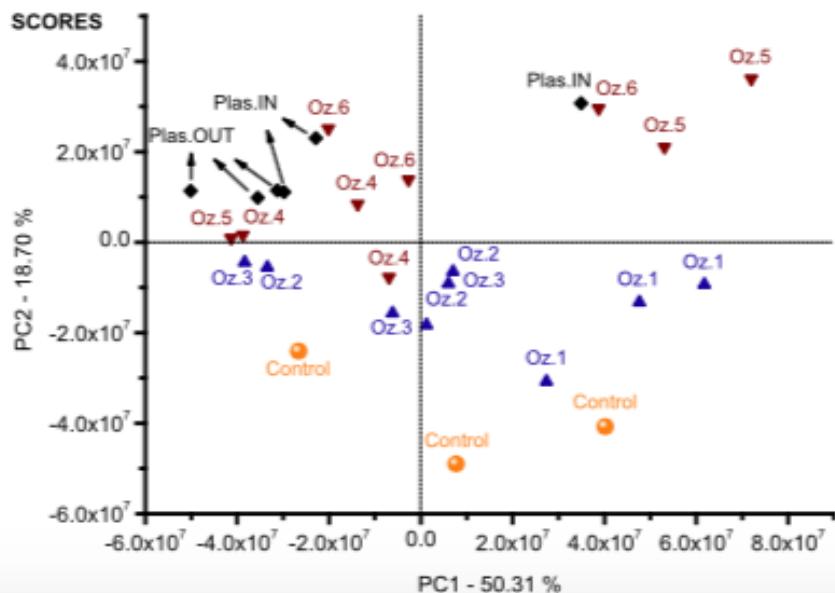


Fig. 4. PC1 vs PC2 loadings plotted in two dimensions (a); and PC2 plotted in line (b). The excluded regions reflect interfering signals removed for chemometrics.

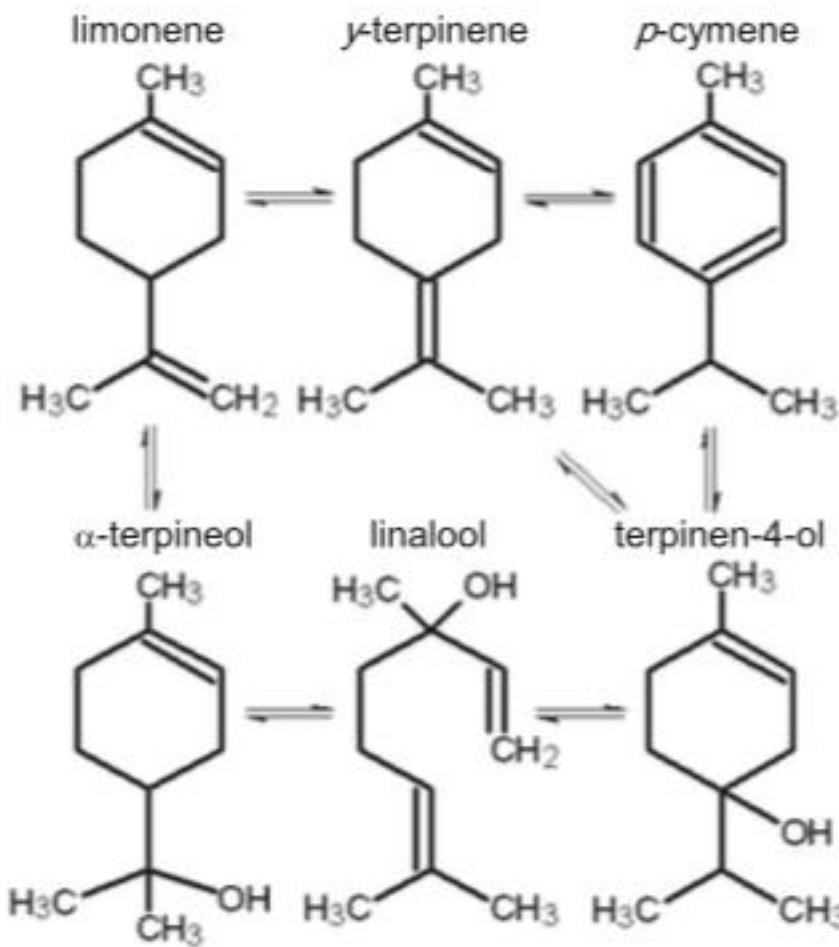


Fig. 5. Proposed oxidation, hydrolysis and reduction of terpenes under plasma and ozone processing of orange juice.

NMR spectroscopy and chemometrics to evaluate different processing of coconut water

Amostra: água de coco

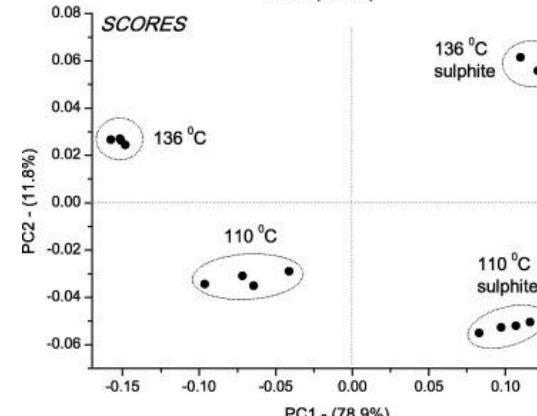
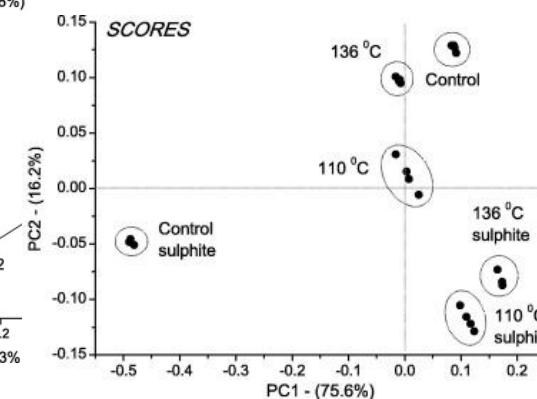
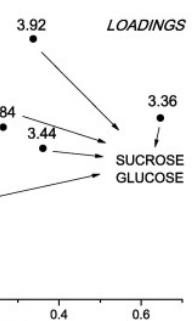
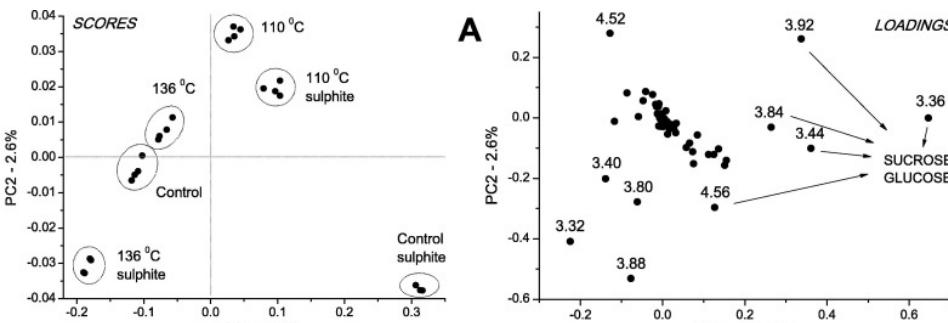
Processamento: térmico (UHT) + com/sem SO₂

Análise: RMN ¹H

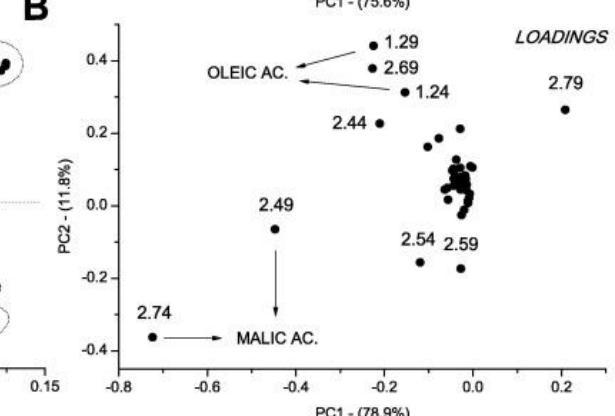
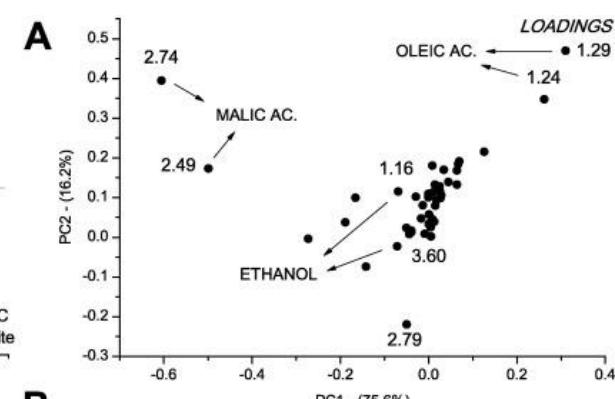
Quimiometria: PCA e quantificação (*q*NMR)



Completo



Alifático



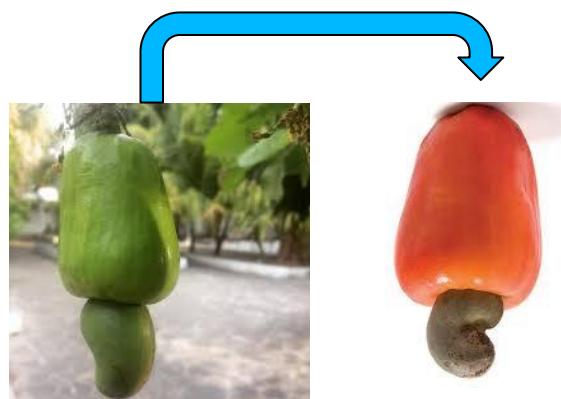
UPLC-qTOF-MS/MS-based phenolic profile and their biosynthetic enzyme activity used to discriminate between cashew apple (*Anacardium occidentale* L.) maturation stages

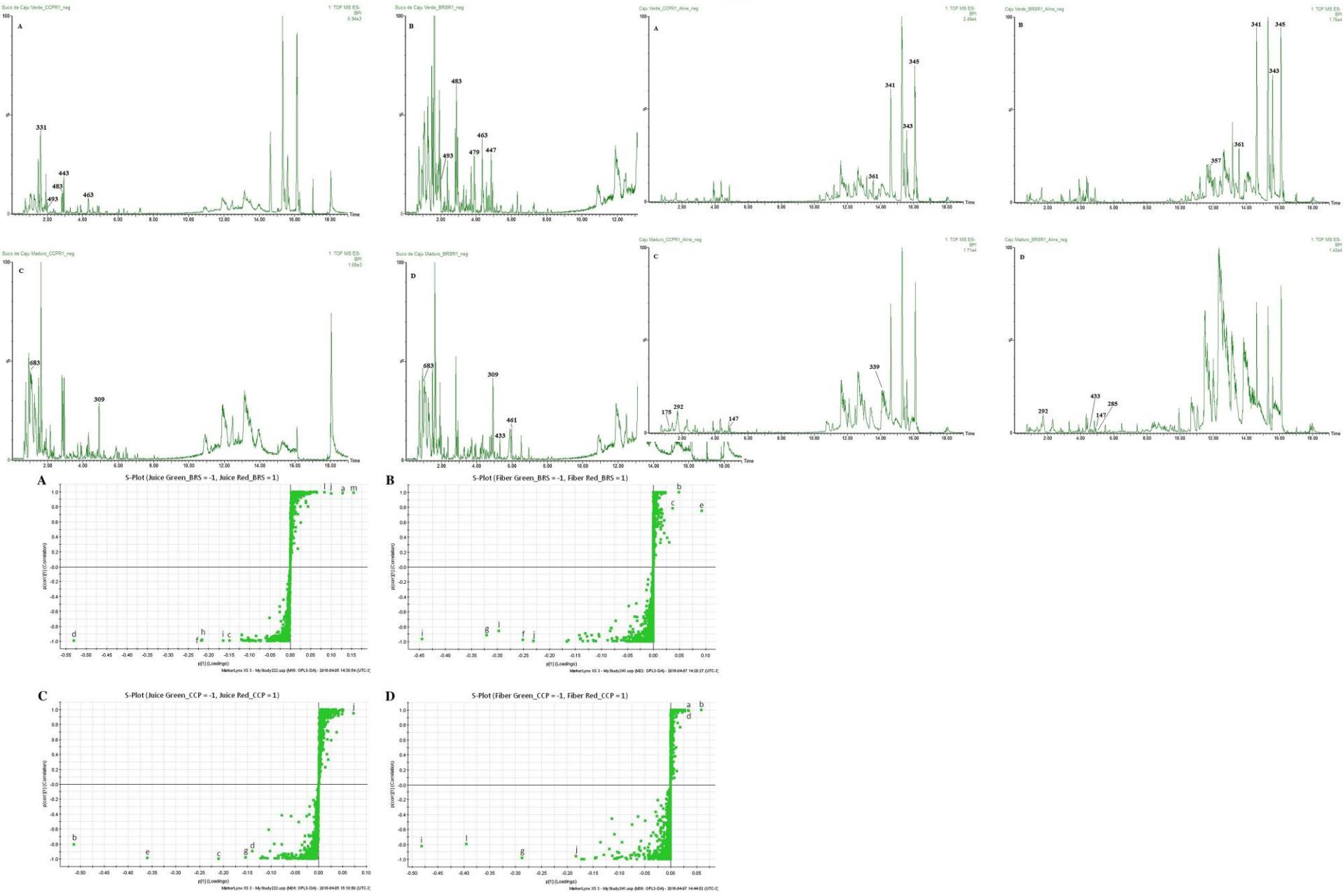
Amostra: caju

Processamento: diferentes estádios de maturação

Análise: LC-MS

Quimiometria: PCA e OPLS-DA





cinnamoyl glucoside may be used as a chemical marker stage 7-ripe juice samples, while monogalloyl diglucoside (c) and digalloyl glucoside (d) as chemical markers of stage 2-green juice samples from both cashew clones. Meanwhile, four compounds could be used as markers for stage 2-green fiber of both clones, anacardic acids (i, j and l) and GA₁₉ (g) which have low solubility in aqueous solutions due to their chemical natures as phenolic lipids and a diterpenoid acid, respectively.

The evaluation of enzymes of phenolic biosynthetic pathway in juice samples showed that PAL activity decreased significantly during the development of cashew apple clones, despite of which, it was much higher in ripe CCP 76 cashew apple. UGT activity differed between clones, however its main product, cinnamoyl glucoside was the most characteristic compound, thus a chemical marker of ripe juice samples from both clones. FLS showed the highest specific activity in both cashew clones and its product, flavonols (glycosylated probably due to UGT action), were identified in cashew apple at immature and ripe stages. LAR activity was not detected during the development of cashew apple from clones BRS 189 and CCP 76 agreeing with previous publications [12], [13] which found no LAR substrates, flavan-3-ols or proanthocyanindins in cashew apples.

Chemical profiling of guarana seeds (*Paullinia cupana*) from different geographical origins using UPLC-QTOF-MS combined with chemometrics

¹H quantitative nuclear magnetic resonance and principal component analysis as tool for discrimination of guarana seeds from different geographic regions of Brazil

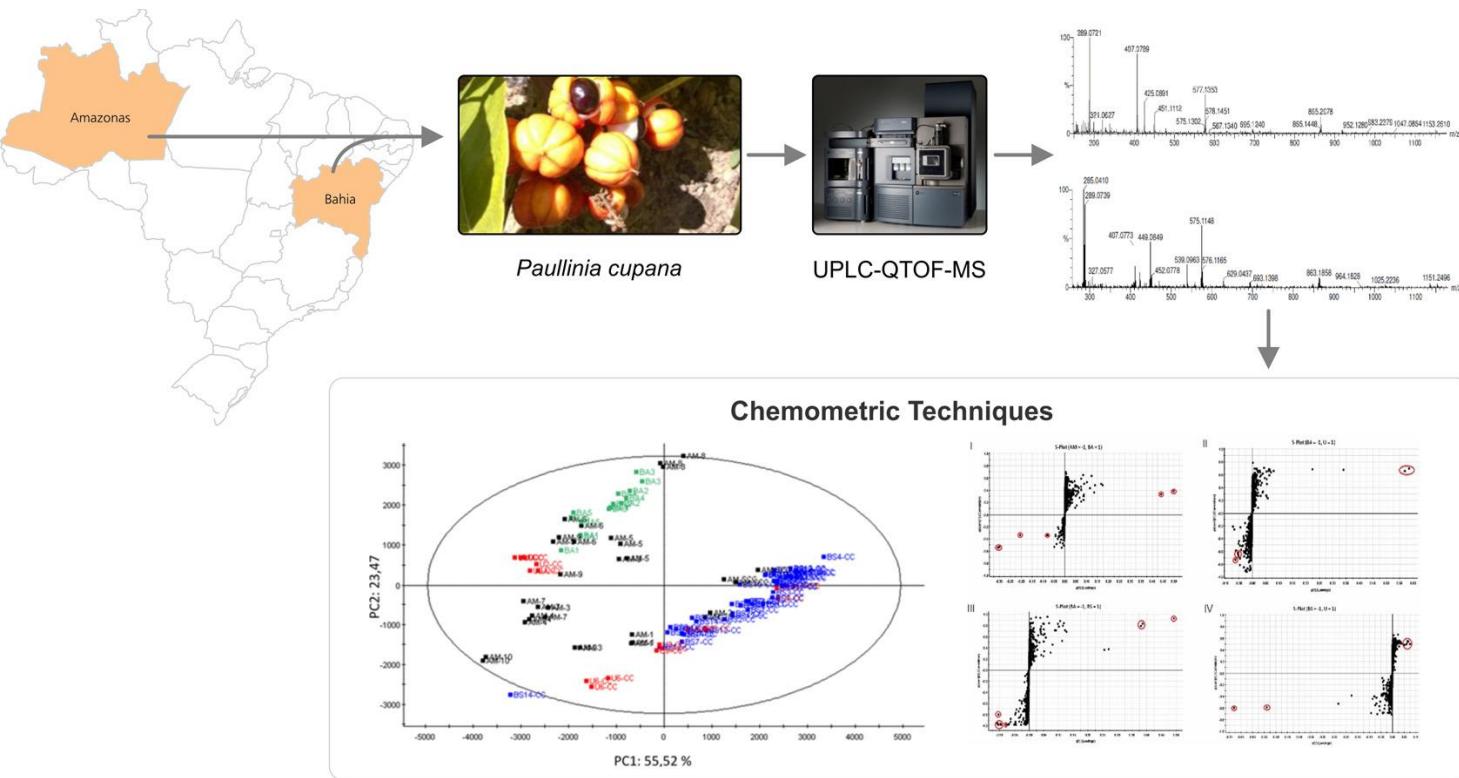
Amostra: guaraná

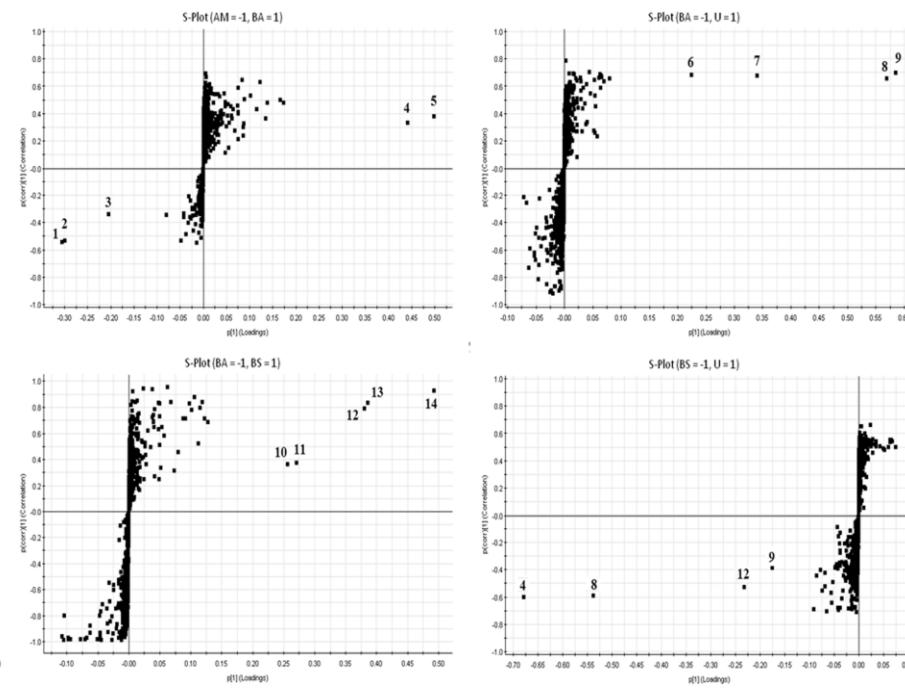
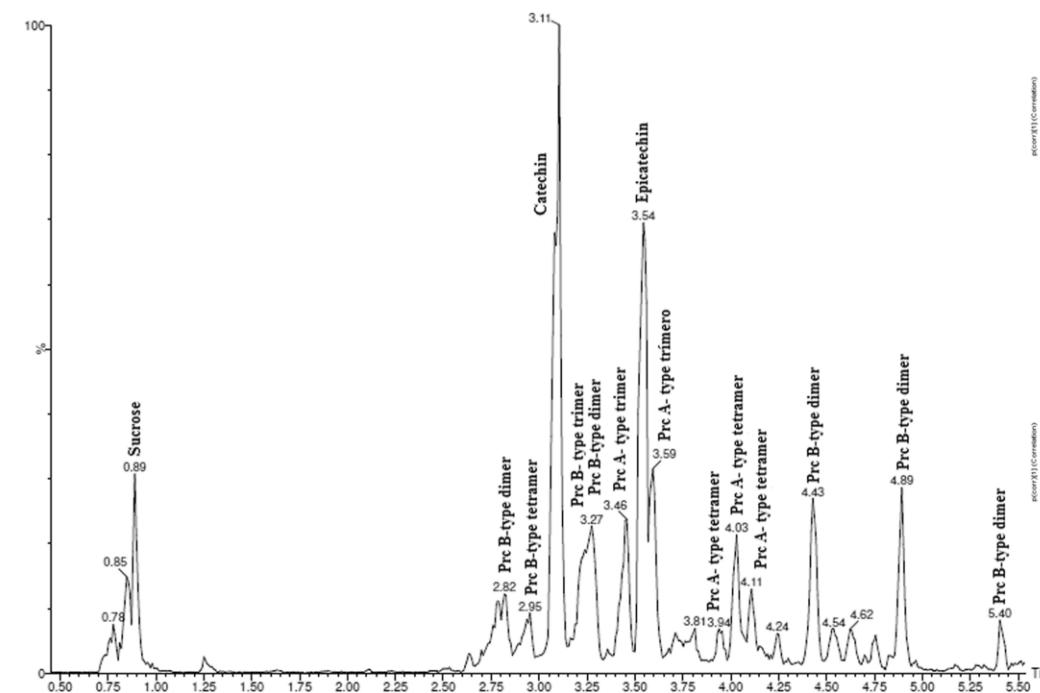
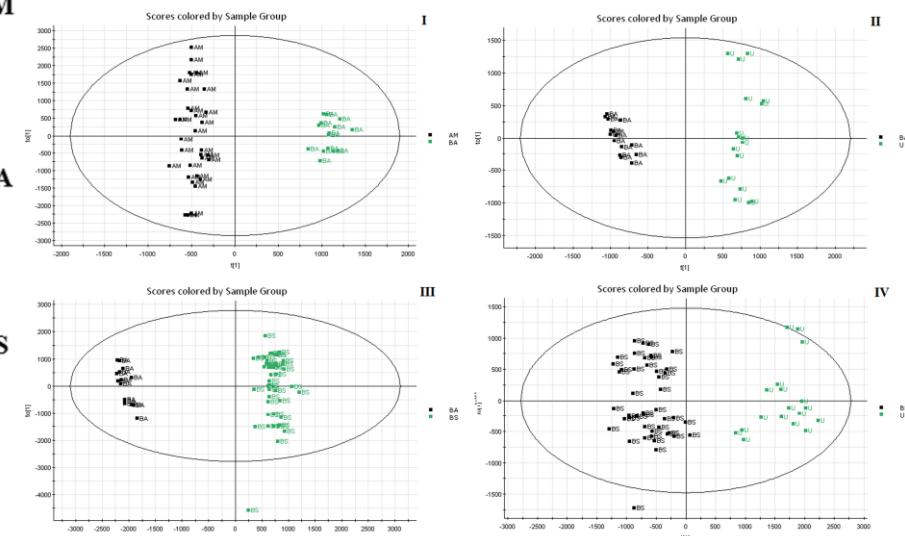
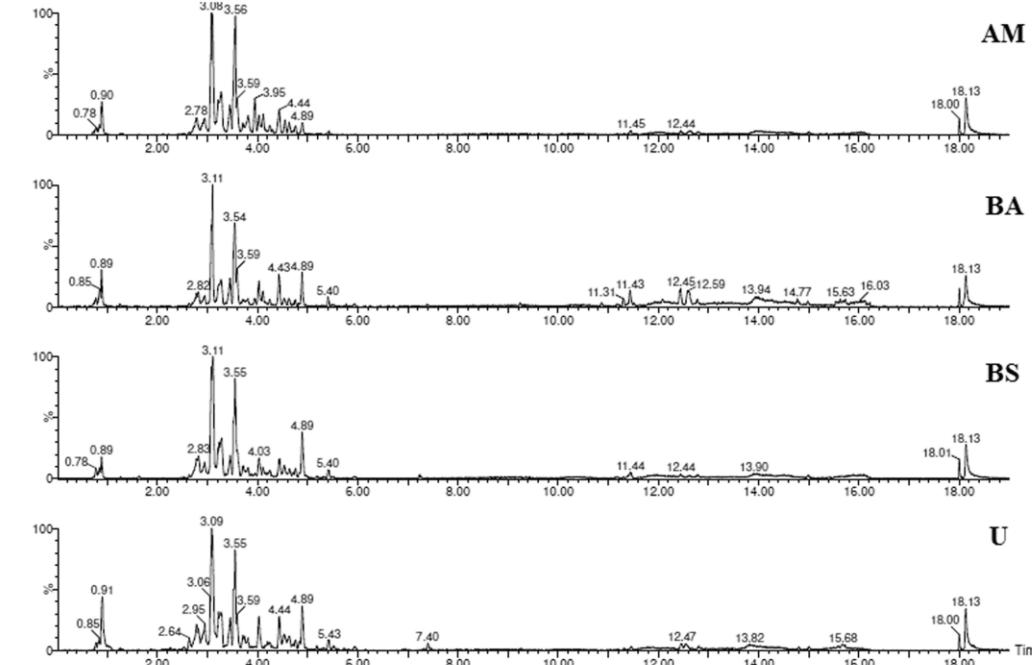
Processamento: diferentes origens

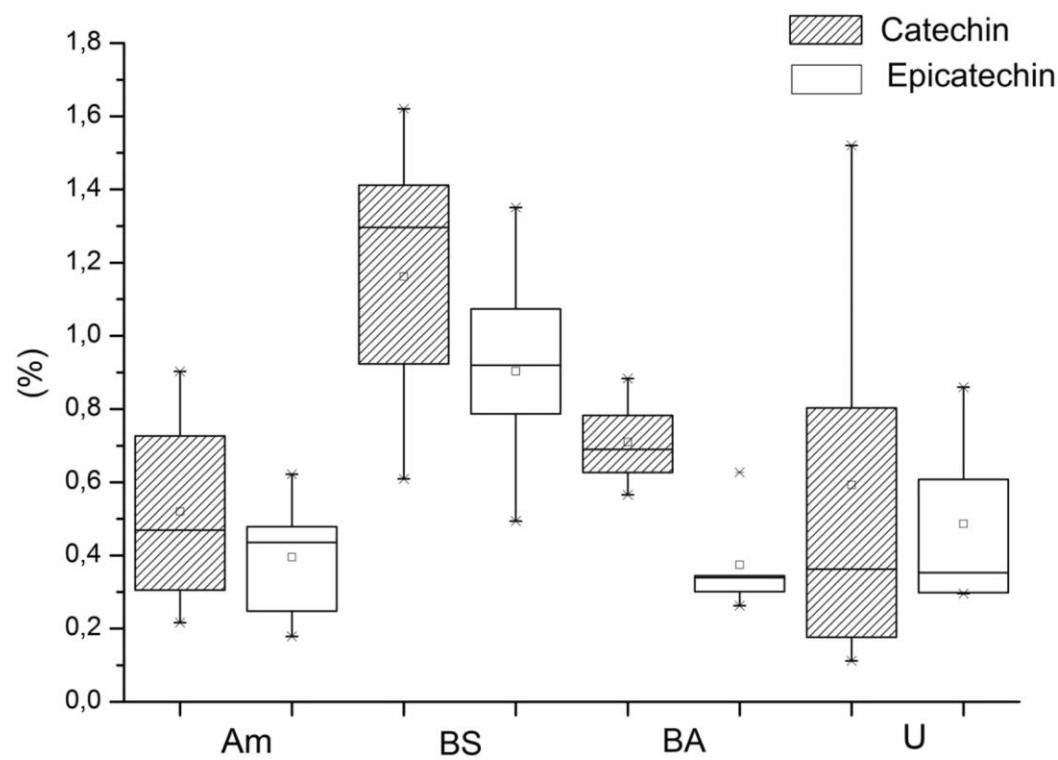
Análise: LC-MS e RMN ¹H

Quimiometria: PCA e quantificação









Genotype evaluation of cowpea seeds (*Vigna unguiculata*) using ^1H qNMR combined with exploratory tools and solid-state NMR

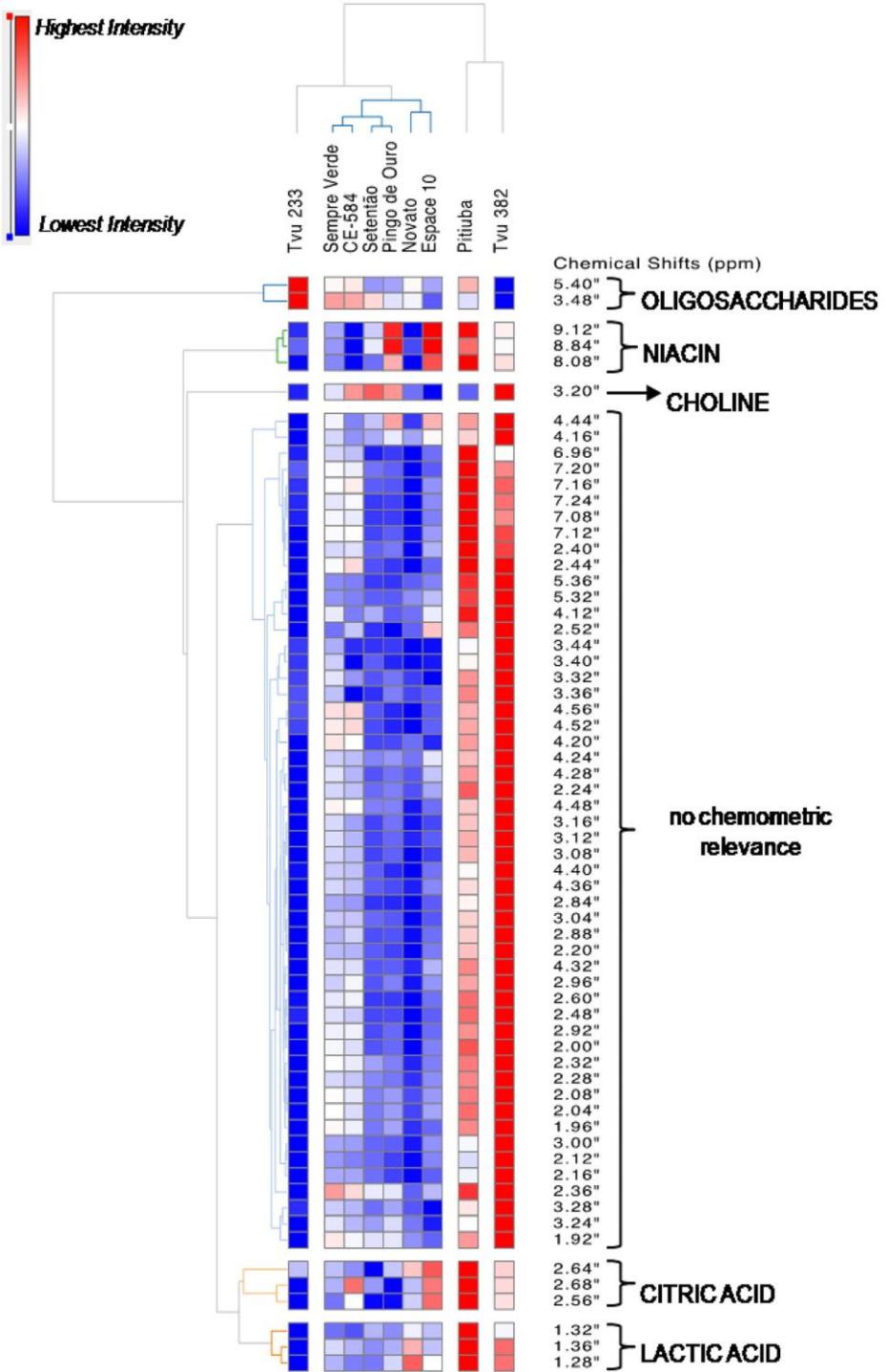
Amostra: feijão caupi

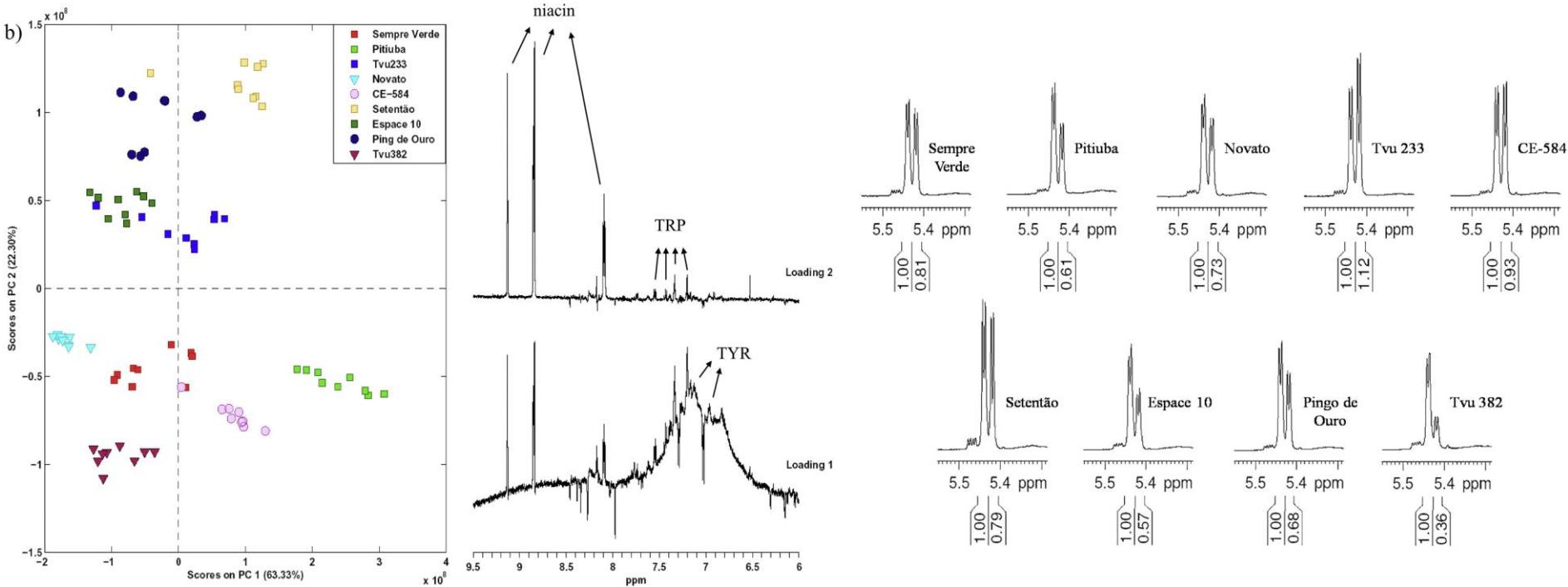
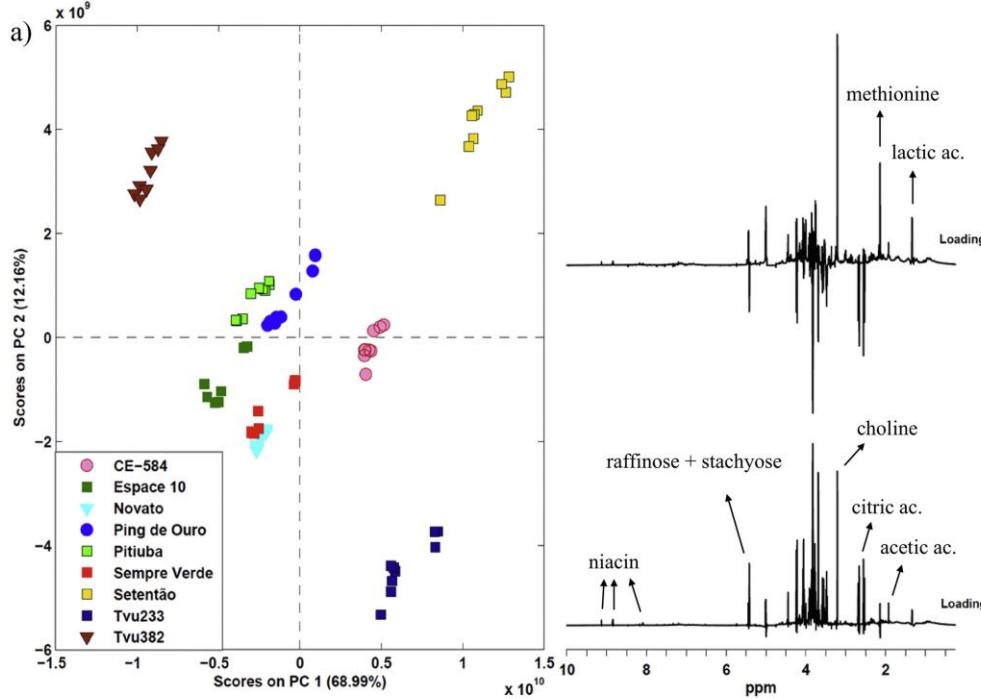
Processamento: (9 genótipos)

Análise: RMN ^1H

Quimiometria: PCA e quantificação (qNMR)







Tracking thermal degradation on passion fruit juice through Nuclear Magnetic Resonance and chemometrics

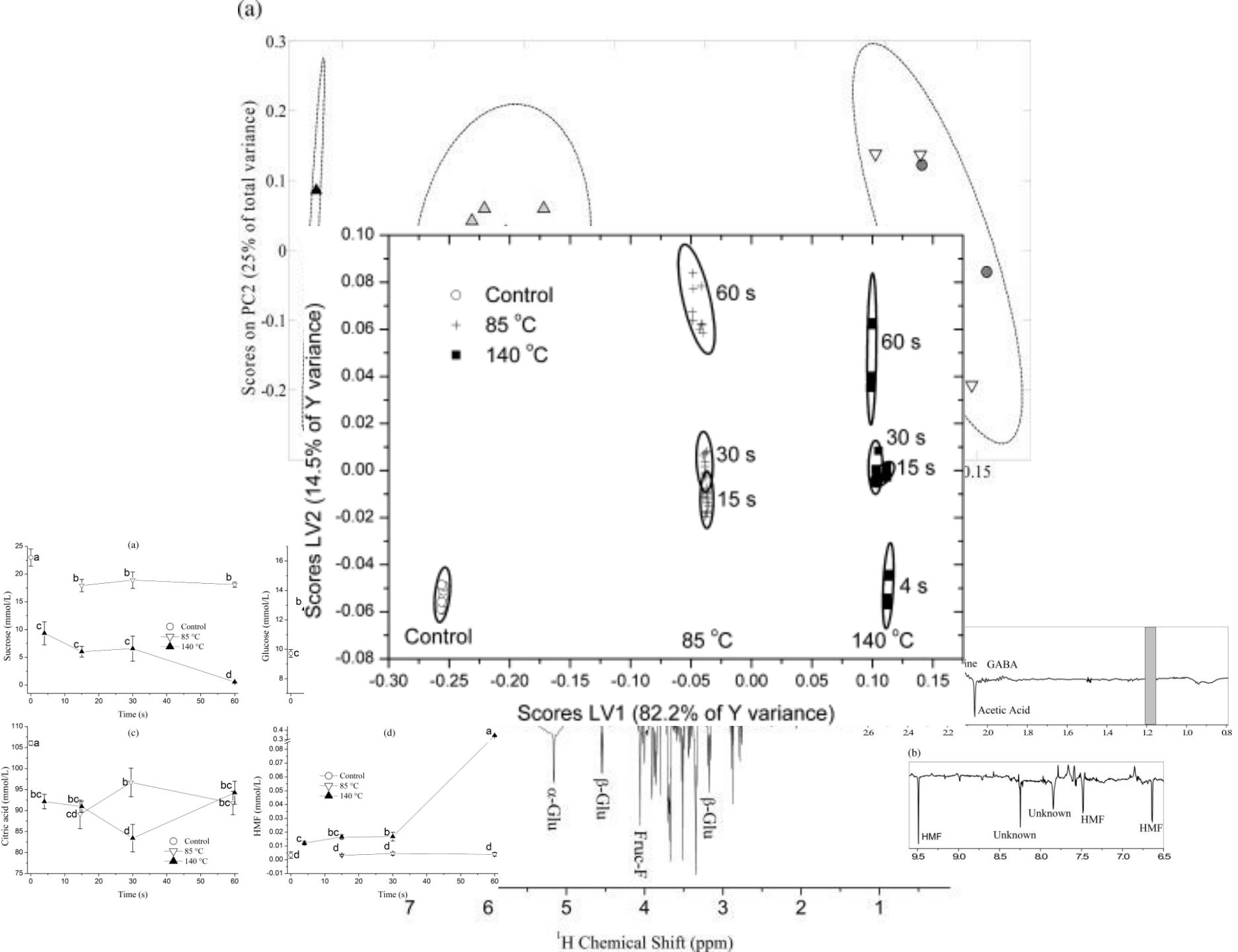
Amostra: suco maracujá

Processamento: térmico UHT e HTST

Análise: RMN ^1H

Quimiometria: PCA e quantificação (q NMR)





Chemometric evaluation of the volatile profile of probiotic melon and probiotic cashew juice

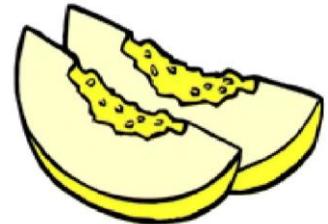
Amostra: suco melão e caju

Processamento: probiotico + térmico UHT e HTST

Análise: GC-MS

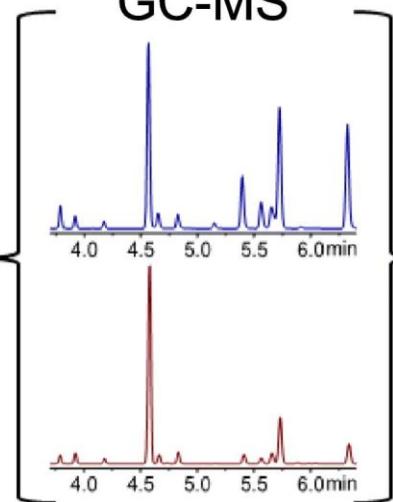
Quimiometria: PCA e quantificação (*q*NMR)



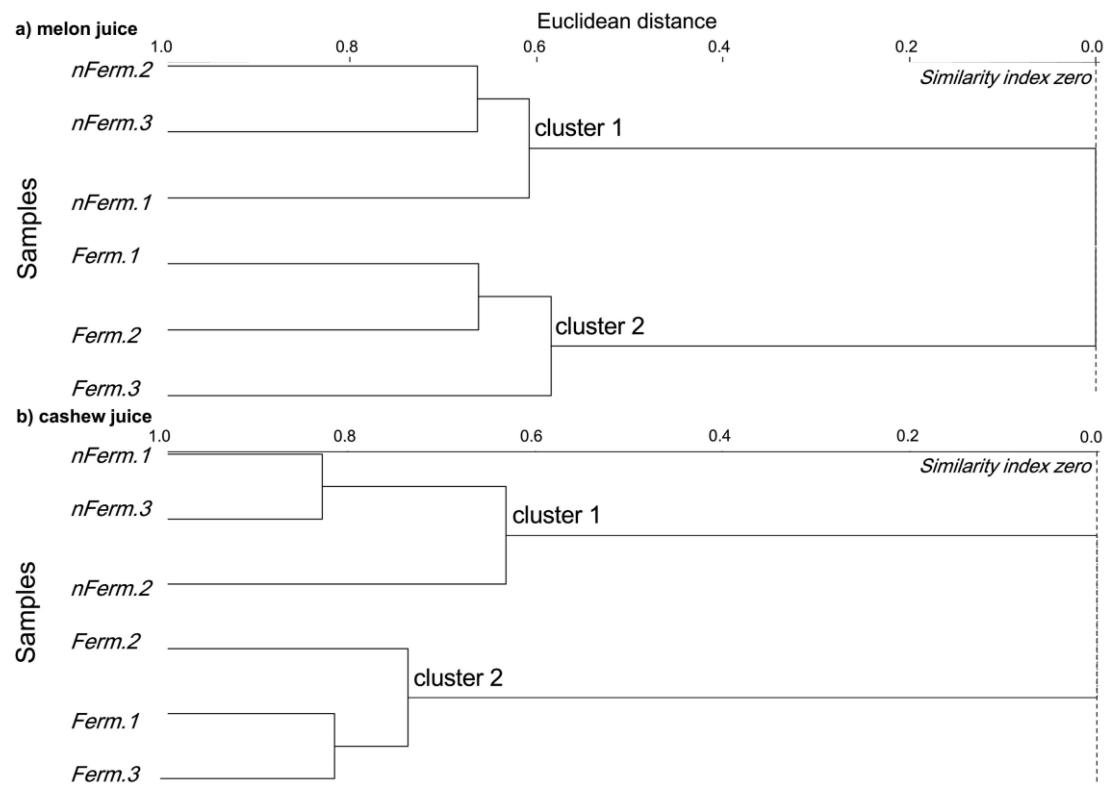
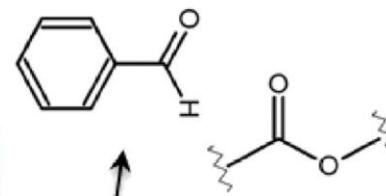


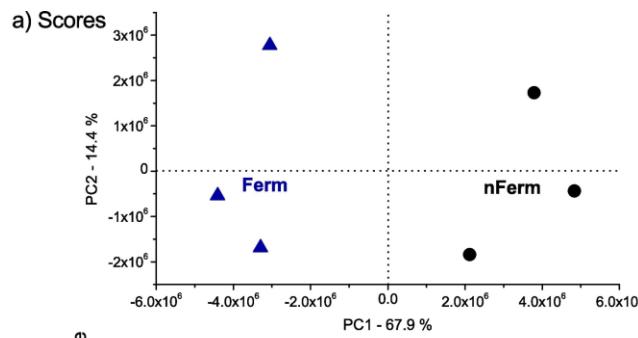
GC-MS

Lactobacillus casei

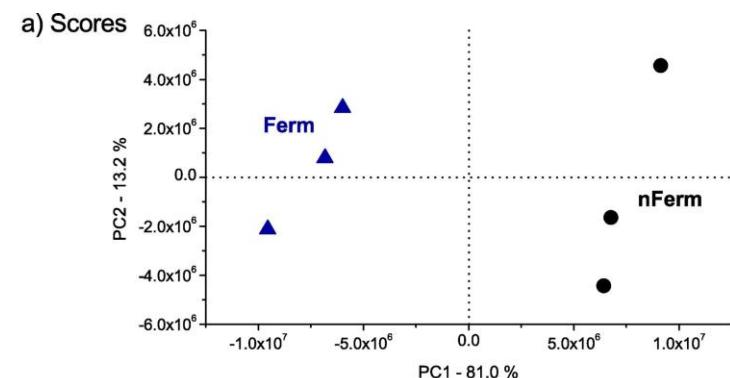
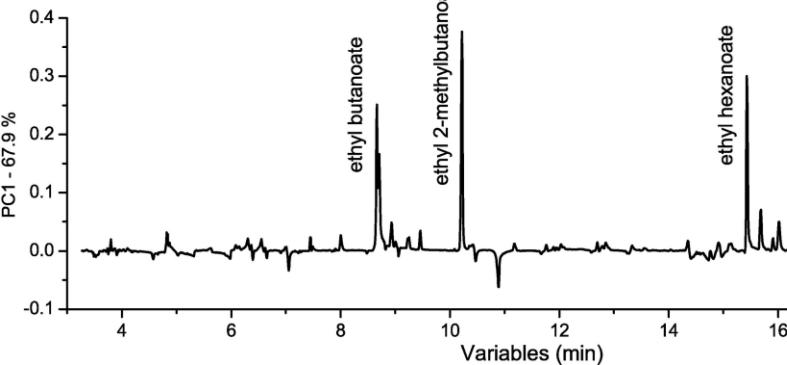


chemometrics

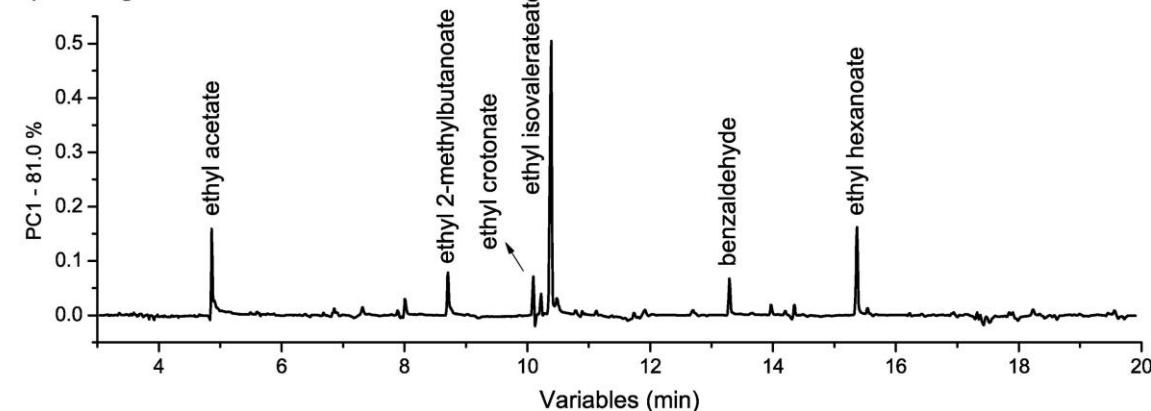


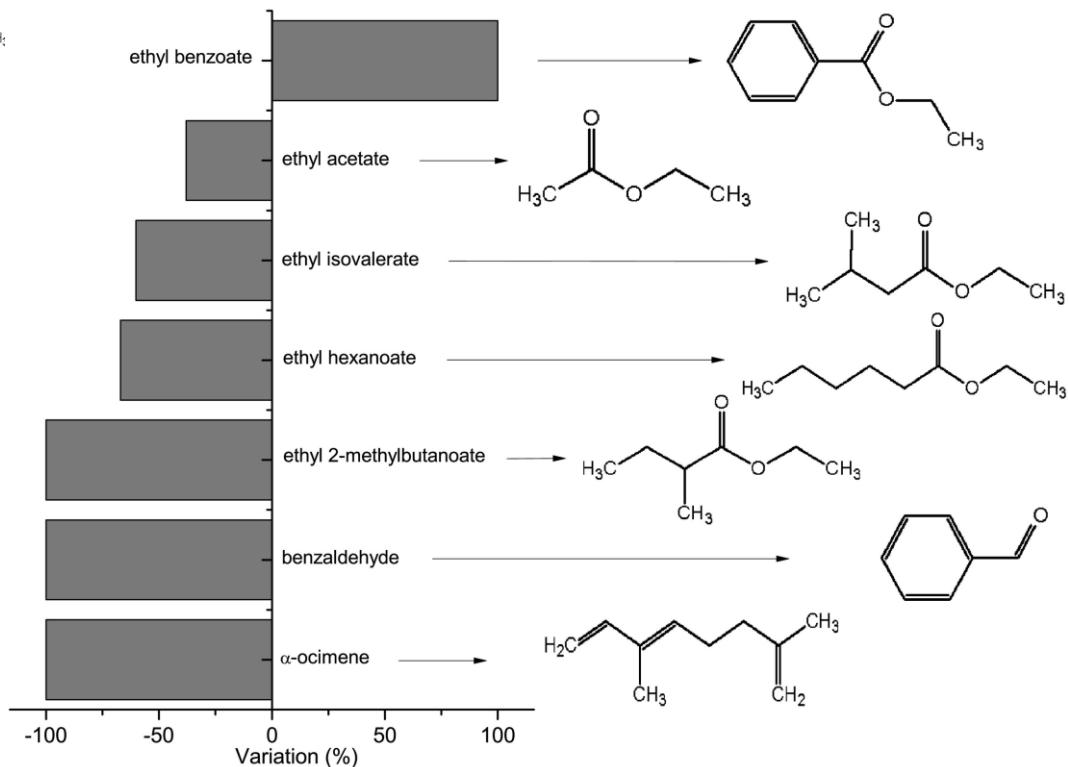
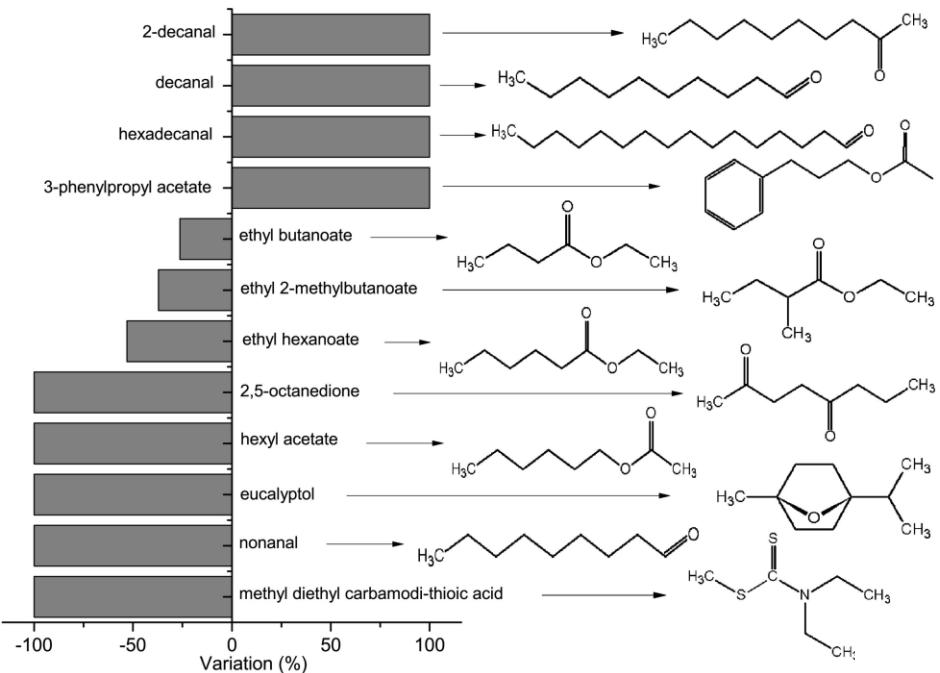


b) Loadings



b) Loadings





Evaluation of thermal and non-thermal processing effect on non-prebiotic and prebiotic acerola juices using ^1H qNMR and GC-MS coupled to chemometrics

Amostra: suco acerola

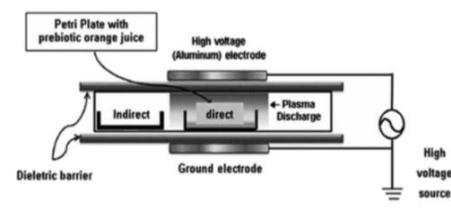
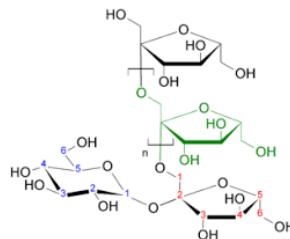
Processamento: prébiotico + térmico UHT e HTST + US+ Plasma

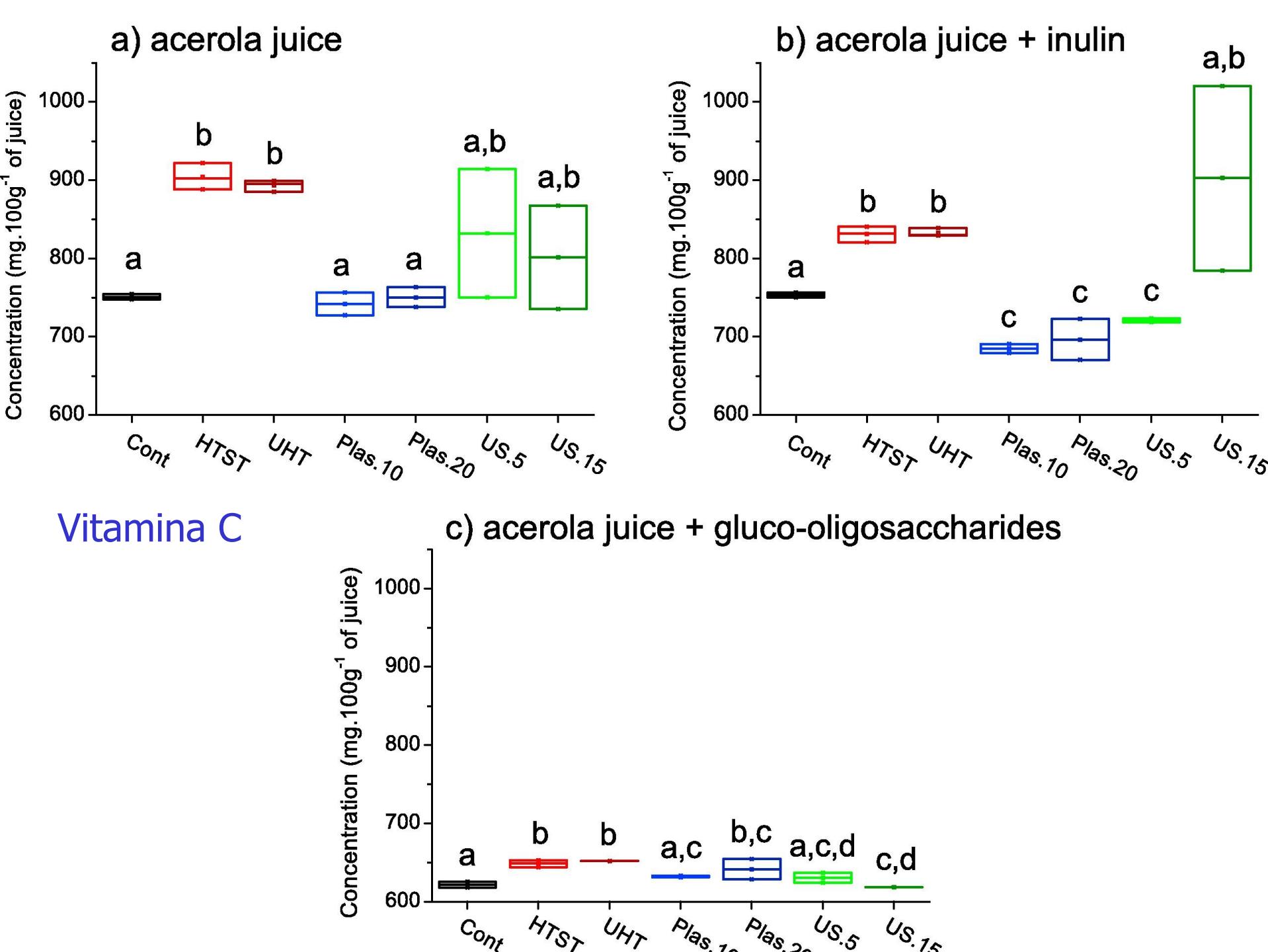
Análise: RMN ^1H + GC-MS

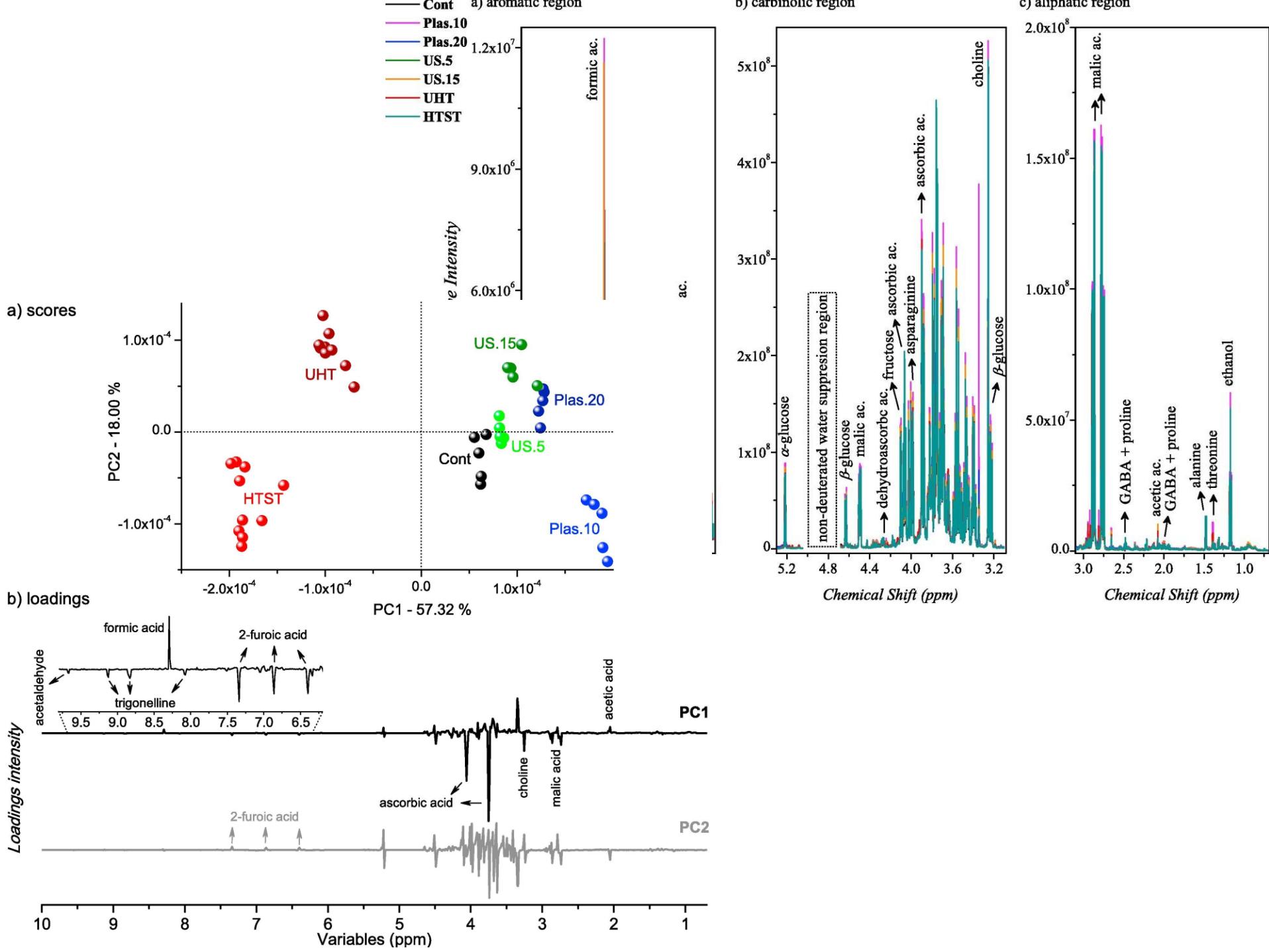
Quimiometria: PCA e quantificação (qNMR)

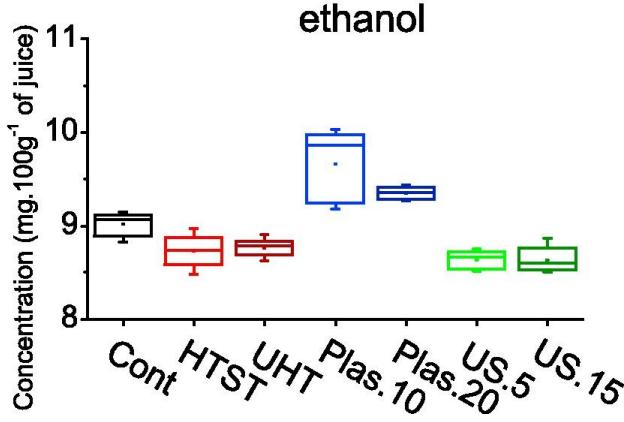
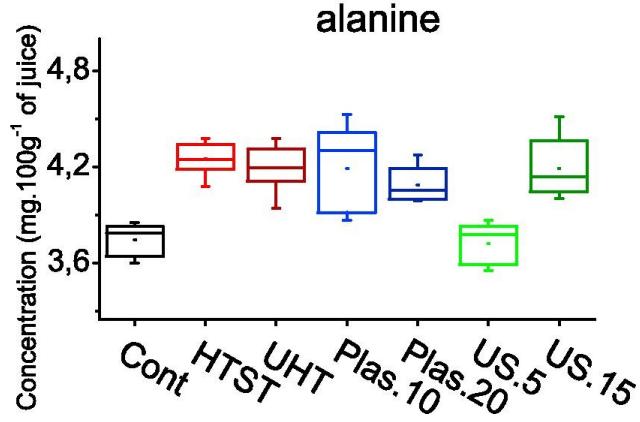
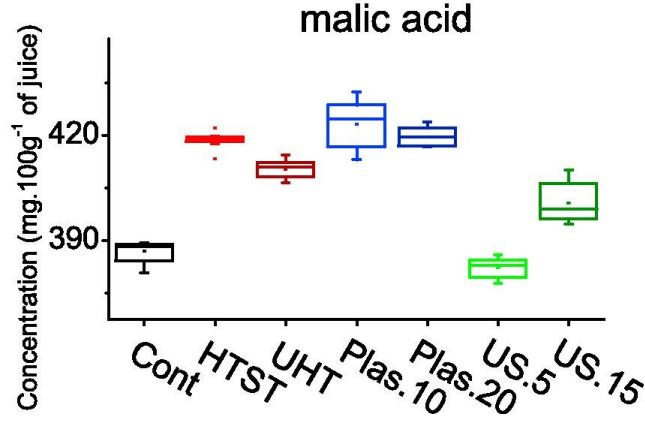
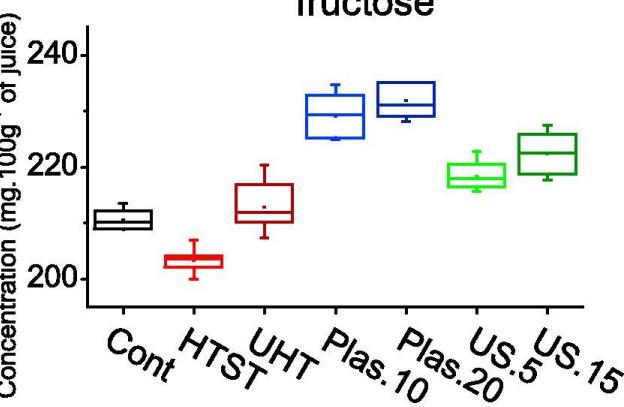
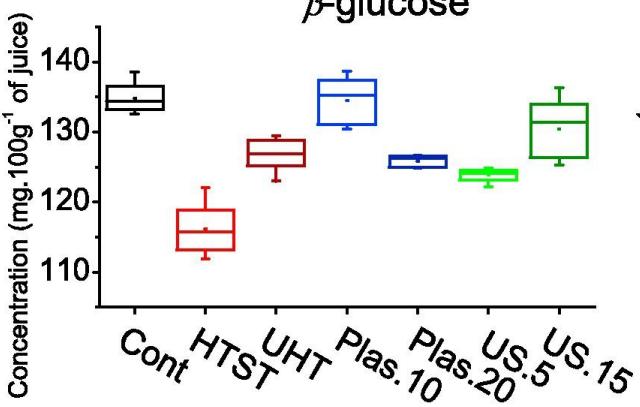
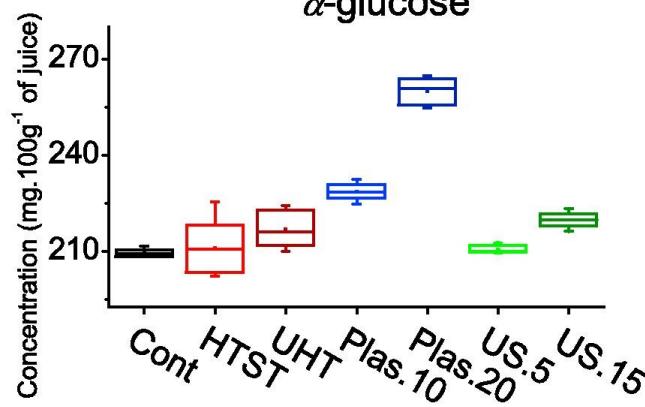
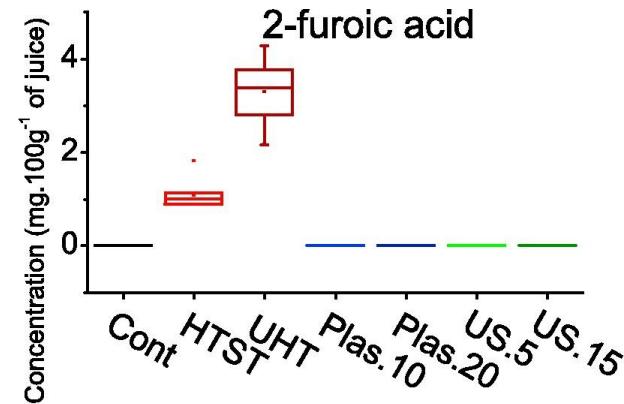
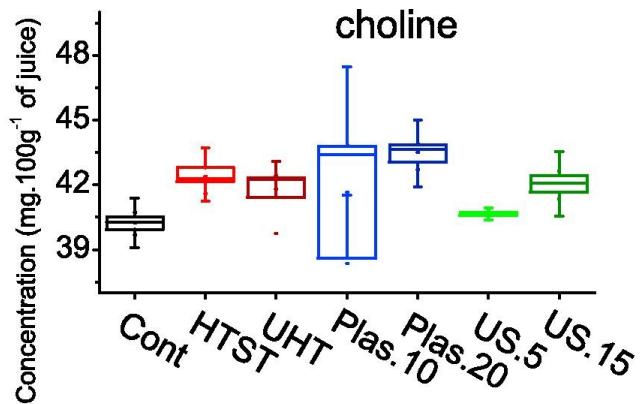
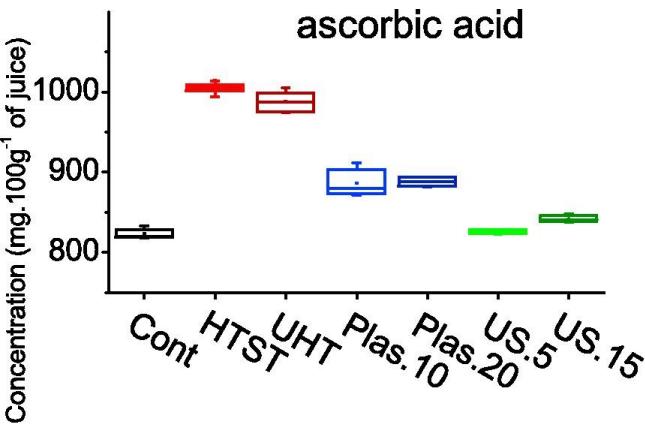


+



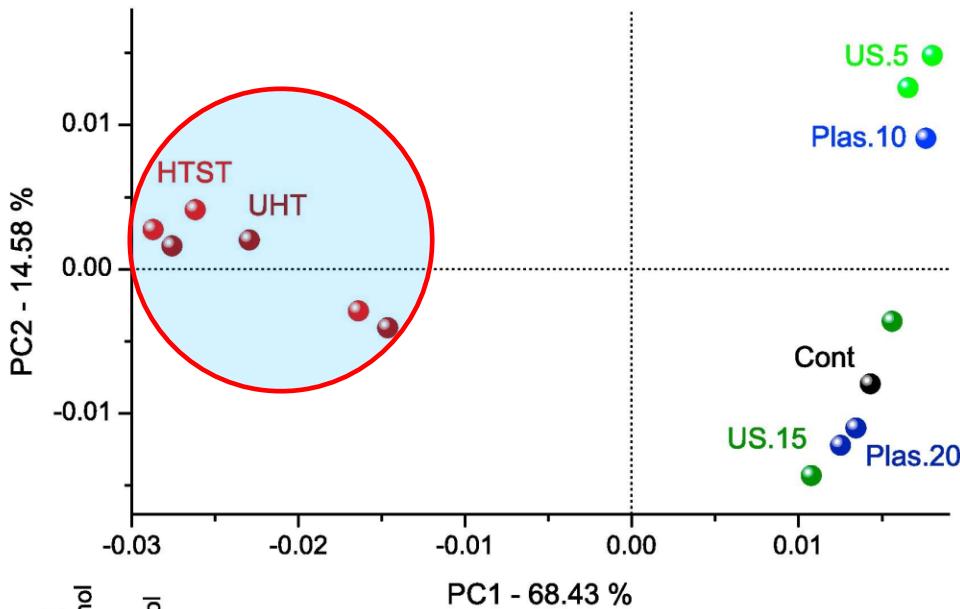






voláteis

a) scores



b) loadings

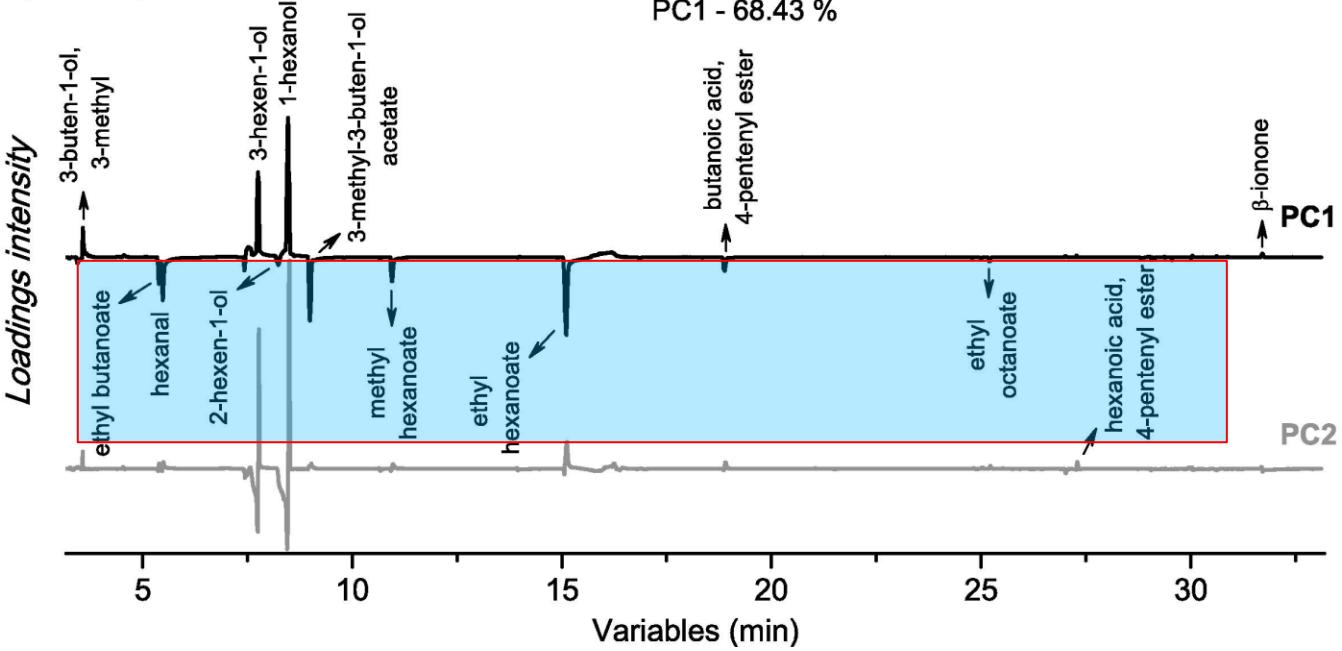


Table 1

Structures of the relevant volatile flavor components detected in acerola juices according to PCA, with respective retention times (RT), experimental and reference retention index (RI), major m/z peak, and percentage (%) of match.

RT (min)	Compound name	Structure	RI ^{exp.}	RI ^{refer.}	Major m/z	Match (%)
3.577	3-methyl 3-buten-1-ol		727	723 ⁱ	41	94
5.384	hexanal		801	801 ⁱ	44	86
5.479	ethyl butanoate		804	802 ⁱ	43	92
7.706	3-hexen-1-ol		856	850 ⁱ	41	97
8.021	2-hexen-1-ol		864	859 ⁱ	57	95
8.431	1-hexanol		873	871 ⁱ	56	94
8.972	3-methyl-3-buten-1-ol, acetate		886	880 ⁱ	43	95
10.957	methyl hexanoate		926	921 ⁱ	74	94
15.086	ethyl hexanoate		1001	997 ⁱ	88	95
18.897	butanoic acid, 4-pentenyl ester		1067	1064 ⁱ	68	84
25.208	ethyl octanoate		1199	1196 ⁱ	94	88
27.310	hexanoic acid, 3-methyl-2-butenyl ester		1268	1267 ⁱ	88	68
31.670	β -ionone		1490	1487 ⁱ	177	92

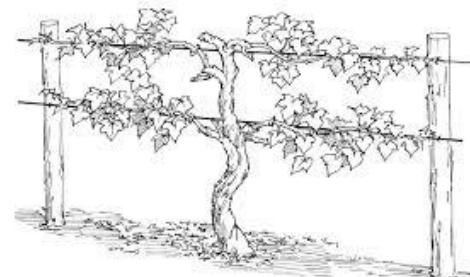
¹H NMR and LC-MS-based metabolomic approach for evaluation of the seasonality and viticultural practices in wines from São Francisco River Valley, a Brazilian semi-arid region

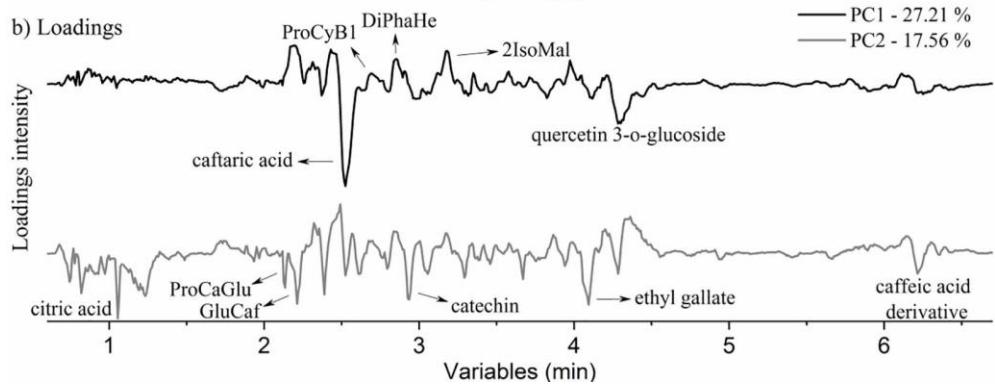
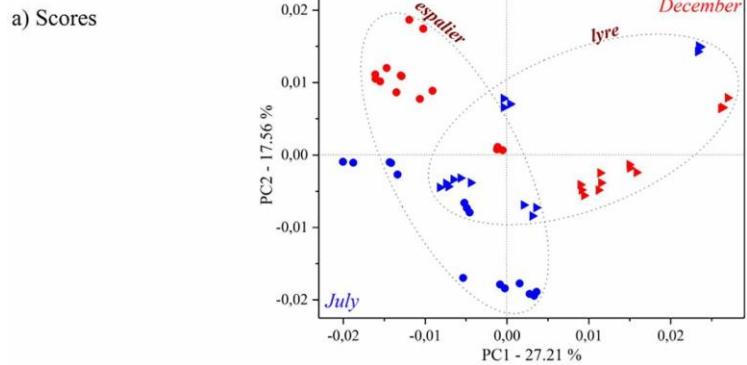
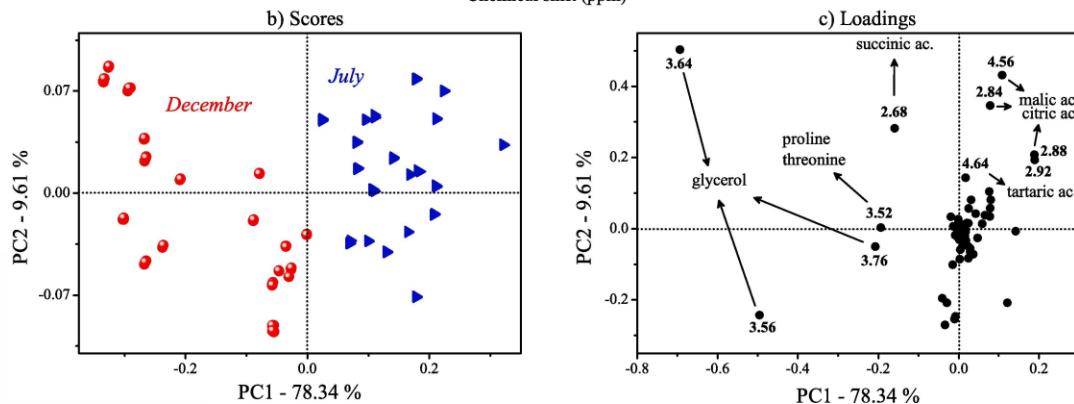
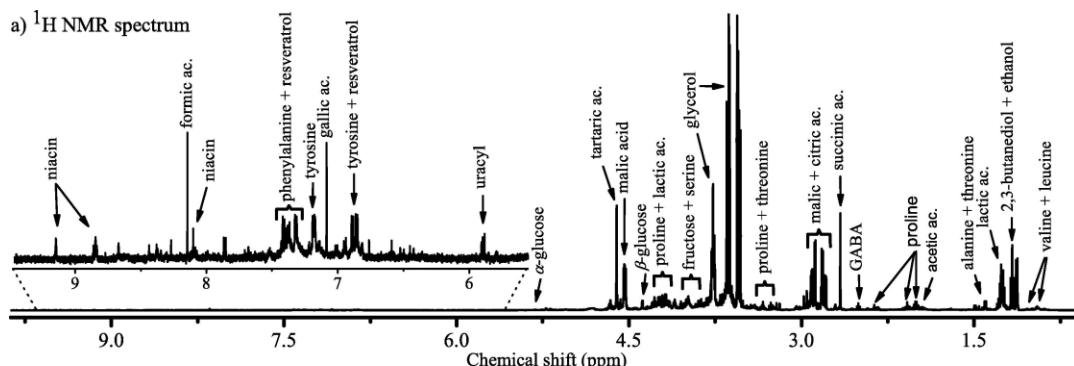
Amostra: vinho

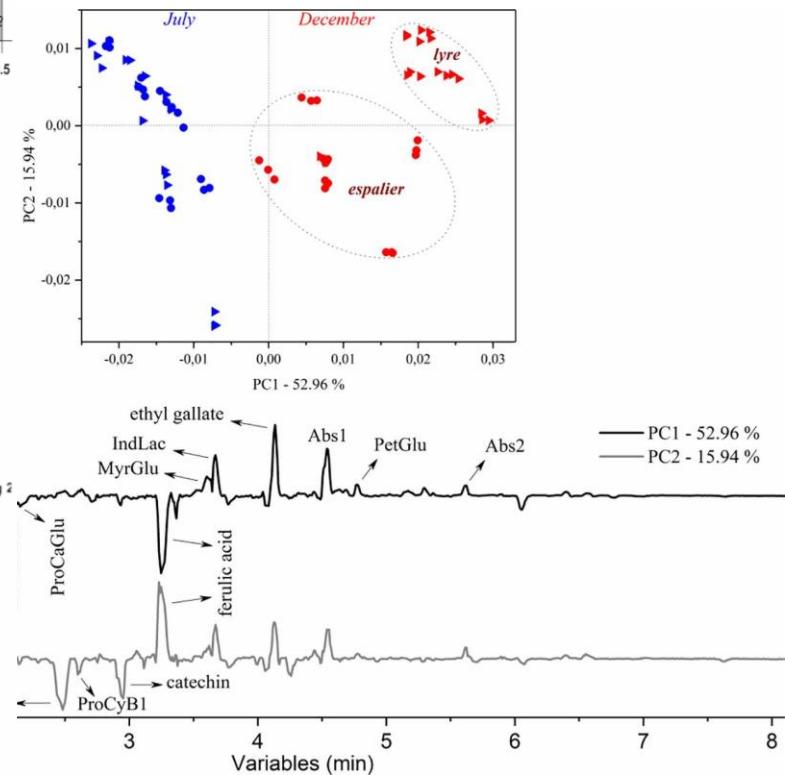
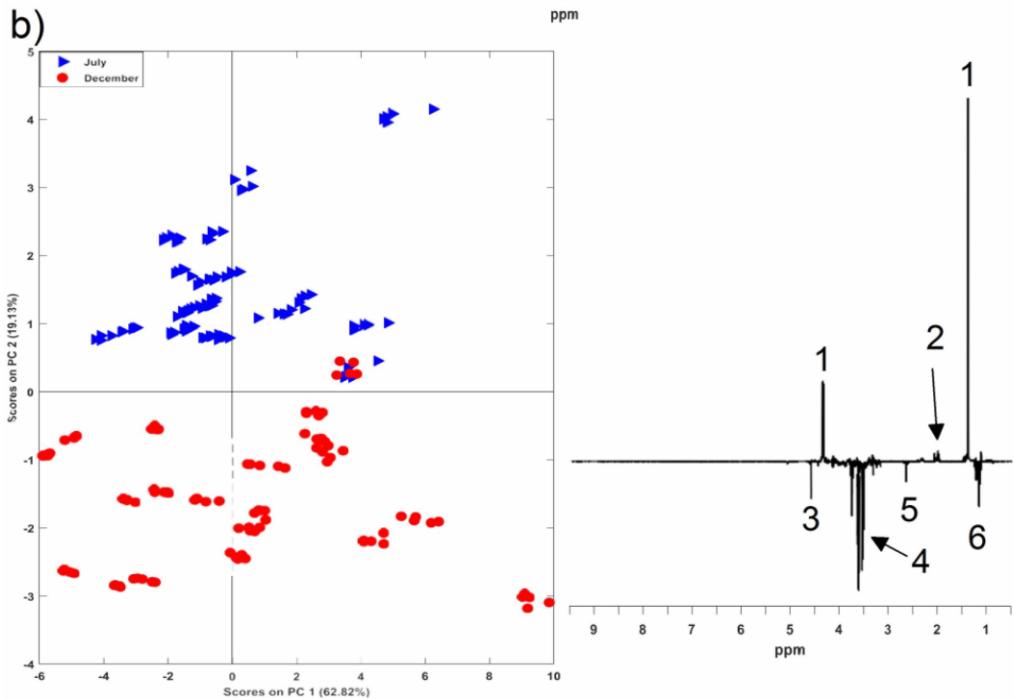
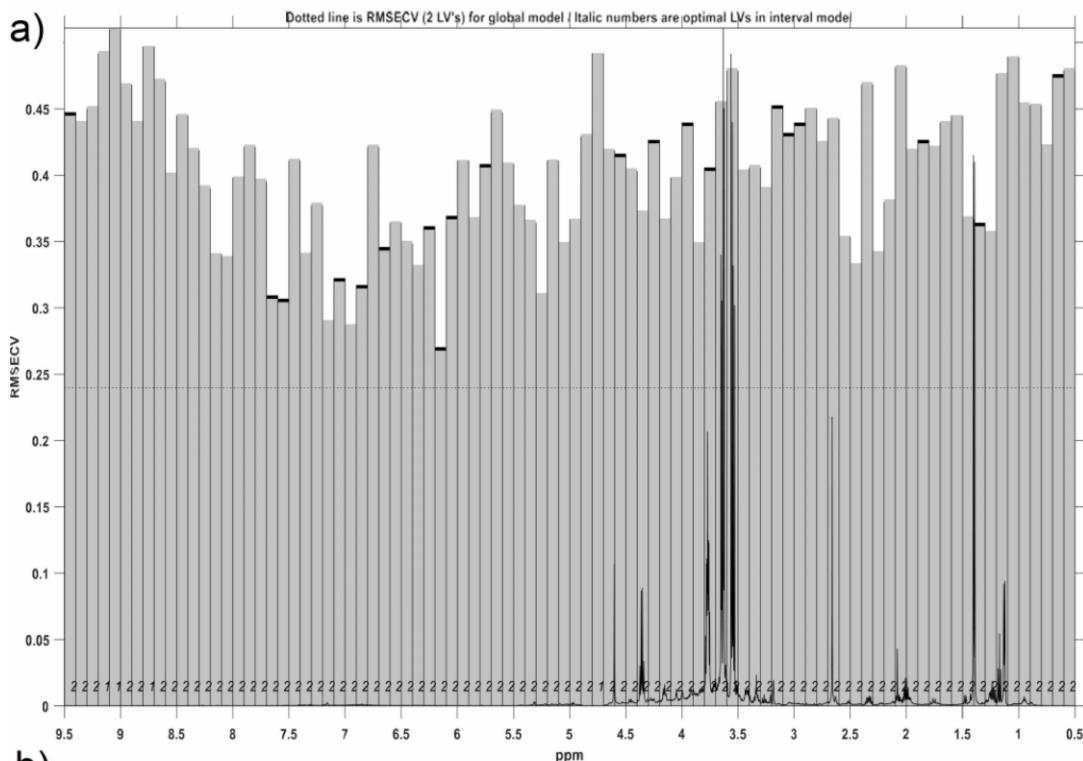
Processamento: prébiotico + térmico UHT e HTST + US+ Plasma

Análise: RMN ¹H + LC-MS

Quimiometria: PCA e quantificação (*q*NMR)









Cite this: *Food Funct.*, 2019, **10**, 1671

Cashew apple fiber prevents high fat diet-induced obesity in mice: an NMR metabolomic evaluation†

Diana Valesca Carvalho,^a Lorena Mara Alexandre Silva,^a Elenilson Godoy Alves Filho,^b Flávia Almeida Santos,^a Renan Pereira de Lima,^a Ana Flávia Seraine Custódio Viana,^a Paulo Iury Gomes Nunes,^b Said Gonçalves da Cruz Fonseca,^a Tiago Sousa de Melo,^c Daniel de Araújo Viana,^d Maria Izabel Gallão^a and Edy Sousa de Brito ^{a*}^b

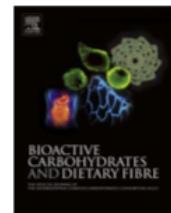


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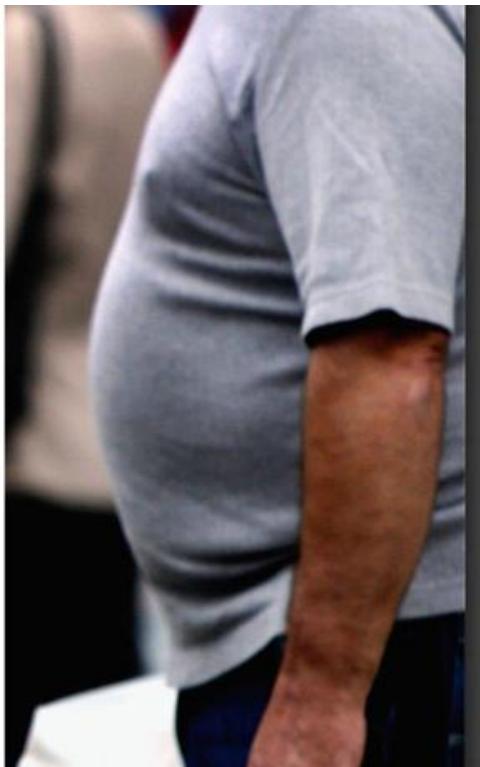
Influence of low molecular weight compounds associated to cashew (*Anacardium occidentale* L.) fiber on lipid metabolism, glycemia and insulinemia of normal mice

Diana Valesca Carvalho^a, Flávia Almeida Santos^b, Renan Pereira de Lima^c, Ana Flávia Seraine Custódio Viana^b, Said Gonçalves Cruz Fonseca^d, Paulo Iury Gomes Nunes^b, Tiago Sousa de Melo^e, Maria Izabel Gallão^f, Edy Sousa de Brito^{g,*}



Brasil

53,8%

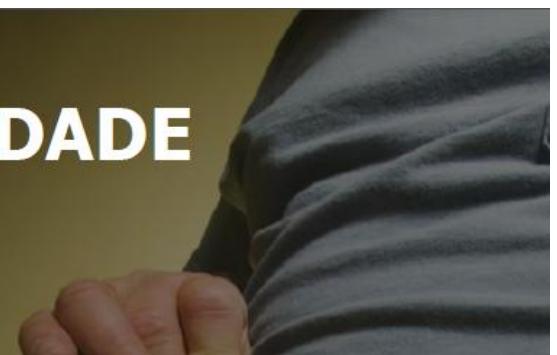


EXCESSO DE PESO

* IMC $\geq 25\text{kg/m}^2$

18,9%

OBESIDADE



* IMC $\geq 30\text{kg/m}^2$

(Vigitel, 2017)

Bagaço de frutas – fonte alternativa de fibra alimentar

- » Subproduto produção de sucos
- » Geralmente descartado no meio ambiente
- » Fibra e compostos fenólicos ligados à matriz da fibra
- » Caju: Importância sócio-econômica/riqueza em nutrientes



Fonte: Siqueira, de Brito. Foto: Cláudio de Norões Rocha

Preparação da ração dos camundongos

Metabolismo normal



FcI (10%)



Dieta Normal (ND)
– Ração comercial



ND-FcI



FcSM (10%)



Dieta Normal (ND)
– Ração comercial



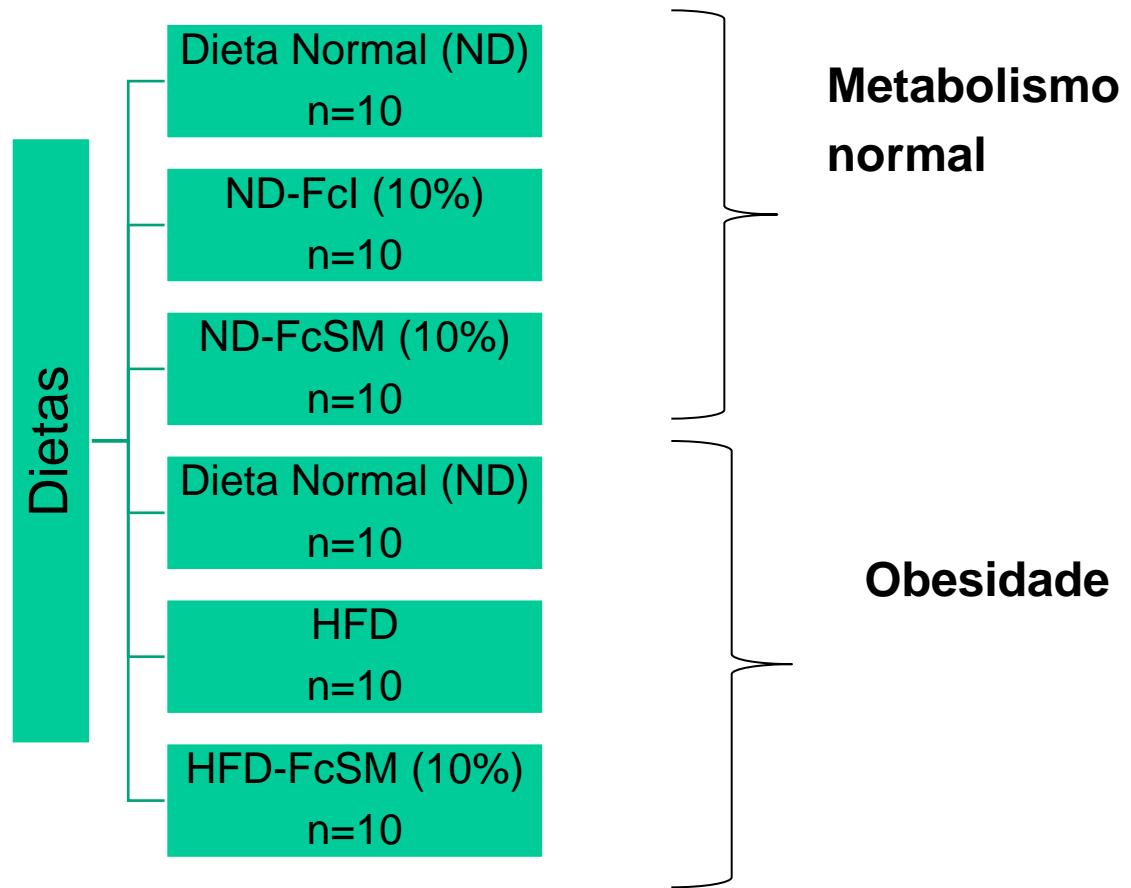
ND-FcSM





(p:19-25g)

**(CEPA/UFC
21/15)**



Alimentados durante 15 semanas; ração e água ad libidum

Peso animal

Consumo de ração e água 2x/semana

Coleta de amostras

14^a semana



Fezes

Obesidade

15^a
semana

Jejum de 6h



Sangue

Metabolismo
normal
e
Obesidade

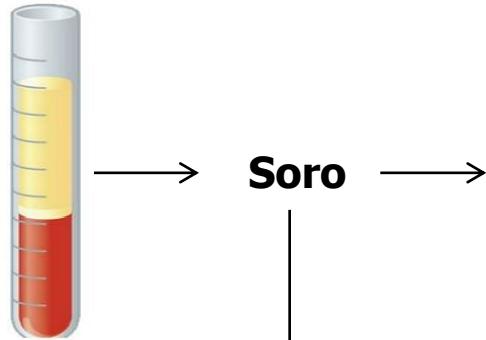
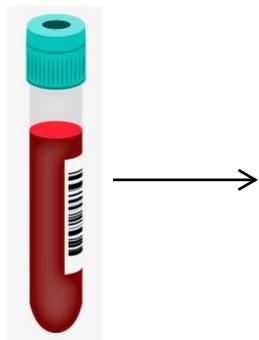
Eutanásia



Fígado



Tecido adiposo
branco
abdominal



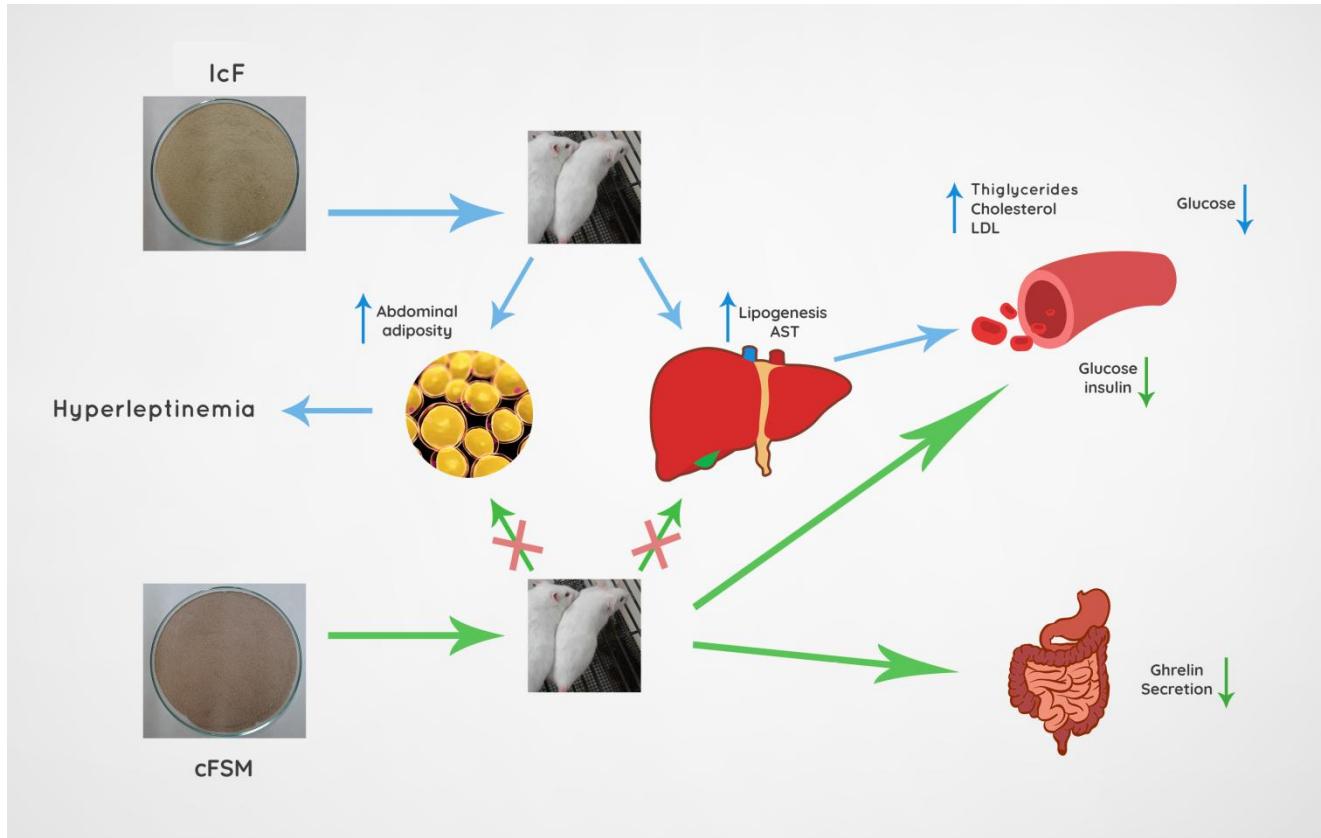
- Glicose
- Colesterol total, HDL, LDL, triglicerídeos
- Enzimas hepáticas (AST, ALT)
- Enzimas digestivas (amilase, lipase)
- Hormônios (Insulina, leptina, grelina)
- Citocinas (IL-6 e TNF- α , adiponectina)



Peso

- Peso
- MDA
- NP-SH
- Colesterol
- Histologia

Metabolismo normal

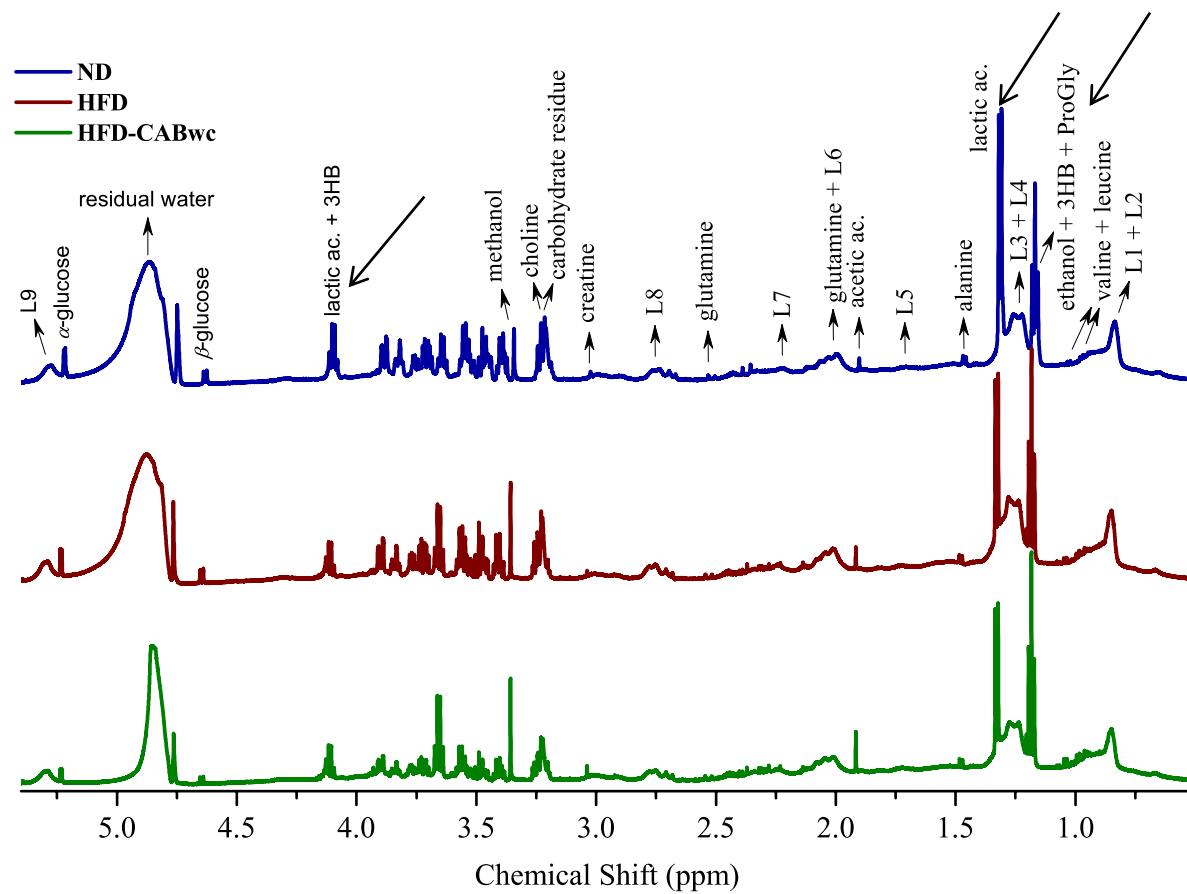


Ingestão alimentar e energética: sem diferença

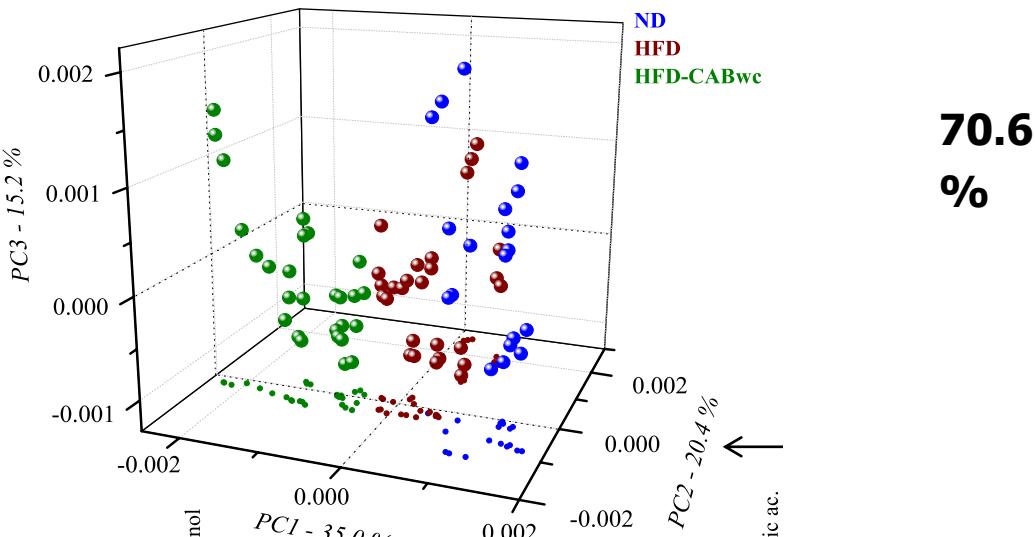
Ingestão de água: elevada FcSM

Peso corporal: sem diferença

Perfil do soro dos camundongos através de RMN acoplado a quimiometria

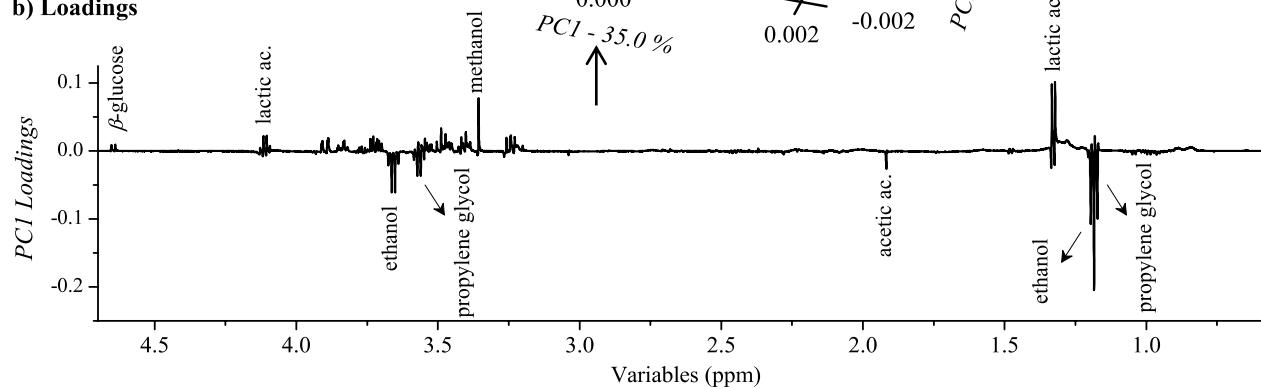


a) Scores



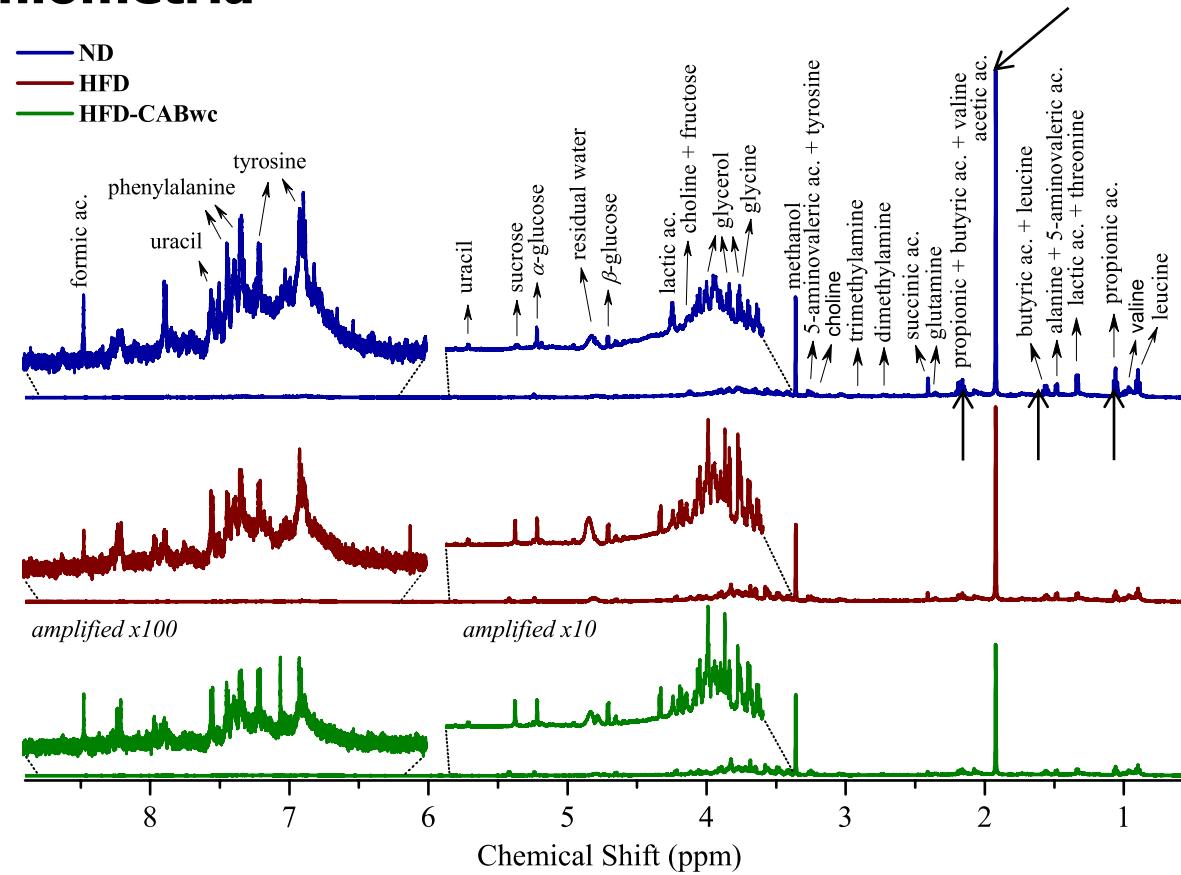
70.6
%

b) Loadings



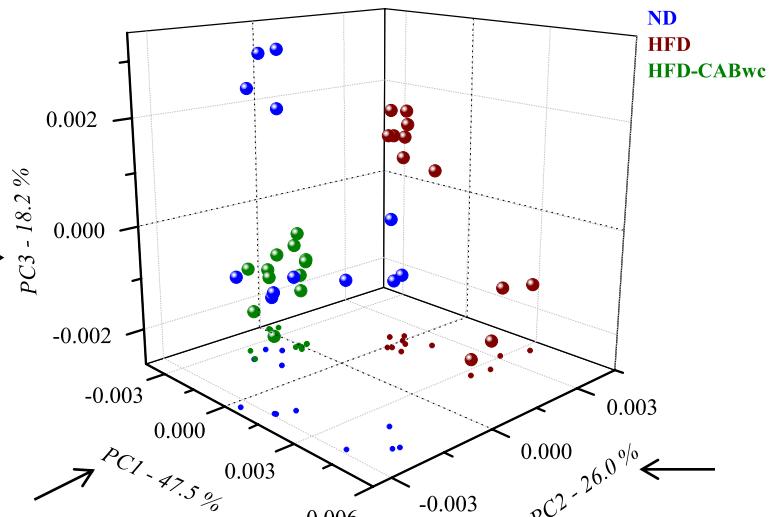
» etanol pode ser produzido por microrganismos intestinais sob diferentes condições nutricionais (Elshaghabee et. al, 2016).

Perfil das fezes dos camundongos através de RMN acoplado a quimiometria



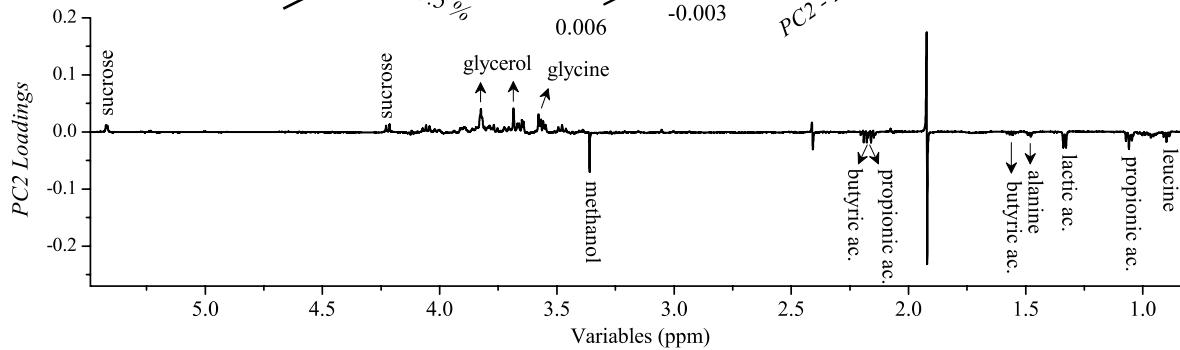
A comparação entre os espectros mostrou que as fezes são principalmente compostos de ácidos orgânicos de cadeia curta (acético, propiônico, butírico, láctico, succínico e fórmico), aminoácidos (leucina, alanina, uracila, tirosina e fenilalanina), açúcares (sacarose, glicose e frutose) e metanol.

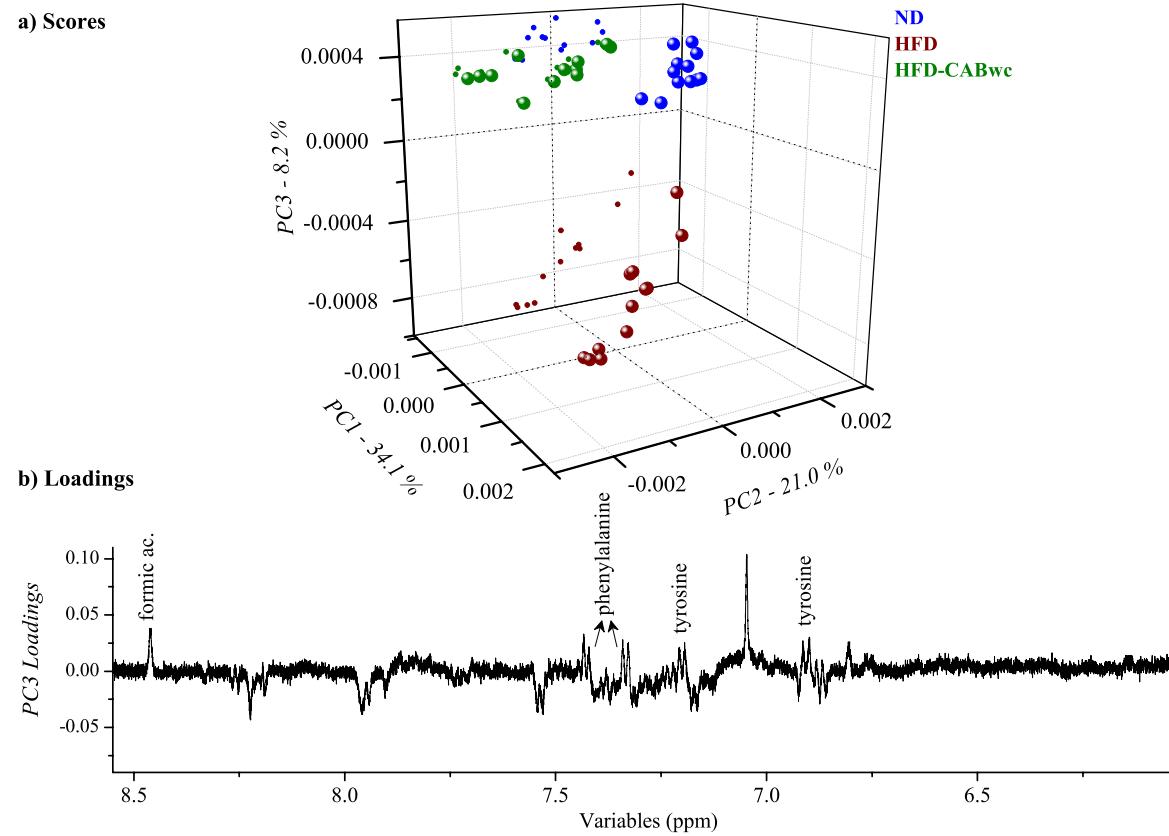
a) Scores



91.7%

b) Loadings

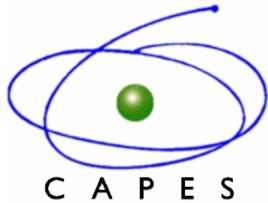




A presença desses aminoácidos está possivelmente relacionada às atividades metabólicas das bactérias intestinais do gênero *Adlercreutzia*, *Anaerostipes*, *Coprococcus* da família Lachnospiraceae (Lin et. al, 2016).



Agradecimentos



C A P E S



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E ABASTECIMENTO





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